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(54) Title: NOVEL DNA CLONING METHOD

(57) Abstract

The invention refers to a novel method for cloning DNA molecules using a homologous recombination mechanism between at least two DNA molecules comprising: a) providing a host cell capable of performing homologous recombination, b) contacting in said host cell a first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred.

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Novel DNA cloning method

Description

The invention refers to a novel method for cloning DNA molecules using a homologous recombination mechanism between at least two DNA molecules. Further, novel reagent kits suitable for DNA cloning are provided.

Current methods for cloning foreign DNA in bacterial cells usually comprise the steps of providing a suitable bacterial vector, cleaving said vector with a restriction enzyme and in vitro-inserting a foreign DNA fragment in said vector. The resulting recombinant vectors are then used to transform bacteria. Although such cloning methods have been used successfully for about 20 years they suffer from several drawbacks. These drawbacks are, in particular, that the in vitro steps required for inserting foreign DNA in a vector are often very complicated and time-consuming, if no suitable restriction sites are available on the foreign DNA or the vector.

Furthermore, current methods usually rely on the presence of suitable restriction enzyme cleavage sites in the vector into which the foreign DNA fragment is placed. This imposes two limitations on the final cloning product. First, the foreign DNA fragment can usually only be inserted into the vector at the position of such a restriction site or sites. Thus, the cloning product is limited by the disposition of suitable restriction sites and cloning into regions of the vector where there is no suitable restriction site, is difficult and often imprecise. Second, since restriction sites are typically 4 to 8 base pairs in length, they occur a multiple number of times as the size of the DNA molecules being used increases. This represents a practical limitation to the size of the DNA molecules that can be manipulated by most current cloning techniques. In particular, the larger sizes of DNA cloned into vectors such as cosmids, BACs, PACs and P1s are such that it is usually impractical to manipulate them directly by restriction enzyme based

techniques. Therefore, there is a need for providing a new cloning method, from which the drawbacks of the prior art have at least partly been eliminated.

According to the present invention it was found that an efficient homologous recombination mechanism between two DNA molecules occurs at usable frequencies in a bacterial host cell which is capable of expressing the products of the recE and recT genes or functionally related genes such as the reda and redß genes, or the phage P22 recombination system (Kolodner et al., Mol.Microbiol. 11 (1994) 23-30; Fenton, A.C. and Poteete, A.R., Virology 134 (1984) 148-160; Poteete, A.R. and Fenton, A.C., Virology 134 (1984) 161-167). This novel method of cloning DNA fragments is termed "ET cloning".

The identification and characterization of the E.coli RecE and RecT proteins is described Gillen et al. (J.Bacteriol. 145 (1981), 521-532) and Hall et al. (J.Bacteriol. 175 (1993), 277-287). Hall and Kolodner (Proc.Natl.Acad.Sci. USA 91 (1994), 3205-3209) disclose in vitro homologous pairing and strand exchange of linear double-stranded DNA and homologous circular single-stranded DNA promoted by the RecT protein. Any references to the use of this method for the cloning of DNA molecules in cells cannot be found therein.

The recET pathway of genetic recombination in E.coli is known (Hall and Kolodner (1994), supra; Gillen et al. (1981), supra). This pathway requires the expression of two genes, recE and recT. The DNA sequence of these genes has been published (Hall et al., supra). The RecE protein is similar to bacteriophage proteins, such as λ exo or λ Reda (Gillen et al., J.Mol.Biol.113 (1977), 27-41; Little, J.Biol.Chem. 242 (1967), 679-686; Radding and Carter, J.Biol.Chem. 246 (1971), 2513-2518; Joseph and Kolodner, J.Biol.Chem. 258 (1983), 10418-10424). The RecT protein is similar to bacteriophage proteins, such as λ ß-protein or λ Redß (Hall et al.

(1993), supra; Muniyappa and Radding, J.Biol.Chem. 261 (1986), 7472-7478; Kmiec and Hollomon, J.Biol.Chem.256 (1981), 12636-12639). The content of the above-cited documents is incorporated herein by reference.

Oliner et al. (Nucl.Acids Res. 21 (1993), 5192-5197) describe in vivo cloning of PCR products in E.coli by intermolecular homologous recombination between a linear PCR product and a linearized plasmid vector. Other previous attempts to develop new cloning methods based on homologous recombination in prokaryotes, too, relied on the use of restriction enzymes to linearise the vector (Bubeck et al., Nucleic Acids Res. 21 (1993), 3601-3602; Oliner et al., Nucleic Acids Res. 21 (1993), 5192-5197; Degryse, Gene 170 (1996), 45-50) or on the host-specific recA-dependent recombination system (Hamilton et al., J.Bacteriol. 171 (1989), 4617-4622; Yang et al., Nature Biotech. 15 (1997), 859-865; Dabert and Smith, Genetics 145 (1997), 877-889). These methods are of very limited applicability and are hardly used in practice.

The novel method of cloning DNA according to the present invention does not require in vitro treatments with restriction enzymes or DNA ligases and is therefore fundamentally distinct from the standard methodologies of DNA cloning. The method relies on a pathway of homologous recombination in E.coli involving the recE and recT gene products, or the redα and redß gene products, or functionally equivalent gene products. The method covalently combines one preferably linear and preferably extrachromosomal DNA fragment, the DNA fragment to be cloned, with one second preferably circular DNA vector molecule, either an episome or the endogenous host chromosome or chromosomes. It is therefore distinct from previous descriptions of cloning in E.coli by homologous recombination which either rely on the use of two linear DNA fragments or different recombination pathways.

The present invention provides a flexible way to use homologous recombination to engineer large DNA molecules including an intact > 76 kb plasmid and the E.coli chromosome. Thus, there is practically no limitation of target choice either according to size or site. Therefore, any recipient DNA in a host cell, from high copy plasmid to the genome, is amenable to precise alteration. In addition to engineering large DNA molecules, the invention outlines new, restriction enzyme-independent approaches to DNA design. For example, deletions between any two chosen base pairs in a target episome can be made by choice of oligonucleotide homology arms. Similarly, chosen DNA sequences can be inserted at a chosen base pair to create, for example, altered protein reading frames. Concerted combinations of insertions and deletions, as well as point mutations, are also possible. The application of these strategies is particularly relevant to complex or difficult DNA constructions, for example, those intended for homologous recombinations in eukaryotic cells, e.g. mouse embryonic stem cells. Further, the present invention provides a simple way to position site specific recombination target sites exactly where desired. This will simplify applications of site specific recombination in other living systems, such as plants and mice.

A subject matter of the present invention is a method for cloning DNA molecules in cells comprising the steps:

a) providing a host cell capable of performing homologous recombination,

b) contacting in said host cell a first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and

c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred.

In the method of the present invention the homologous recombination preferably occurs via the recET mechanism, i.e. the homologous recombination is mediated by the gene products of the recE and the recT genes which are preferably selected from the E.coli genes recE and recT or functionally related genes such as the phage λ redα and redß genes.

The host cell suitable for the method of the present invention preferably is a bacterial cell, e.g. a gram-negative bacterial cell. More preferably, the host cell is an enterobacterial cell, such as Salmonella, Klebsiella or Escherichia. Most preferably the host cell is an Escherichia coli cell. It should be noted, however, that the cloning method of the present invention is also suitable for eukaryotic cells, such as fungi, plant or animal cells.

Preferably, the host cell used for homologous recombination and propagation of the cloned DNA can be any cell, e.g. a bacterial strain in which the products of the recE and recT, or redα and redß, genes are expressed. The host cell may comprise the recE and recT genes located on the host cell chromosome or on non-chromosomal DNA, preferably on a vector, e.g. a plasmid. In a preferred case, the RecE and RecT, or Redα and Redß, gene products are expressed from two different regulatable promoters, such as the arabinose-inducible BAD promoter or the lac promoter or from non-regulatable promoters. Alternatively, the recE and recT, or redα and redß, genes are expressed on a polycistronic mRNA from a single regulatable or non-regulatable promoter. Preferably the expression is controlled by regulatable promoters.

Especially preferred is also an embodiment, wherein the recE or redα gene is expressed by a regulatable promoter. Thus, the recombinogenic potential of the system is only elicited when required and, at other times, possible undesired recombination reactions are limited. The recT or redß gene, on the other hand, is preferably overexpressed with respect to recE or redα. This may be accomplished by using a strong constitutive promoter, e.g. the

EM7 promoter and/or by using a higher copy number of recT, or redß, versus recE, or redα, genes.

For the purpose of the present invention any recE and recT genes are suitable insofar as they allow a homologous recombination of first and second DNA molecules with sufficient efficiency to give rise to recombination products in more than 1 in 10^9 cells transfected with DNA. The recE and recT genes may be derived from any bacterial strain or from bacteriophages or may be mutants and variants thereof. Preferred are recE and recT genes which are derived from E.coli or from E.coli bacteriophages, such as the redα and redß genes from lambdoid phages, e.g. bacteriophage λ.

More preferably, the recE or redα gene is selected from a nucleic acid molecule comprising

(a) the nucleic acid sequence from position 1320 (ATG) to 2159 (GAC) as depicted in Fig.7B,

(b) the nucleic acid sequence from position 1320 (ATG) to 1998(CGA) as depicted in Fig.14B,

(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or

(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequence from (a), (b) and/or (c).

More preferably, the recT or redß gene is selected from a nucleic acid molecule comprising

(a) the nucleic acid sequence from position 2155 (ATG) to 2961 (GAA) as depicted in Fig.7B,

(b) the nucleic acid sequence from position 2086 (ATG) to 2868 (GCA) as depicted in Fig.14B,

(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or

(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequences from (a), (b) and/or (c).

It should be noted that the present invention also encompasses mutants and variants of the given sequences, e.g. naturally occurring mutants and variants or mutants and variants obtained by genetic engineering. Further it should be noted that the recE gene depicted in Fig.7B is an already truncated gene encoding amino acids 588-866 of the native protein. Mutants and variants preferably have a nucleotide sequence identity of at least 60%, preferably of at least 70% and more preferably of at least 80% of the recE and recT sequences depicted in Fig.7B and 13B, and of the redα and redß sequences depicted in Fig.14B.

According to the present invention hybridization under stringent conditions preferably is defined according to Sambrook et al. (1989), infra, and comprises a detectable hybridization signal after washing for 30 min in 0.1 x SSC, 0.5% SDS at 55°C, preferably at 62°C and more preferably at 68°C.

In a preferred case the recE and recT genes are derived from the corresponding endogenous genes present in the E.coli K12 strain and its derivatives or from bacteriophages. In particular, strains that carry the sbcA mutation are suitable. Examples of such strains are JC8679 and JC 9604 (Gillen et al. (1981), supra). Alternatively, the corresponding genes may also be obtained from other coliphages such as lambdoid phages or phage P22.

The genotype of JC 8679 and JC 9604 is Sex (Hfr, F+, F-, or F') : F-.JC 8679 comprises the mutations: recBC 21, recC 22, sbcA 23, thr-1, ara-14, leu B 6, DE (gpt-proA) 62, lacY1, tsx-33, gluV44 (AS), galK2 (Oc), LAM-, his-60, relA 1, rps L31 (strR), xyl A5, mtl-1, argE3 (Oc) and thi-1. JC 9604 comprises the same mutations and further the mutation recA 56.

Further, it should be noted that the recE and recT, or redα and redß, genes can be isolated from a first donor source, e.g. a donor bacterial cell and transformed into a second receptor source, e.g. a receptor bacterial or eukaryotic cell in which they are expressed by recombinant DNA means.

In one embodiment of the invention, the host cell used is a bacterial strain having an sbcA mutation, e.g. one of E.coli strains JC 8679 and JC 9604 mentioned above. However, the method of the invention is not limited to host cells having an sbcA mutation or analogous cells. Surprisingly, it has been found that the cloning method of the invention also works in cells without sbcA mutation, whether recBC + or recBC-, e.g. also in prokaryotic recBC + host cells, e.g. in E.coli recBC + cells. In that case preferably those host cells are used in which the product of a recBC type exonuclease inhibitor gene is expressed. Preferably, the exonuclease inhibitor is capable of inhibiting the host recBC system or an equivalent thereof. A suitable example of such exonuclease inhibitor gene is the λ redγ gene (Murphy, J.Bacteriol. 173 (1991), 5808-5821) and functional equivalents thereof, respectively, which, for example, can be obtained from other coliphages such as from phage P22 (Murphy, J.Biol.Chem.269 (1994), 22507-22516).

More preferably, the exonuclease inhibitor gene is selected from a nucleic acid molecule comprising

(a) the nucleic acid sequence from position 3588 (ATG) to 4002 (GTA) as depicted in Fig.14A,

(b) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or

(c) a nucleic acid sequence which hybridizes under stringent conditions (as defined above) with the nucleic acid sequence from (a) and/ or (b).

Surprisingly, it has been found that the expression of an exonuclease inhibitor gene in both recBC + and recBC- strains leads to significant improvement of cloning efficiency.

The cloning method according to the present invention employs a homologous recombination between a first DNA molecule and a second DNA molecule. The first DNA molecule can be any DNA molecule that carries an origin of replication which is operative in the host cell, e.g. an E.coli replication origin. Further, the first DNA molecule is present in a form which is capable of being replicated in the host cell. The first DNA molecule, i.e. the vector, can be any extrachromosomal DNA molecule containing an origin of replication which is operative in said host cell, e.g. a plasmid including single, low, medium or high copy plasmids or other extrachromosomal circular DNA molecules based on cosmid, P1, BAC or PAC vector technology. Examples of such vectors are described, for example, by Sambrook et al. (Molecular Cloning, Laboratory Manual, 2nd Edition (1989), Cold Spring Harbor Laboratory Press) and Ioannou et al. (Nature Genet. 6 (1994), 84-89) or references cited therein. The first DNA molecule can also be a host cell chromosome, particularly the E.coli chromosome. Preferably, the first DNA molecule is a double-stranded DNA molecule.

The second DNA molecule is preferably a linear DNA molecule and comprises at least two regions of sequence homology, preferably of sequence identity to regions on the first DNA molecule. These homology or identity regions are preferably at least 15 nucleotides each, more preferably at least 20 nucleotides and, most preferably, at least 30 nucleotides each. Especially good results were obtained when using sequence homology regions having a length of about 40 or more nucleotides, e.g. 60 or more nucleotides. The two sequence homology regions can be located on the linear DNA fragment so that one is at one end and the other is at the other end, however they may also be located internally. Preferably, also the second DNA molecule is a double-stranded DNA molecule.

The two sequence homology regions are chosen according to the experimental design. There are no limitations on which regions of the first

DNA molecule can be chosen for the two sequence homology regions located on the second DNA molecule, except that the homologous recombination event cannot delete the origin of replication of the first DNA molecule. The sequence homology regions can be interrupted by non-identical sequence regions as long as sufficient sequence homology is retained for the homologous recombination reaction. By using sequence homology arms having non-identical sequence regions compared to the target site mutations such as substitutions, e.g. point mutations, insertions and/or deletions may be introduced into the target site by ET cloning.

The second foreign DNA molecule which is to be cloned in the bacterial cell may be derived from any source. For example, the second DNA molecule may be synthesized by a nucleic acid amplification reaction such as a PCR where both of the DNA oligonucleotides used to prime the amplification contain in addition to sequences at the 3'-ends that serve as a primer for the amplification, one or the other of the two homology regions. Using oligonucleotides of this design, the DNA product of the amplification can be any DNA sequence suitable for amplification and will additionally have a sequence homology region at each end.

A specific example of the generation of the second DNA molecule is the amplification of a gene that serves to convey a phenotypic difference to the bacterial host cells, in particular, antibiotic resistance. A simple variation of this procedure involves the use of oligonucleotides that include other sequences in addition to the PCR primer sequence and the sequence homology region. A further simple variation is the use of more than two amplification primers to generate the amplification product. A further simple variation is the use of more than one amplification reaction to generate the amplification product. A further variation is the use of DNA fragments obtained by methods other than PCR, for example, by endonuclease or restriction enzyme cleavage to linearize fragments from any source of DNA.

It should be noted that the second DNA molecule is not necessarily a single species of DNA molecule. It is of course possible to use a heterogenous population of second DNA molecules, e.g. to generate a DNA library, such as a genomic or cDNA library.

The method of the present invention may comprise the contacting of the first and second DNA molecules in vivo. In one embodiment of the present invention the second DNA fragment is transformed into a bacterial strain that already harbors the first vector DNA molecule. In a different embodiment, the second DNA molecule and the first DNA molecule are mixed together in vitro before co-transformation in the bacterial host cell. These two embodiments of the present invention are schematically depicted in Fig.1. The method of transformation can be any method known in the art (e.g. Sambrook et al. supra). The preferred method of transformation or co-transformation, however, is electroporation.

After contacting the first and second DNA molecules under conditions which favour homologous recombination between first and second DNA molecules via the ET cloning mechanism a host cell is selected, in which homologous recombination between said first and second DNA molecules has occurred. This selection procedure can be carried out by several different methods. In the following three preferred selection methods are depicted in Fig.2 and described in detail below.

In a first selection method a second DNA fragment is employed which carries a gene for a marker placed between the two regions of sequence homology wherein homologous recombination is detectable by expression of the marker gene. The marker gene may be a gene for a phenotypic marker which is not expressed in the host or from the first DNA molecule. Upon recombination by ET cloning, the change in phenotype of the host strain conveyed by the stable acquisition of the second DNA fragment identifies the ET cloning product.

In a preferred case, the phenotypic marker is a gene that conveys resistance to an antibiotic, in particular, genes that convey resistance to kanamycin, ampillicin, chloramphenicol, tetracyclin or any other substance that shows bacteriocidal or bacteriostatic effects on the bacterial strain employed.

A simple variation is the use of a gene that complements a deficiency present within the bacterial host strain employed. For example, the host strain may be mutated so that it is incapable of growth without a metabolic supplement. In the absence of this supplement, a gene on the second DNA fragment can complement the mutational defect thus permitting growth. Only those cells which contain the episome carrying the intended DNA rearrangement caused by the ET cloning step will grow.

In another example, the host strain carries a phenotypic marker gene which is mutated so that one of its codons is a stop codon that truncates the open reading frame. Expression of the full length protein from this phenotypic marker gene requires the introduction of a suppressor tRNA gene which, once expressed, recognizes the stop codon and permits translation of the full open reading frame. The suppressor tRNA gene is introduced by the ET cloning step and successful recombinants identified by selection for, or identification of, the expression of the phenotypic marker gene. In these cases, only those cells which contain the intended DNA rearrangement caused by the ET cloning step will grow.

A further simple variation is the use of a reporter gene that conveys a readily detectable change in colony colour or morphology. In a preferred case, the green fluorescence protein (GFP) can be used and colonies carrying the ET cloning product identified by the fluorescence emissions of GFP. In another preferred case, the lacZ gene can be used and colonies carrying the ET cloning product identified by a blue colony colour when X-gal is added to the culture medium.

In a second selection method the insertion of the second DNA fragment into the first DNA molecule by ET cloning alters the expression of a marker present on the first DNA molecule. In this embodiment the first DNA molecule contains at least one marker gene between the two regions of sequence homology and homologous recombination may be detected by an altered expression, e.g. lack of expression of the marker gene.

In a preferred application, the marker present on the first DNA molecule is a counter-selectable gene product, such as the sacB, ccdB or tetracycline-resistance genes. In these cases, bacterial cells that carry the first DNA molecule unmodified by the ET cloning step after transformation with the second DNA fragment, or co-transformation with the second DNA fragment and the first DNA molecule, are plated onto a medium so the expression of the counter-selectable marker conveys a toxic or bacteriostatic effect on the host. Only those bacterial cells which contain the first DNA molecule carrying the intended DNA rearrangement caused by the ET cloning step will grow.

In another preferred application, the first DNA molecule carries a reporter gene that conveys a readily detectable change in colony colour or morphology. In a preferred case, the green fluorescence protein (GFP) can be present on the first DNA molecule and colonies carrying the first DNA molecule with or without the ET cloning product can be distinguished by differences in the fluorescence emissions of GFP. In another preferred case, the lacZ gene can be present on the first DNA molecule and colonies carrying the first DNA molecule with or without the ET cloning product identified by a blue or white colony colour when X-gal is added to the culture medium.

In a third selection method the integration of the second DNA fragment into the first DNA molecule by ET cloning removes a target site for a site specific recombinase, termed here an RT (for recombinase target) present

on the first DNA molecule between the two regions of sequence homology. A homologous recombination event may be detected by removal of the target site.

In the absence of the ET cloning product, the RT is available for use by the corresponding site specific recombinase. The difference between the presence or not of this RT is the basis for selection of the ET cloning product. In the presence of this RT and the corresponding site specific recombinase, the site specific recombinase mediates recombination at this RT and changes the phenotype of the host so that it is either not able to grow or presents a readily observable phenotype. In the absence of this RT, the corresponding site specific recombinase is not able to mediate recombination.

In a preferred case, the first DNA molecule to which the second DNA fragment is directed, contains two RTs, one of which is adjacent to, but not part of, an antibiotic resistance gene. The second DNA fragment is directed, by design, to remove this RT. Upon exposure to the corresponding site specific recombinase, those first DNA molecules that do not carry the ET cloning product will be subject to a site specific recombination reaction between the RTs that remove the antibiotic resistance gene and therefore the first DNA molecule fails to convey resistance to the corresponding antibiotic. Only those first DNA molecules that contain the ET cloning product, or have failed to be site specifically recombined for some other reason, will convey resistance to the antibiotic.

In another preferred case, the RT to be removed by ET cloning of the second DNA fragment is adjacent to a gene that complements a deficiency present within the host strain employed. In another preferred case, the RT to be removed by ET cloning of the second DNA fragment is adjacent to a reporter gene that conveys a readily detectable change in colony colour or morphology.

In another preferred case, the RT to be removed by ET cloning of the second DNA fragment is anywhere on a first episomal DNA molecule and the episome carries an origin of replication incompatible with survival of the bacterial host cell if it is integrated into the host genome. In this case the host genome carries a second RT, which may or may not be a mutated RT so that the corresponding site specific recombinase can integrate the episome, via its RT, into the RT sited in the host genome. Other preferred RTs include RTs for site specific recombinases of the resolvase/transposase class. RTs include those described from existing examples of site specific recombination as well as natural or mutated variations thereof.

The preferred site specific recombinases include Cre, FLP, Kw or any site specific recombinase of the integrase class. Other preferred site specific recombinases include site specific recombinases of the resolvase/transposase class.

There are no limitations on the method of expression of the site specific recombinase in the host cell. In a preferred method, the expression of the site specific recombinase is regulated so that expression can be induced and quenched according to the optimisation of the ET cloning efficiency. In this case, the site specific recombinase gene can be either integrated into the host genome or carried on an episome. In another preferred case, the site specific recombinase is expressed from an episome that carries a conditional origin of replication so that it can be eliminated from the host cell.

In another preferred case, at least two of the above three selection methods are combined. A particularly preferred case involves a two-step use of the first selection method above, followed by use of the second selection method. This combined use requires, most simply, that the DNA fragment to be cloned includes a gene, or genes that permits the identification, in the first step, of correct ET cloning products by the acquisition of a phenotypic

change. In a second step, expression of the gene or genes introduced in the first step is altered so that a second round of ET cloning products can be identified. In a preferred example, the gene employed is the tetracycline resistance gene and the first step ET cloning products are identified by the acquisition of tetracycline resistance. In the second step, loss of expression of the tetracycline gene is identified by loss of sensitivity to nickel chloride, fusaric acid or any other agent that is toxic to the host cell when the tetracycline gene is expressed. This two-step procedure permits the identification of ET cloning products by first the integration of a gene that conveys a phenotypic change on the host, and second by the loss of a related phenotypic change, most simply by removal of some of the DNA sequences integrated in the first step. Thereby the genes used to identify ET cloning products can be inserted and then removed to leave ET cloning products that are free of these genes.

In a further embodiment of the present invention the ET cloning may also be used for a recombination method comprising the steps of

a) providing a source of RecE and RecT, or Redα and Redß, proteins,

b) contacting a first DNA molecule which is capable of being replicated in a suitable host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and

c) selecting DNA molecules in which a homologous recombination between said first and second DNA molecules has occurred.

The source of RecE and RecT, or Redα and Redß, proteins may be either purified or partially purified RecE and RecT, or Redα and Redß, proteins or cell extracts comprising RecE and RecT, or Redα and Redß, proteins.

The homologous recombination event in this embodiment may occur in vitro, e.g. when providing a cell extract containing further components

required for homologous recombination. The homologous recombination event, however, may also occur in vivo, e.g. by introducing RecE and RecT, or Reda and Redß, proteins or the extract in a host cell (which may be recET positive or not, or redaß positive or not) and contacting the DNA molecules in the host cell. When the recombination occurs in vitro the selection of DNA molecules may be accomplished by transforming the recombination mixture in a suitable host cell and selecting for positive clones as described above. When the recombination occurs in vivo the selection methods as described above may directly be applied.

A further subject matter of the invention is the use of cells, preferably bacterial cells, most preferably, E.coli cells capable of expressing the recE and recT, or reda and redß, genes as a host cell for a cloning method involving homologous recombination.

Still a further subject matter of the invention is a vector system capable of expressing recE and recT, or reda and redß, genes in a host cell and its use for a cloning method involving homologous recombination. Preferably, the vector system is also capable of expressing an exonuclease inhibitor gene as defined above, e.g. the λ redγ gene. The vector system may comprise at least one vector. The recE and recT, or reda and redß, genes are preferably located on a single vector and more preferably under control of a regulatable promoter which may be the same for both genes or a single promoter for each gene. Especially preferred is a vector system which is capable of overexpressing the recT, or redß, gene versus the recE, or reda, gene.

Still a further subject matter of the invention is the use of a source of RecE and RecT, or Reda and Redß, proteins for a cloning method involving homologous recombination.

A still further subject matter of the invention is a reagent kit for cloning comprising

(a) a host cell, preferably a bacterial host cell,

(b) means of expressing recE and recT, or redα and redß, genes in said host cell, e.g. comprising a vector system, and

(c) a recipient cloning vehicle, e.g. a vector, capable of being replicated in said cell.

On the one hand, the recipient cloning vehicle which corresponds to the first DNA molecule of the process of the invention can already be present in the bacterial cell. On the other hand, it can be present separated from the bacterial cell.

In a further embodiment the reagent kit comprises

(a) a source for RecE and RecT, or Redα and Redß, proteins and

(b) a recipient cloning vehicle capable of being propagated in a host cell and

(c) optionally a host cell suitable for propagating said recipient cloning vehicle.

The reagent kit furthermore contains, preferably, means for expressing a site specific recombinase in said host cell, in particular, when the recipient ET cloning product contains at least one site specific recombinase target site. Moreover, the reagent kit can also contain DNA molecules suitable for use as a source of linear DNA fragments used for ET cloning, preferably by serving as templates for PCR generation of the linear fragment, also as specifically designed DNA vectors from which the linear DNA fragment is released by restriction enzyme cleavage, or as prepared linear fragments included in the kit for use as positive controls or other tasks. Moreover, the reagent kit can also contain nucleic acid amplification primers comprising a region of homology to said vector. Preferably, this region of homology is located at the 5'-end of the nucleic acid amplification primer.

The invention is further illustrated by the following Sequence listings, Figures and Examples.

SEQ ID NO. 1: shows the nucleic acid sequence of the plasmid pBAD24-rec ET (Fig. 7).

SEQ ID NOs 2/3: show the nucleic acid and amino acid sequences of the truncated recE gene (t-recE) present on pBAD24-recET at positions 1320-2162.

SEQ ID NOs 4/5: show the nucleic acid and amino acid sequences of the recT gene present on pBAD24-recET at position 2155-2972.

SEQ ID NOs 6/7: show the nucleic acid and amino acid sequences of the araC gene present on the complementary stand to the one shown of pBAD24-recET at positions 974-996.

SEQ ID NOs 8/9: show the nucleic acid an amino acid sequences of the bla gene present on pBAD24-recET at positions 3493-4353.

SEQ ID NO 10: shows the nucleic acid sequence of the plasmid pBAD-ETγ (Fig. 13).

SEQ ID No 11: shows the nucleic acid sequence of the plasmid pBAD-αßγ (Fig. 14) as well as the coding regions for the genes redα (1320-200), redß (2086-2871) and redγ (3403-3819).

SEQ ID NOs 12-14: show the amino acid sequences of the Redα, Redß and Redγ proteins, respectively. The redγ sequence is present on each of pBAD-ETγ (Fig. 13) and pBAD-αßγ (Fig. 14).

Figure 1

A preferred method for ET cloning is shown by diagram. The linear DNA fragment to be cloned is synthesized by PCR using oligonucleotide primers that contain a left homology arm chosen to match sequences in the recipient episome and a sequence for priming in the PCR reaction, and a

right homology arm chosen to match another sequence in the recipient episome and a sequence for priming in the PCR reaction. The product of the PCR reaction, here a selectable marker gene (sm1), is consequently flanked by the left and right homology arms and can be mixed together in vitro with the episome before co-transformation, or transformed into a host cell harboring the target episome. The host cell contains the products of the recE and recT genes. ET cloning products are identified by the combination of two selectable markers, sm1 and sm2 on the recipient episome.

Figure 2

Three ways to identify ET cloning products are depicted. The first, (on the left of the figure), shows the acquisition, by ET cloning, of a gene that conveys a phenotypic difference to the host, here a selectable marker gene (sm). The second (in the centre of the figure) shows the loss, by ET cloning, of a gene that conveys a phenotypic difference to the host, here a counter selectable marker gene (counter-sm). The third shows the loss of a target site (RT, shown as triangles on the circular episome) for a site specific recombinase (SSR), by ET cloning. In this case, the correct ET cloning product deletes one of the target sites required by the SSR to delete a selectable marker gene (sm). The failure of the SSR to delete the sm gene identifies the correct ET cloning product.

Figure 3

A simple example of ET cloning is presented.

(a) Top panel - PCR products (left lane) synthesized from oligonucleotides designed as described in Fig.1 to amplify by PCR a kanamycin resistance gene and to be flanked by homology arms present in the recipient vector, were mixed in vitro with the recipient vector (2nd lane) and cotransformed into a recET+ E.coli host. The recipient vector carried an ampillicin

resistance gene. (b) Transformation of the sbcA E.coli strain JC9604 with either the PCR product alone (0.2 μg) or the vector alone (0.3 μg) did not convey resistance to double selection with ampicillin and kanamycin (amp + kan), however cotransformation of both the PCR product and the vector produced double resistant colonies. More than 95% of these colonies contained the correct ET cloning product where the kanamycin gene had precisely integrated into the recipient vector according to the choice of homology arms. The two lanes on the right of (a) show Pvu II restriction enzyme digestion of the recipient vector before and after ET cloning. (c) As for b, except that six PCR products (0.2 μg each) were cotransformed with pSVpaZ11 (0.3 μg each) into JC9604 and plated onto Amp + Kan plates or Amp plates. Results are plotted as Amp + Kan-resistant colonies, representing recombination products, divided by Amp-resistant colonies, representing the plasmid transformation efficiency of the competent cell preparation, x 10^6. The PCR products were equivalent to the a-b PCR product except that homology arm lengths were varied. Results are from five experiments that used the same batches of competent cells and DNAs. Error bars represent standard deviation. (d) Eight products flanked by 50 bp homology arms were cotransformed with pSVpaZ11 into JC9604. All eight PCR products contained the same left homology arm and amplified neo gene. The right homology arms were chosen from the pSVpaZ11 sequence to be adjacent to (0), or at increasing distances (7-3100 bp), from the left. Results are from four experiments.

Figure 4

ET cloning in an approximately 100kb P1 vector to exchange the selectable marker.

A P1 clone which uses a kanamycin resistance gene as selectable marker and which contains at least 70kb of the mouse Hox a gene cluster was used. Before ET cloning, this episome conveys kanamycin resistance (top

panel, upper left) to its host E.coli which are ampillicin sensitive (top panel, upper right). A linear DNA fragment designed to replace the kanamycin resistance gene with an ampillicin resistance gene was made by PCR as outlined in Fig.1 and transformed into E.coli host cells in which the recipient Hox a/P1 vector was resident. ET cloning resulted in the deletion of the kanamycin resistance gene, and restoration of kanamycin sensitivity (top panel, lower left) and the acquisition of ampillicin resistance (top panel, lower right). Precise DNA recombination was verified by restriction digestion and Southern blotting analyses of isolated DNA before and after ET cloning (lower panel).

Figure 5

ET cloning to remove a counter selectable marker

A PCR fragment (upper panel, left, third lane) made as outlined in Figs.1 and 2 to contain the kanamycin resistance gene was directed by its chosen homology arms to delete the counter selectable ccdB gene present in the vector, pZero-2.1. The PCR product and the pZero vector were mixed in vitro (upper panel, left, 1st lane) before cotransformation into a recE/recT + E.coli host. Transformation of pZero-2.1 alone and plating onto kanamycin selection medium resulted in little colony growth (lower panel, left). Cotransformation of pZero-2.1 and the PCR product presented ET cloning products (lower panel, right) which showed the intended molecular event as visualized by Pvu II digestion (upper panel, right).

Figure 6

ET cloning mediated by inducible expression of recE and recT from an episome.

RecE/RecT mediate homologous recombination between linear and circular DNA molecules. (a) The plasmid pBAD24-recET was transformed into E.coli JC5547, and then batches of competent cells were prepared after induction of RecE/RecT expression by addition of L-arabinose for the times indicated

before harvesting. A PCR product, made using oligonucleotides e and f to contain the chloramphenicol resistance gene (cm) of pMAK705 and 50 bp homology arms chosen to flank the ampicillin resistance gene (bla) of pBAD24-recET, was then transformed and recombinants identified on chloramphenicol plates. (b) Arabinose was added to cultures of pBAD24-recET transformed JC5547 for different times immediately before harvesting for competent cell preparation. Total protein expression was analyzed by SDS-PAGE and Coomassie blue staining. (c) The number of chloramphenicol resistant colonies per μg of PCR product was normalized against a control for transformation efficiency, determined by including 5 pg pZero2.1, conveying kanamycin resistance, in the transformation and plating an aliquot onto Kan plates.

Figure 7A

The plasmid pBAD24-recET is shown by diagram. The plasmid contains the genes recE (in a truncated form) and recT under control of the inducible BAD promoter (P_{BAD}). The plasmid further contains an ampillicin resistance gene (Amp') and an araC gene.

Figure 7B

The nucleic acid sequence and the protein coding portions of pBAD24-recET are depicted.

Figure 8

Manipulation of a large E.coli episome by multiple recombination steps. a Scheme of the recombination reactions. A P1 clone of the Mouse Hoxa complex, resident in JC9604, was modified by recombination with PCR products that contained the neo gene and two Flp recombination targets

(FRTs). The two PCR products were identical except that one was flanked by g and h homology arms (insertion), and the other was flanked by i and h homology arms (deletion). In a second step, the neo gene was removed by Flp recombination between the FRTs by transient transformation of a Flp expression plasmid based on the pSC101 temperature-sensitive origin (ts ori). b Upper panel; ethidium bromide stained agarose gel showing EcoR1 digestions of P1 DNA preparations from three independent colonies for each step. Middle panel; a Southern blot of the upper panel hybridized with a neo gene probe. Lower panel; a Southern blot of the upper panel hybridized with a Hoxa3 probe to visualize the site of recombination. Lanes 1, the original Hoxa3 P1 clone grown in E.coli strain NS3145. Lanes 2, replacement of the Tn903 kanamycin resistance gene resident in the P1 vector with an ampicillin resistance gene increased the 8.1 kb band (lanes 1), to 9.0 kb. Lanes 3, insertion of the Tn5-neo gene with g-h homology arms upstream of Hoxa3, increased the 6.7 kb band (lanes 1,2) to 9.0 kb. Lanes 4, Flp recombinase deleted the g-h neo gene reducing the 9.0 kb band (lanes 3) back to 6.7 kb. Lanes 5, deletion of 6 kb of Hoxa3 - 4 intergenic DNA by replacement with the i-h neo gene, decreased the 6.7 kb band (lanes 2) to 4.5 kb. Lanes 6, Flp recombinase deleted the i-h neo gene reducing the 4.5 kb band to 2.3 kb.

Figure 9

Manipulation of the E.coli chromosome. A Scheme of the recombination reactions. The endogenous lacZ gene of JC9604 at 7.8′ of the E.coli chromosome, shown in expanded form with relevant Ava I sites and coordinates, was targeted by a PCR fragment that contained the neo gene flanked by homology arms j and k, and loxP sites, as depicted. Integration of the neo gene removed most of the lacZ gene including an Ava I site to alter the 1443 and 3027 bp bands into a 3277 bp band. In a second step, the neo gene was removed by Cre recombination between the loxPs by transient transformation of a Cre expression plasmid based on the pSC101

temperature-sensitive origin (ts ori). Removal of the neo gene by Cre recombinase reduces the 3277 band to 2111 bp. b ß-galactosidase expression evaluated by streaking colonies on X-Gal plates. The top row of three streaks show ß-galactosidase expression in the host JC9604 strain (w.t.), the lower three rows (Km) show 24 independent primary colonies, 20 of which display a loss of ß-galactosidase expression indicactive of the intended recombination event. c Southern analysis of E.coli chromosomal DNA digested with Ava I using a random primed probe made from the entire lacZ coding region; lanes 1,2, w.t.; lanes 3-6, four independent white colonies after integration of the j-k neo gene; lanes 7-10; the same four colonies after transient transformation with the Cre expression plasmid.

Figure 10

Two rounds of ET cloning to introduce a point mutation. a Scheme of the recombination reactions. The lacZ gene of pSVpaX1 was disrupted in JC9604lacZ, a strain made by the experiment of Fig.9 to ablate endogenous lacZ expression and remove competitive sequences, by a sacB-neo gene cassette, synthesized by PCR to pIB279 and flanked by l and m homology arms. The recombinants, termed pSV-sacB-neo, were selected on Amp + Kan plates. The lacZ gene of pSV-sacB-neo was then repaired by a PCR fragment made from the intact lacZ gene using l* and m* homology arms. The m* homology arm included a silent C to G change that created a BamH1 site. The recombinants, termed pSVpaX1*, were identified by counter selection against the sacB gene using 7% sucrose. b ß-galactosidase expression from pSVpaX1 was disrupted in pSV-sacB-neo and restored in pSVpaX1*. Expression was analyzed on X-gal plates. Three independent colonies of each pSV-sacB-neo and pSVpaX1* are shown. c Ethidium bromide stained agarose gels of BamH1 digested DNA prepared from independent colonies taken after counter selection with sucrose. All ß-galactosidase expressing colonies (blue) contained the introduced BamH1 restriction site (upper panel). All white colonies displayed large

rearrangements and no product carried the diagnostic 1.5kb BamH1 restriction fragment (lower panel).

Figure 11

Transferance of ET cloning into a recBC+ host to modify a large episome. a Scheme of the plasmid, pBAD-ETγ, which carries the mobile ET system, and the strategy employed to target the Hoxa P1 episome. pBAD-ETγ is based on pBAD24 and includes (i) the truncated recE gene (t-recE) under the arabinose-inducible P_{BAD} promoter; (ii) the recT gene under the EM7 promoter; and (iii) the redγ gene under the Tn5 promoter. It was transformed into NS3145, a recA E.coli strain which contained the Hoxa P1 episome. After arabinose induction, competent cells were prepared and transformed with a PCR product carrying the chloramphenicol resistance gene (cm) flanked by n and p homology arms. n and p were chosen to recombine with a segment of the P1 vector. b Southern blots of Pvu II digested DNAs hybridized with a probe made from the P1 vector to visualize the recombination target site (upper panel) and a probe made from the chloramphenicol resistance gene (lower panel). Lane 1, DNA prepared from cells harboring the Hoxa P1 episome before ET cloning. Lanes 2-17, DNA prepared from 16 independent chloramphenicol resistant colonies.

Figure 12

Comparison of ET cloning using the recE/recT genes in pBAD-ETγ with redα/redß genes in pBAD-αßγ.

The plasmids pBAD-ETγ or pBAD-αßγ, depicted, were transformed into the E.coli recA-, recBC+ strain, DK1 and targeted by a chloramphenicol gene as described in Fig.6 to evaluate ET cloning efficiencies. Arabinose induction of protein expression was for 1 hour.

Figure 13A

The plasmid pBAD-ETγ is shown by diagram.

Figure 13B

The nucleic acid sequence and the protein coding portions of pBAD-ETγ are depicted.

Figure 14A

The plasmid pBAD-αßγ is shown by diagram. This plasmid substantially corresponds to the plasmid shown in Fig.13 except that the recE and recT genes are substituted by the redα and redß genes.

Figure 14B

The nucleic acid sequence and the protein coding portions of pBAD-αßγ are depicted.

1. Methods

1.1. Preparation of linear fragments

Standard PCR reaction conditions were used to amplify linear DNA fragments. The sequences of the primers used are depicted in Table 1.

Table 1

The Tn5-neo gene from pJP5603 (Penfold and Pemberton, Gene 118 (1992), 145-146) was amplified by using oligo pairs a/b and c/d. The chloramphenicol (cm) resistant gene from pMAK705 (Hashimoto-Gotoh and

Sekiguchi, J.Bacteriol.131 (1977), 405-412) was amplified by using primer pairs e/f and n/p. The Tn5-neo gene flanked by FRT or loxP sites was amplified from pKaZ or pKaX (http://www.embl-heidelberg.de/ExternalInfo /stewart) using oligo pairs i/h, g/h and j/k. The sacB-neo cassette from pIB279 (Blomfield et al., Mol.Microbiol.5 (1991), 1447-1457) was amplified by using oligo pair l/m. The lacZ gene fragment from pSVpaZ11 (Buchholz et al., Nucleic Acids Res.24 (1996), 4256-4262) was amplified using oligo pair l*/m*. PCR products were purified using the QIAGEN PCR Purification Kit and eluted with H_2O_2, followed by digestion of any residual template DNA with Dpn I. After digestion, PCR products were extracted once with Phenol:$CHCl_3$, ethanol precipitated and resuspended in H_2O at approximately 0.5 μg/μl.

1.2 Preparation of competent cells and electroporation

Saturated overnight cultures were diluted 50 fold into LB medium, grown to an OD600 of 0.5, following by chilling on ice for 15 min. Bacterial cells were centrifuged at 7,000 rpm for 10 min at 0°C. The pellet was resuspended in ice-cold 10% glycerol and centrifuged again (7,000 rpm, -5°C, 10 min). This was repeated twice more and the cell pellet was suspended in an equal volume of ice-cold 10% glycerol. Aliquots of 50 μl were frozen in liquid nitrogen and stored at -80°C. Cells were thawed on ice and 1 μl DNA solution (containing, for co-transformation, 0.3 μg plasmid and 0.2 μg PCR products; or, for transformation, 0.2 μg PCR products) was added. Electroporation was performed using ice-cold cuvettes and a Bio-Rad Gene Pulser set to 25 μFD, 2.3 kV with Pulse Controller set at 200 ohms. LB medium (1 ml) was added after electroporation. The cells were incubated at 37°C for 1 hour with shaking and then spread on antibiotic plates.

1.3 Induction of RecE and RecT expression

E.coli JC5547 carrying pBAD24-recET was cultured overnight in LB medium plus 0.2% glucose, 100 μg/ml ampicillin. Five parallel LB cultures, one of which (0) included 0.2% glucose, were started by a 1/100 inoculation. The cultures were incubated at 37°C with shaking for 4 hours and 0.1% L-arabinose was added 3, 2, 1 or 1/2 hour before harvesting and processing as above. Immediately before harvesting, 100 μl was removed for analysis on a 10% SDS-polyacrylamide gel. E.coli NS3145 carrying Hoxa-P1 and pBAD-ETγ was induced by 0.1% L-arabinose for 90 min before harvesting.

1.4 Transient transformation of FLP and Cre expression plasmids

The FLP and Cre expression plasmids, 705-Cre and 705-FLP (Buchholz et al, Nucleic Acids Res. 24 (1996), 3118-3119), based on the pSC101 temperature sensitive origin, were transformed into rubidium chloride competent bacterial cells. Cells were spread on 25 μg/ml chloramphenicol plates, and grown for 2 days at 30°C, whereupon colonies were picked, replated on L-agar plates without any antibiotics and incubated at 40°C overnight. Single colonies were analyzed on various antibiotic plates and all showed the expected loss of chloramphenicol and kanamycin resistance.

1.5 Sucrose counter selection of sacB expression

The E.coli JC9604lacZ strain, generated as described in Fig.11, was cotransformed with a sacB-neo PCR fragment and pSVpaX1 (Buchholz et al, Nucleic Acids Res. 24 (1996), 4256-4262). After selection on 100 μg/ml ampicillin, 50 μg/ml kanamycin plates, pSVpaX-sacB-neo plasmids were isolated and cotransformed into fresh JC9604lacZ cells with a PCR fragment amplified from pSVpaX1 using primers l*/m*. Oligo m* carried a silent point mutation which generated a BamHI site. Cells were plated on 7% sucrose, 100 μg/ml ampicillin, 40 μg/ml X-gal plates and incubated at

28°C for 2 days. The blue and white colonies grown on sucrose plates were counted and further checked by restriction analysis.

1.6 Other methods

DNA preparation and Southern analysis were performed according to standard procedures. Hybridization probes were generated by random priming of fragments isolated from the Tn5 neo gene (PvuII), Hoxa3 gene (both HindIII fragments), lacZ genes (EcoR1 and BamH1 fragments from pSVpaX1), cm gene (BstB1 fragments from pMAK705) and P1 vector fragments (2.2 kb EcoR1 fragments from P1 vector).

2. Results

2.1 Identification of recombination events in E.coli

To identify a flexible homologous recombination reaction in E.coli, an assay based on recombination between linear and circular DNAs was designed (Fig.1, Fig.3). Linear DNA carrying the Tn5 kanamycin resistance gene (neo) was made by PCR (Fig.3a). Initially, the oligonucleotides used for PCR amplification of neo were 60mers consisting of 42 nucleotides at their 5' ends identical to chosen regions in the plasmid and, at the 3' ends, 18 nucleotides to serve as PCR primers. Linear and circular DNAs were mixed in equimolar proportions and co-transformed into a variety of E.coli hosts. Homologous recombination was only detected in sbcA E.coli hosts. More than 95% of double ampicillin/kanamycin resistant colonies (Fig.3b) contained the expected homologously recombined plasmid as determined by restriction digestion and sequencing. Only a low background of kanamycin resistance, due to genomic integration of the neo gene, was apparent (not shown).

The linear plus circular recombination reaction was characterized in two ways. The relationship betweeen homology arm length and recombination efficiency was simple, with longer arms recombining more efficiently (Fig.3c). Efficiency increased within the range tested, up to 60 bp. The effect of distance between the two chosen homology sites in the recipient plasmid was examined (Fig.3d). A set of eight PCR fragments was generated by use of a constant left homology arm with differing right homology arms. The right homology arms were chosen from the plasmid sequence to be 0 - 3100 bp from the left. Correct products were readily obtained from all, with less than 4 fold difference between them, although the insertional product (0) was least efficient. Correct products also depended on the presence of both homology arms, since PCR fragments containing only one arm failed to work.

2.2 Involvement of RecE and RecT

The relationship between host genotype and this homologous recombination reaction was more systematically examined using a panel of E.coli strains deficient in various recombination components (Table 2).

Table 2

Only the two sbcA strains, JC8679 and JC9604 presented the intended recombination products and RecA was not required. In sbcA strains, expression of RecE and RecT is activated. Dependence on recE can be inferred from comparison of JC8679 with JC8691. Notably no recombination products were observed in JC9387 suggesting that the sbcBC background is not capable of supporting homologous recombination based on 50 nucleotide homology arms.

To demonstrate that RecE and RecT are involved, part of the recET operon was cloned into an inducible expression vector to create pBAD24-recET

(Fig.6a). the recE gene was truncated at its N-terminal end, as the first 588 a.a.s of RecE are dispensable. The recBC strain, JC5547, was transformed with pBAD24-recET and a time course of RecE/RecT induction performed by adding arabinose to the culture media at various times before harvesting for competent cells. The batches of harvested competent cells were evaluated for protein expression by gel electrophoresis (Fig.6b) and for recombination between a linear DNA fragment and the endogenous pBAD24-recET plasmid (Fig.6c). Without induction of RecE/RecT, no recombinant products were found, whereas recombination increased in approximate concordance with increased RecE/RecT expression. This experiment also shows that co-transformation of linear and circular DNAs is not essential and the circular recipient can be endogenous in the host. From the results shown in Figs.3, 6 and Table 2, we conclude that RecE and RecT mediate a very useful homologous recombination reaction in recBC E.coli at workable frequencies. Since RecE and RecT are involved, we refer to this way of recombining linear and circular DNA fragments as "ET cloning".

2.3 Application of ET cloning to large target DNAs

To show that large DNA episomes could be manipulated in E.coli, a > 76 kb P1 clone that contains at least 59 kb of the intact mouse Hoxa complex, (confirmed by DNA sequencing and Southern blotting), was transferred to an E.coli strain having an sbcA background (JC9604) and subjected to two rounds of ET cloning. In the first round, the Tn903 kanamycin resistance gene resident in the P1 vector was replaced by an ampicillin resistance gene (Fig.4). In the second round, the interval between the Hoxa3 and a4 genes was targeted either by inserting the neo gene between two base pairs upstream of the Hoxa3 proximal promoter, or by deleting 6203 bp between the Hoxa3 and a4 genes (Fig.8a). Both insertional and deletional ET cloning products were readily obtained (Fig.8b, lanes 2, 3 and 5) showing that the

two rounds of ET cloning took place in this large E.coli episome with precision and no apparent unintended recombination.

The general applicability of ET cloning was further examined by targeting a gene in the E.coli chromosome (Fig.9a). The ß-galactosidase (lacZ) gene of JC9604 was chosen so that the ratio between correct and incorrect recombinants could be determined by evaluating ß-galactosidase expression. Standard conditions (0.2 μg PCR fragment; 50 μl competent cells), produced 24 primary colonies, 20 of which were correct as determined by ß-galactosidase expression (Fig.9b), and DNA analysis (Fig.9c, lanes 3-6).

2.4 Secondary recombination reactions to remove operational sequences

The products of ET cloning as described above are limited by the necessary inclusion of selectable marker genes. Two different ways to use a further recombination step to remove this limitation were developed. In the first way, site specific recombination mediated by either Flp or Cre recombinase was employed. In the experiments of Figs.8 and 9, either Flp recombination target sites (FRTs) or Cre recombination target sites (loxPs) were included to flank the neo gene in the linear substrates. Recombination between the FRTs or loxPs was accomplished by Flp or Cre, respectively, expressed from plasmids with the pSC101 temperature sensitive replication origin (Hashimoto-Gotoh and Sekiguchi, J.Bacteriol. 131 (1977), 405-412) to permit simple elimination of these plasmids after site specific recombination by temperature shift. The precisely recombined Hoxa P1 vector was recovered after both ET and Flp recombination with no other recombination products apparent (Fig.8, lanes 4 and 6). Similarly, Cre recombinase precisely recombined the targeted lacZ allele (Fig.9, lanes 7-10). Thus site specific recombination can be readily coupled with ET cloning to remove operational sequences and leave a 34 bp site specific recombination target site at the point of DNA manipulation.

In the second way to remove the selectable marker gene, two rounds of ET cloning, combining positive and counter selection steps, were used to leave the DNA product free of any operational sequences (Fig.10a).

Additionally this experiment was designed to evaluate, by a functional test based on ß-galactosidase activity, whether ET cloning promoted small mutations such as frame shift or point mutations within the region being manipulated. In the first round, the lacZ gene of pSVpaX1 was disrupted with a 3.3 kb PCR fragment carrying the neo and B.subtilis sacB (Blomfield et al., Mol.Microbiol. 5 (1991), 1447-1457) genes, by selection for kanamycin resistance (Fig.10a). As shown above for other positively selected recombination products, virtually all selected colonies were white (Fig.10b), indicative of successful lacZ disruption, and 17 of 17 were confirmed as correct recombinants by DNA analysis. In the second round, a 1.5 kb PCR fragment designed to repair lacZ was introduced by counter selection against the sacB gene. Repair of lacZ included a silent point mutation to create a BamH1 restriction site. Approximately one quarter of sucrose resistant colonies expressed ß-galactosidase, and all analyzed (17 of 17; Fig.10c) carried the repaired lacZ gene with the BamH1 point mutation. The remaining three quarters of sucrose resistant colonies did not express ß-galactosidase, and all analyzed (17 of 17; Fig.10c) had undergone a variety of large mutational events, none of which resembled the ET cloning product. Thus, in two rounds of ET cloning directed at the lacZ gene, no disturbances of ß-galactosidase activity by small mutations were observed, indicating the RecE/RecT recombination works with high fidelity. The significant presence of incorrect products observed in the counter selection step is an inherent limitation of the use of counter selection, since any mutation that ablates expression of the counter selection gene will be selected. Notably, all incorrect products were large mutations and therefore easily distinguished from the correct ET product by DNA analysis. In a different experiment (Fig.5), we observed that ET cloning into pZero2.1 (InVitroGen) by counter selection against the ccdB gene gave

a lower background of incorrect products (8%), indicating that the counter selection background is variable according to parameters that differ from those that influence ET cloning efficiencies.

2.5 Transference of ET cloning between E.coli hosts

The experiments shown above were performed in recBC- E.coli hosts since the sbcA mutation had been identified as a suppressor of recBC (Barbour et al., Proc.Natl.Acad.Sci. USA 67 (1970), 128-135; Clark, Genetics 78 (1974), 259-271). However, many useful E.coli strains are recBC+, including strains commonly used for propagation of P1, BAC or PAC episomes. To transfer ET cloning into recBC + strains, we developed pBAD-ETγ and pBAD-αßγ (Figs.13 and 14). These plasmids incorporate three features important to the mobility of ET cloning. First, RecBC is the major E.coli exonuclease and degrades introduced linear fragments. Therefore the RecBC inhibitor, Redγ (Murphy, J.Bacteriol. 173 (1991), 5808-5821), was included. Second, the recombinogenic potential of RecE/RecT, or Redα/Redß, was regulated by placing recE or redα under an inducible promoter. Consequently ET cloning can be induced when required and undesired recombination events which are restricted at other times. Third, we observed that ET cloning efficiencies are enhanced when RecT, or Redß, but not RecE, or Redα, is overexpressed. Therefore we placed recT, or redß, under the strong, constitutive, EM7 promoter.

pBAD-ETγ was transformed into NS3145 E.coli harboring the original Hoxa P1 episome (Fig.11a). A region in the P1 vector backbone was targeted by PCR amplification of the chloramphenicol resistance gene (cm) flanked by n and p homology arms. As described above for positively selected ET cloning reactions, most (> 90%) chloramphenicol resistant colonies were correct. Notably, the overall efficiency of ET cloning, in terms of linear DNA transformed, was nearly three times better using pBAD-ETγ than with similar experiments based on targeting the same episome in the sbcA host,

JC9604. This is consistent with our observation that overexpression of RecT improves ET cloning efficiencies.

A comparison between ET cloning efficiencies mediated by RecE/RecT, expressed from pBAD-ETγ, and Redα/Redß, expressed from pBAD-αßγ was made in the recA-, recBC + E.coli strain, DK1 (Fig.12). After transformation of E.coli DK1 with either pBAD-ETγ or pBAD-αßγ, the same experiment as described in Figure 6a,c, to replace the bla gene of the pBAD vector with a chloramphenicol gene was performed. Both pBAD-ETγ or pBAD-αßγ presented similar ET cloning efficiencies in terms of responsiveness to arabinose induction of RecE and Redα, and number of targeted events.

Table 2

E.coli Strains	Genotypes	Amp+Kan	Amp
			x 10^8/µg
JC8679	*recBC sbcA*	318	2.30
JC9604	*recA recBC sbcA*	114	0.30
JC8691	*recBC sbcA recE*	0	0.37
JC5547	*recA recBC*	0	0.37
JC5519	*recBC*	0	1.80
JC15329	*recA recBC sbcBC*	0	0.03
JC9387	*recBC sbcBC*	0	2.20
JC8111	*recBC sbcBC recF*	0	2.40
JC9366	*recA*	0	0.37
JC13031	*recJ*	0	0.45

Claims

1. A method for cloning DNA molecules in cells comprising the steps of:

   a) providing a host cell capable of performing homologous recombination,

   b) contacting in said host cell a first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and

   c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred.

2. The method according to claim 1 wherein the homologous recombination occurs via the recET cloning mechanism.

3. The method according to claim 2 wherein the host cell is capable of expressing recE and recT genes.

4. The method according to claim 3 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

5. The method according to claim 3 or 4 wherein the host cell is transformed with at least one vector capable of expressing recE and/or recT genes.

6. The method of claim 3, 4 or 5 wherein the expression of the recE and/or recT genes is under control of a regulatable promoter.

7. The method of claim 5 or 6 wherein the recT gene is overexpressed versus the recE gene.

8. The method according to any one of claims 3 to 7 wherein the recE gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 1320 (ATG) to 2159 (GAC) as depicted in Fig.7B,
(b) the nucleic acid sequence from position 1320 (ATG) to 1998 (CGA) as depicted in Fig.13B,
(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequence from (a), (b) and/or (c).

9. The method according to any one of claims 3 to 8 wherein the recT gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 2155 (ATG) to 2961 (GAA) as depicted in Fig.7B,
(b) the nucleic acid sequence from position 2086 (ATG) to 2868 (GCA) as depicted in Fig.13B,
(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequences from (a), (b) and/or (c).

10. The method according to any one of the previous claims wherein the host cell is a gram-negative bacterial cell.

11. The method according to claim 10 wherein the host cell is an Escherichia coli cell.

12. The method according to claim 11 wherein the host cell is an Escherichia coli K12 strain.

13. The method according to claim 12 wherein the E.coli strain is selected from JC 8679 and JC 9604.

14. The method according to any one of the previous claims wherein the host cell further is capable of expressing a recBC inhibitor gene.

15. The method according to claim 14 wherein the host cell is transformed with a vector expressing the recBC inhibitor gene.

16. The method according to claim 14 or 15 wherein the recBC inhibitor gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 3588 (ATG) to 4002 (GTA) as depicted in Fig.13B,
(b) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(c) a nucleic acid sequence which hybridizes under stringent conditions (as defined above) with the nucleic acid sequence from (a) and/ or (b).

17. The method according to any one of claims 13 to 16 wherein the host cell is a prokaryotic recBC+ cell.

18. The method according to any one of the previous claims wherein the first DNA molecule is circular.

19. The method according to any one of the previous claims wherein the first DNA molecule is an extrachromosomal DNA molecule containing an origin of replication which is operative in the host cell.

20. The method according to claim 18 or 19 wherein the first DNA molecule is selected from plasmids, cosmids, P1 vectors, BAC vectors and PAC vectors.

21. The method according to any one of claims 1-18 wherein the first DNA molecule is a host cell chromosome.

22. The method according to any one of the previous claims wherein the second DNA molecule is linear.

23. The method according to any one of the previous claims wherein the regions of sequence homology are at least 15 nucleotides each.

24. The method according to one of claims 1 to 16 wherein the second DNA molecule is obtained by an amplification reaction.

25. The method according to one of the previous claims wherein the first and/or second DNA molecules are introduced into the host cells by transformation.

26. The method according to claim 25 wherein the transformation method is electroporation.

27. The method according to one of claims 1 to 26 wherein the first and second DNA molecules are introduced into the host cell simultaneously by co-transformation.

28. The method according to one of claims 1 to 26 wherein the second DNA molecule is introduced into a host cell in which the first DNA molecule is already present.

29. The method according to one of the previous claims wherein the second DNA molecule contains at least one marker gene placed between the two regions of sequence homology and wherein homologous recombination is detected by expression of said marker gene.

30. The method according to claim 29 wherein gene presence is selected from antibiotic resistance genes, deficiency complementation genes and reporter genes.

31. The method of any one of claims 1 to 30 wherein the first DNA molecule contains at least one marker gene between the two regions of sequence homology and wherein homologous recombination is detected by lack of expression of said marker gene.

32. The method of any one of claims 1 to 31 wherein said marker gene is selected from genes which, under selected conditions, convey a toxic or bacteriostatic effect on the cell, and reporter genes.

33. A method according to any one of the previous claims wherein the first DNA molecule contains at least one target site for a site specific recombinase between the two regions of sequence homology and wherein homologous recombination is detected by removal of said target site.

34. A method for cloning DNA molecules comprising the steps of:
(a) providing a source of RecE and RecT proteins,
(b) contacting a first DNA molecule which is capable of being replicated in a suitable host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DMA molecules and

(c) selecting DNA molecules in which homologous recombination between said first and second DNA molecules has occurred.

35. The method of claim 34 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

36. The method of claim 34 or 35 wherein the recombination occurs in vitro.

37. The method of claim 34 or 35 wherein the recombination occurs in vivo.

38. Use of cells capable of expressing the recE and recT genes as a host cell for a cloning method involving homologous recombination.

39. Use of a vector system capable of expressing recE and recT genes in a host cell for a cloning method involving homologous recombination.

40. Use of claims 38 or 39 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

41. Use of a source of RecE and RecT proteins for a cloning method involving homologous recombination.

42. Use of claim 41 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

43. A reagent kit for cloning comprising
(a) a host cell
(b) means of expressing recE and recT genes in said host cell and
(c) a recipient cloning vehicle capable of being replicated in said cell.

44. The reagent kit according to claim 43 wherein the means (b) comprise a vector system capable of expressing the recE and recT genes in the host cell.

45. The reagent kit according to claim 43 or 44 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

46. A reagent kit for cloning comprising
(a) a source for RecE and RecT proteins and
(b) a recipient cloning vehicle capable of being propagated in a host cell.

47. The reagent kit according to claim 46 further comprising a host cell suitable for propagating said recipient cloning vehicle.

48. The reagent kit according to claim 46 or 47 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

49. The reagent kit according to any one of claims 43-48 further comprising means for expressing a site specific recombinase in said host cell.

50. The reagent kit according to any one of claims 43-49 further comprising nucleic acid amplification primers comprising a region of homology to said recipient cloning vehicle.

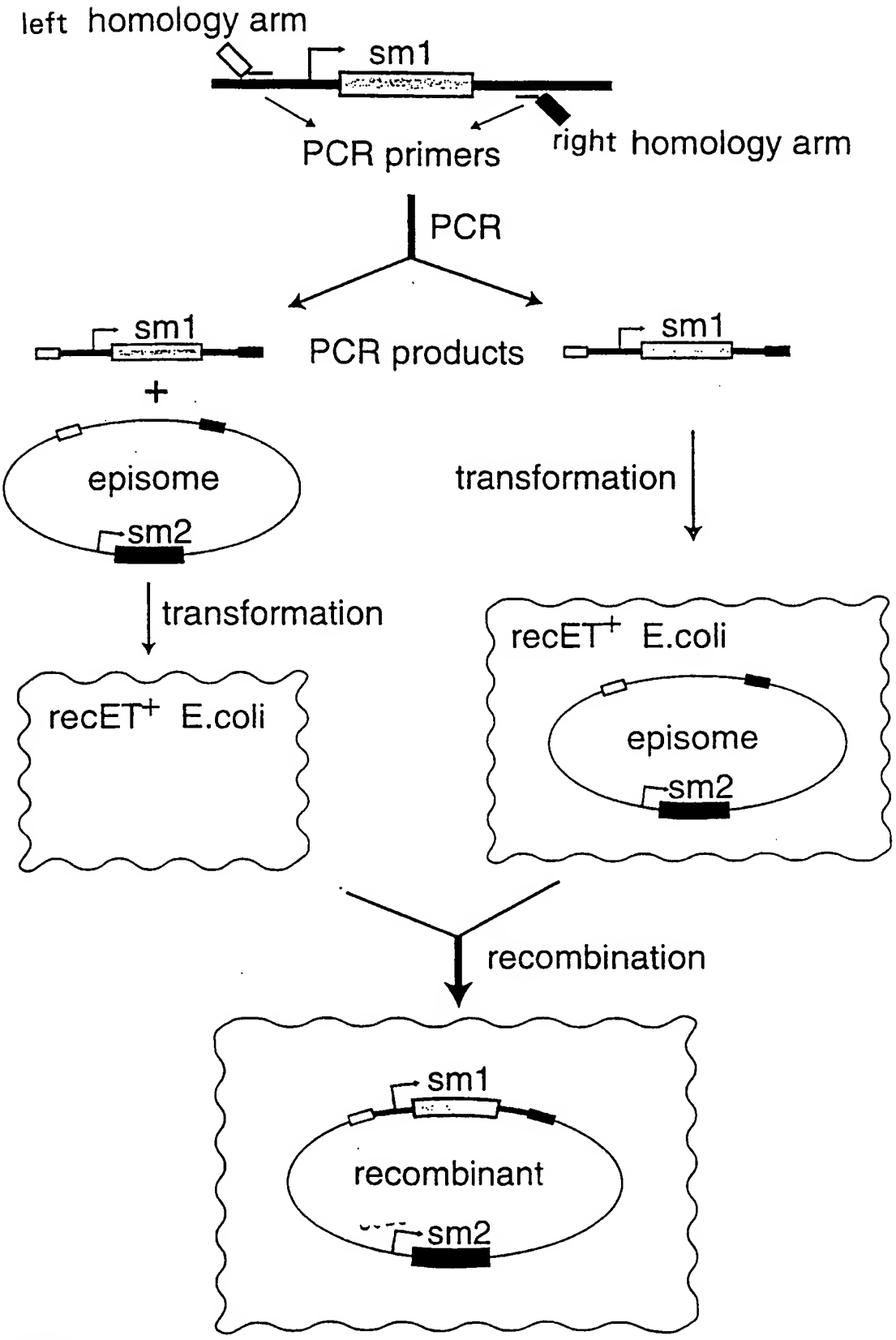

left homology arm
sm1
PCR primers
right homology arm
PCR
sm1
PCR products
sm1
+
episome
sm2
transformation
transformation
recET+ E.coli
recET+ E.coli
episome
sm2
recombination
sm1
recombinant
sm2

Figure 1

Three ways to select recombinants

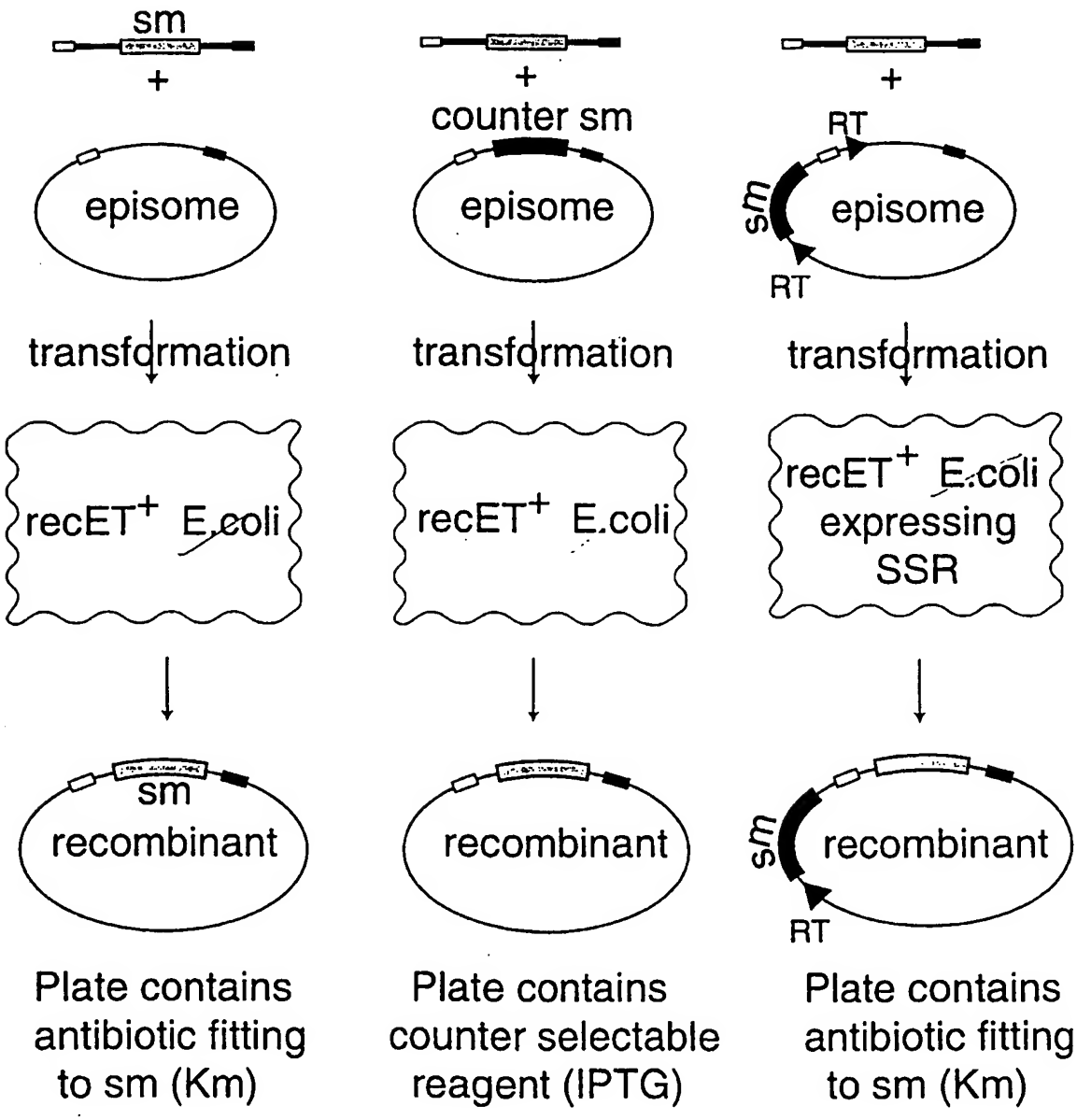
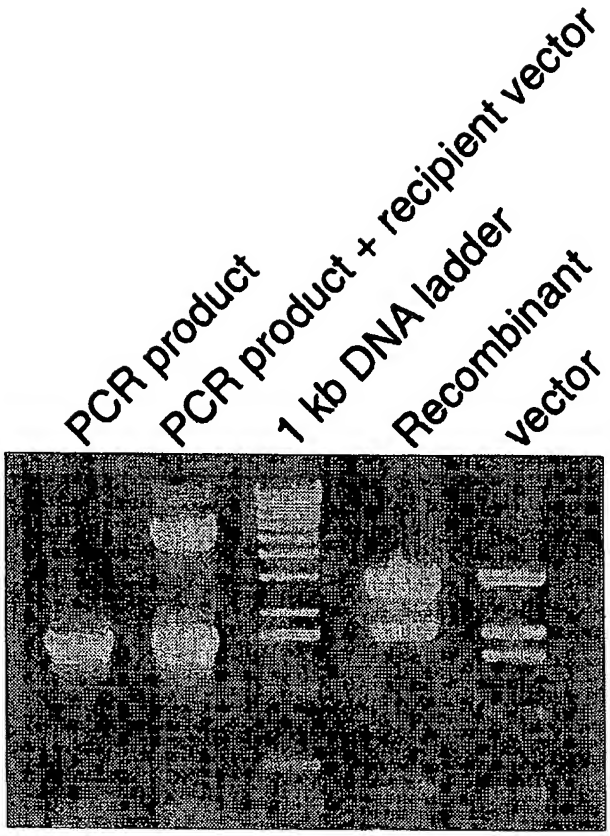


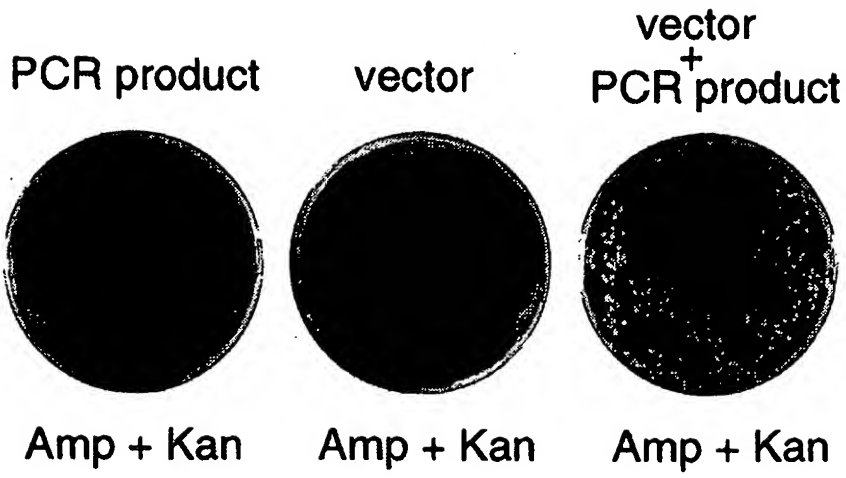
Figure 2

Figure 3

a


PCR product
PCR product + recipient vector
1 kb DNA ladder
Recombinant
vector

b


PCR product
vector
vector + PCR product
Amp + Kan
Amp + Kan
Amp + Kan

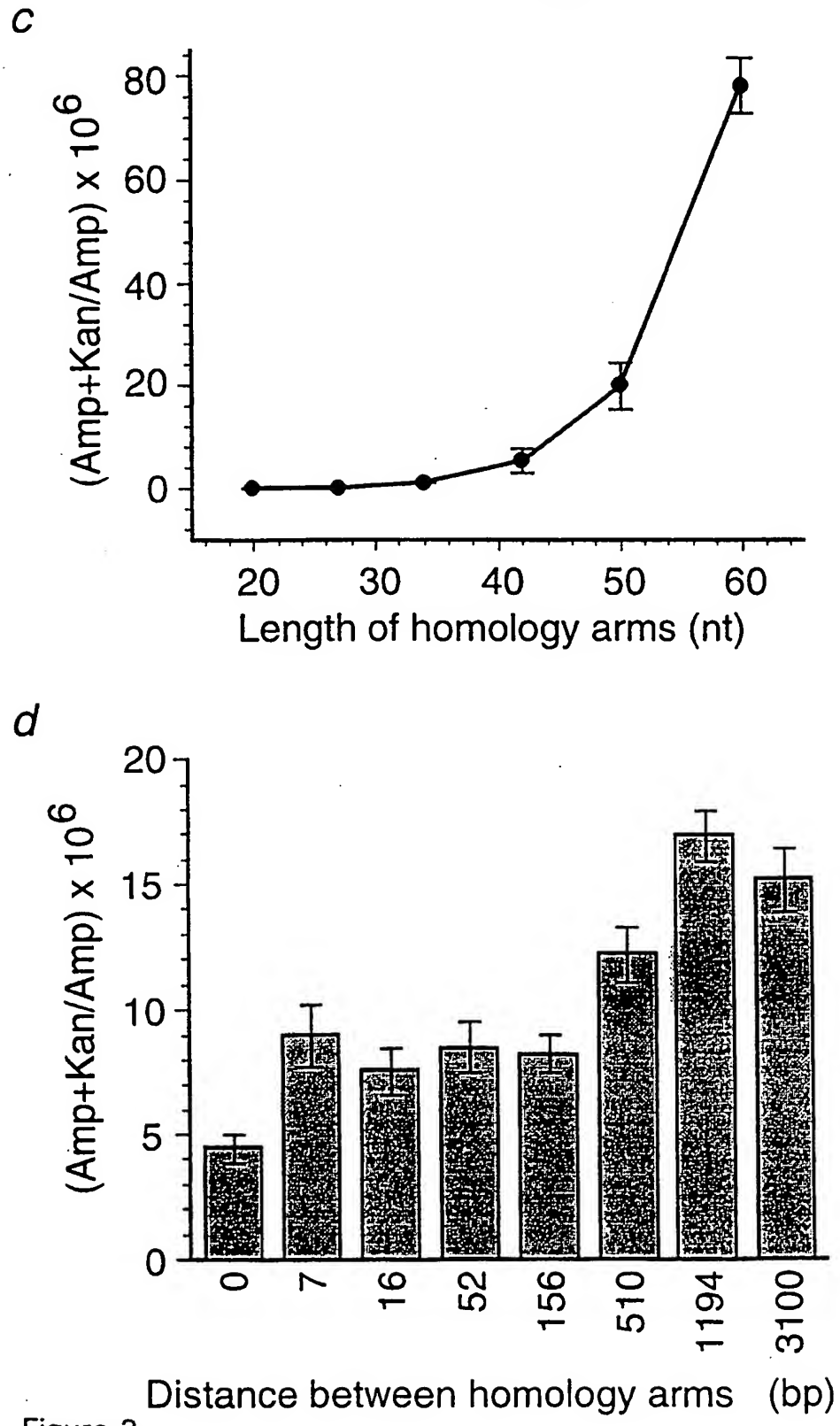

c
(Amp+Kan/Amp) x 10^6
0
20
40
60
80
20
30
40
50
60
Length of homology arms (nt)
d
(Amp+Kan/Amp) x 10^6
0
5
10
15
20
0
7
16
52
156
510
1194
3100
Distance between homology arms (bp)

Figure 3

Figure 4a

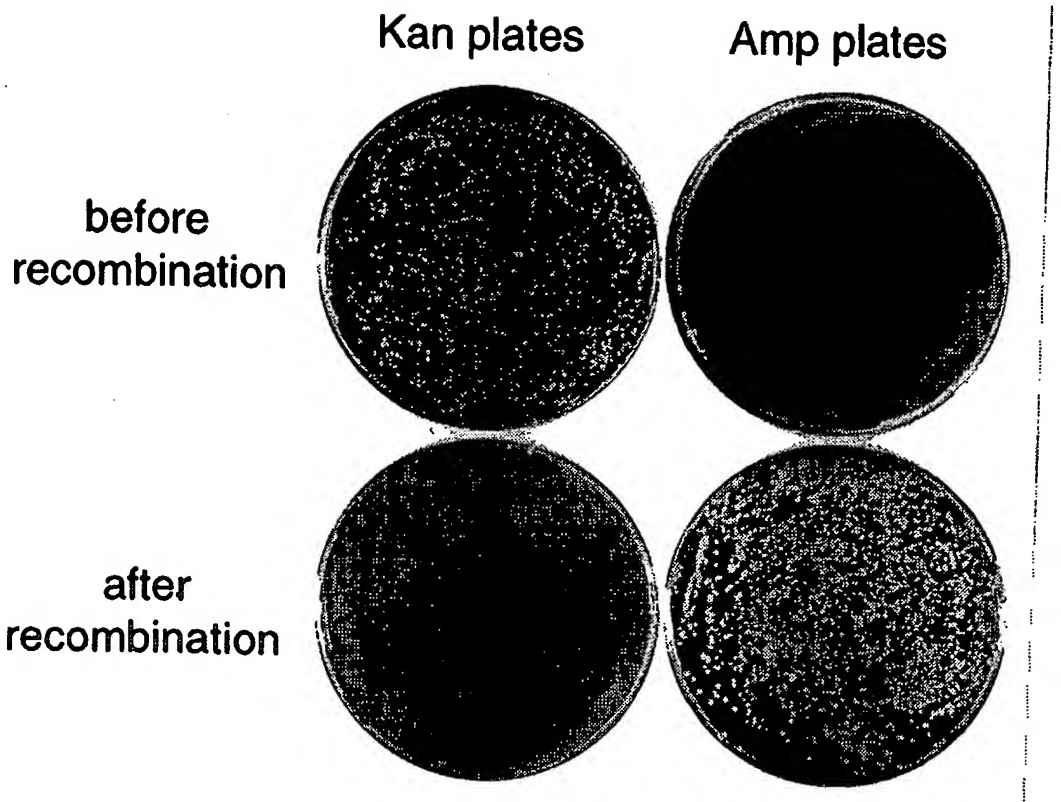
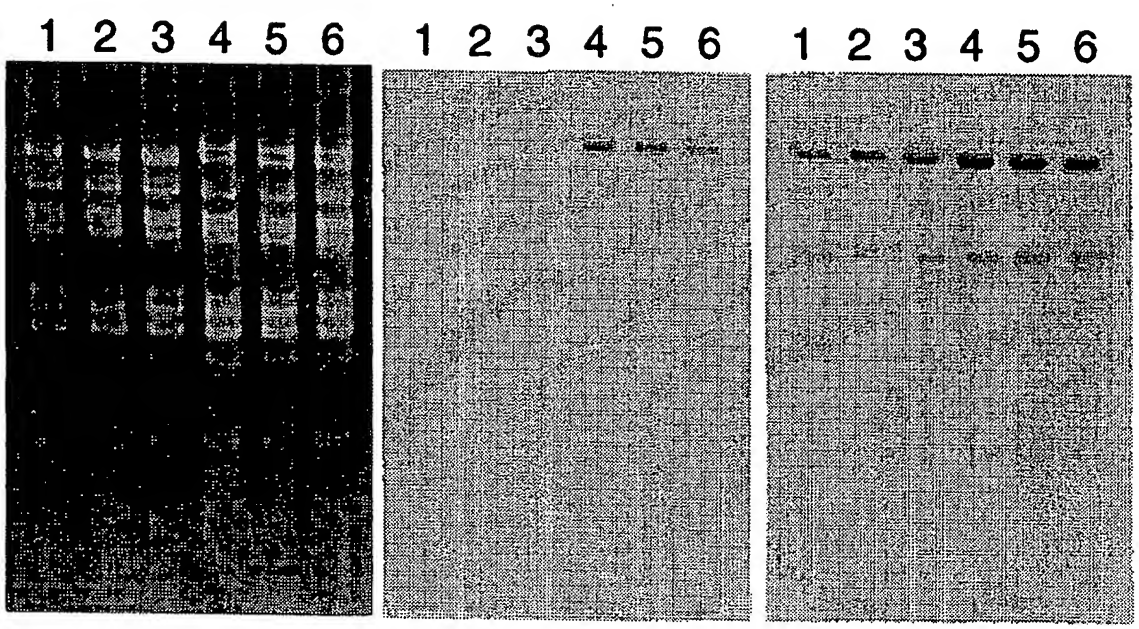

Kan plates
Amp plates
before recombination
after recombination

Figure 4b



P1 DNA digested with EcoR I | hybridized with a bla probe (Amp) | hybridized with a Hoxa-3 probe

Lane 1: 1 of P1-Hox clone in NS3145 original bacterial strain (Kan resistance)
Lane 2-3: 2 of P1-Hox clones in JC9604 before homologous recombination (Kan resistance)
Lane 4-6: 3 of P1-Hox clones in JC9604 after homologous recombination (Amp resistance)

Figure 5

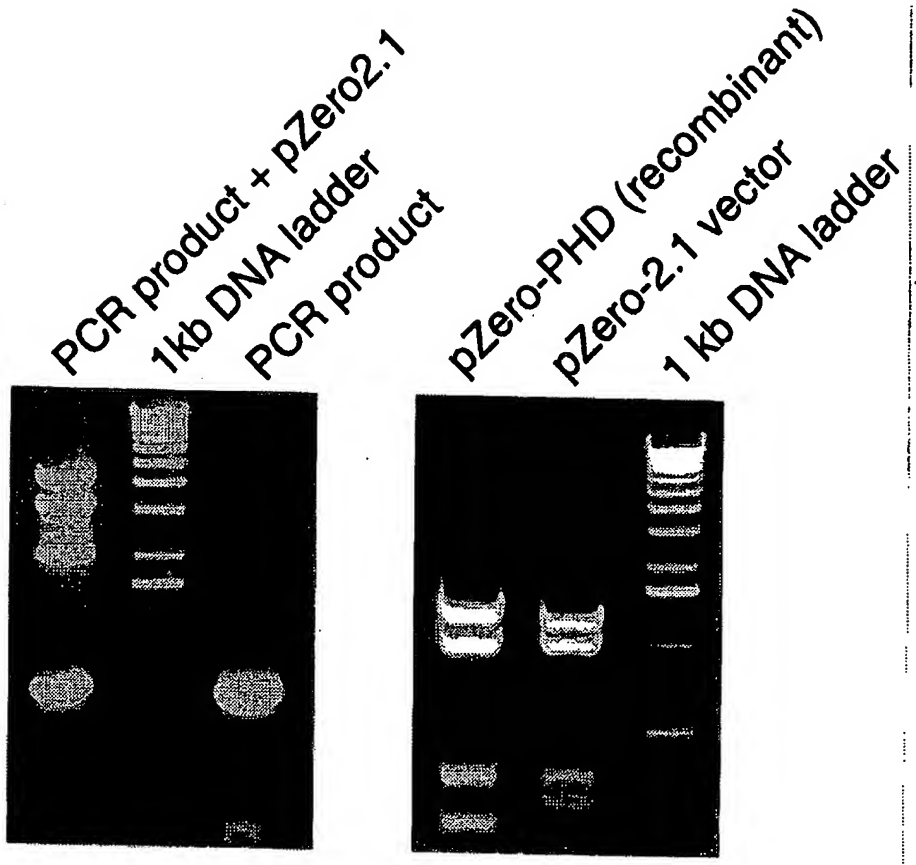

a
PCR product + pZero2.1
1kb DNA ladder
PCR product
pZero-PHD (recombinant)
pZero-2.1 vector
1 kb DNA ladder
b
pZero-2.1
alone
pZero-2.1
+
PCR products
IPTG + Kan

Figure 6

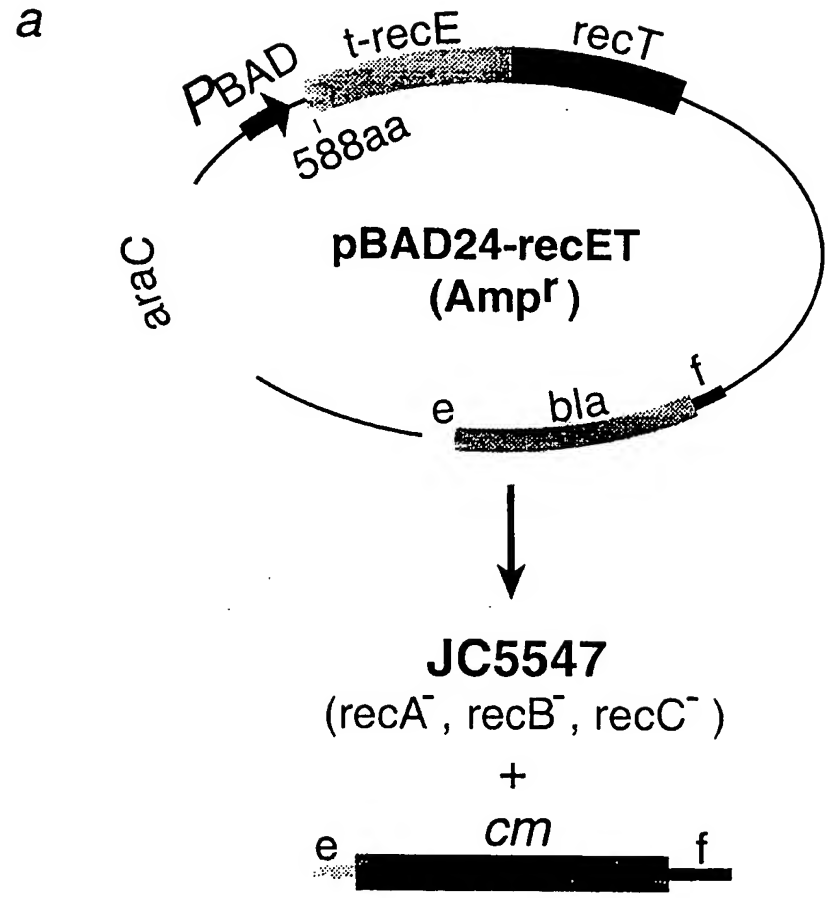
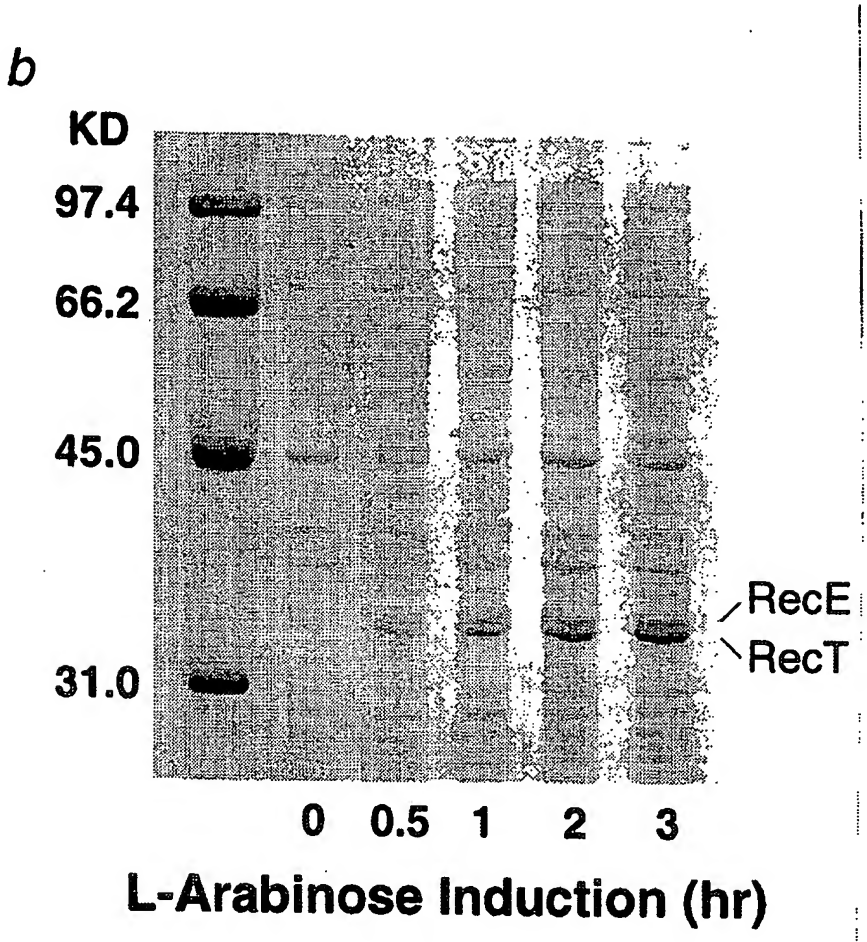

a
PBAD
t-recE
recT
588aa
pBAD24-recET
(Ampr)
araC
e
bla
f
JC5547
(recA-, recB-, recC-)
+
cm
e
f

Figure 6


b
KD
97.4
66.2
45.0
31.0
RecE
RecT
0 0.5 1 2 3
L-Arabinose Induction (hr)

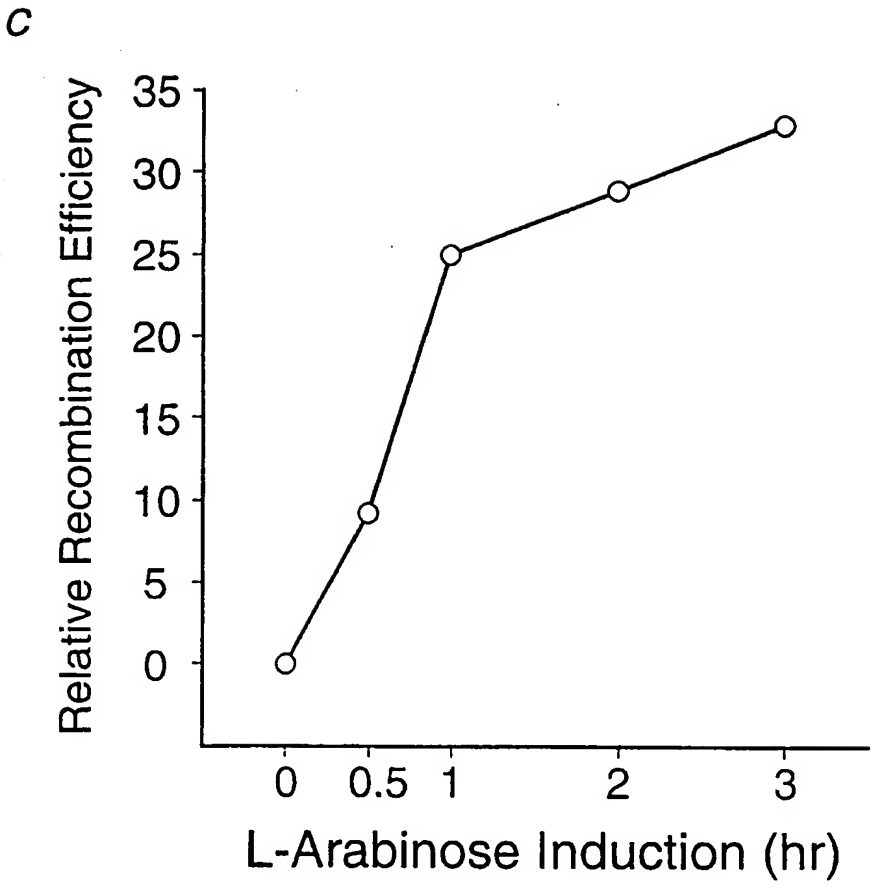
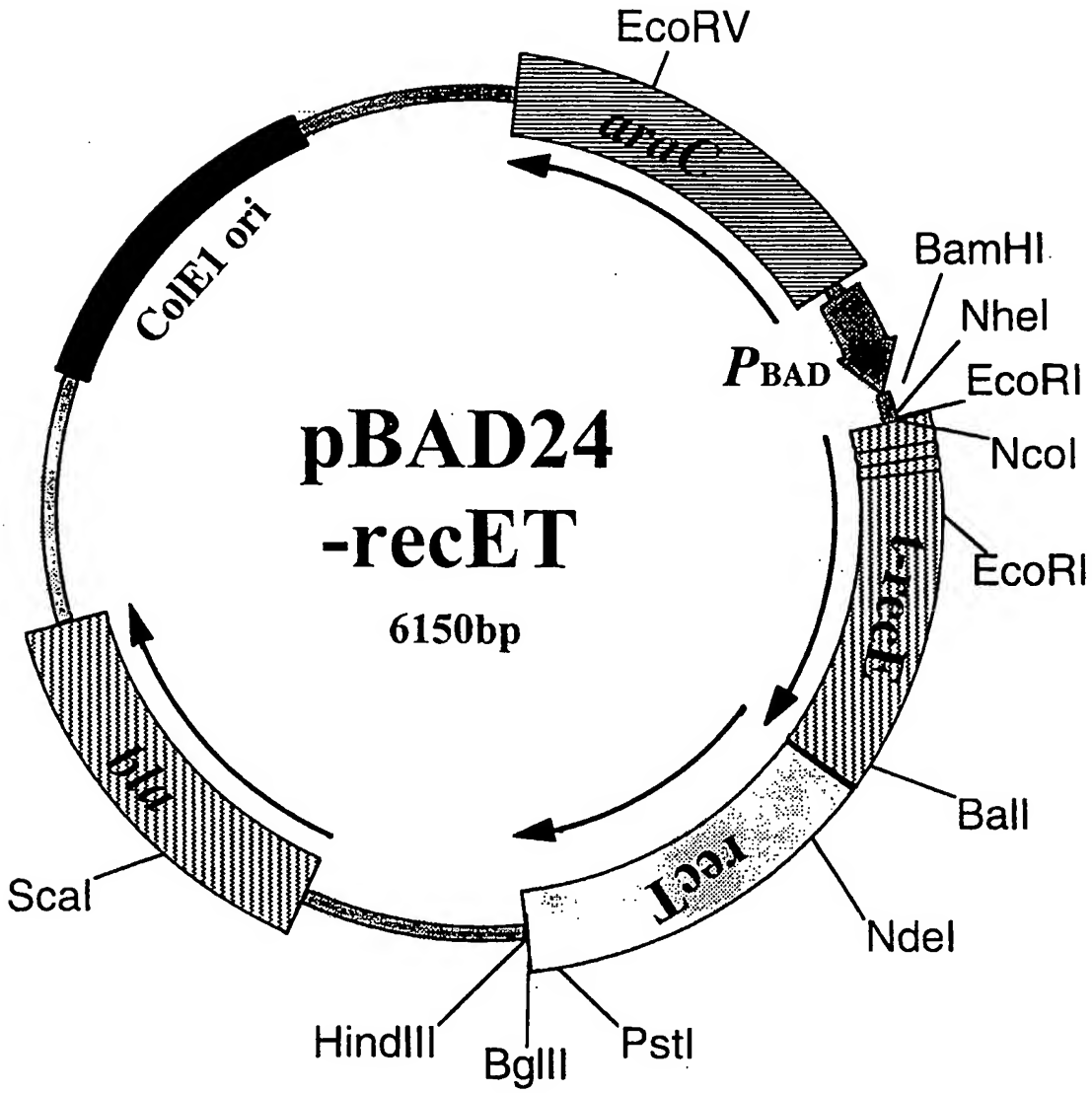

C
Relative Recombination Efficiency
35
30
25
20
15
10
5
0
0
0.5
1
2
3
L-Arabinose Induction (hr)

Figure 6

Figure 7a



t-recE --- truncated recE (from 588 aa ---> end. 866 aa)

Figure 7b

```
  1 ATCGATGCATAATGTGCCTGTCAAATGGACGAAGCAGGGATTC
 44 TGCAAACCCTATGCTACTCCGTCAAGCCGTCAATTGTCTGATT
 87 CGTTACCAA TTA TGA CAA CTT GAC GGC TAC ATC
          293◀••• Ser Leu Lys Val Ala Val Asp
120 ATT CAC TTT TTC TTC ACA ACC GGC ACG GAA CTC
285◀Asn Val Lys Glu Glu Cys Gly Ala Arg Phe Glu
153 GCT CGG GCT GGC CCC GGT GCA TTT TTT AAA TAC
274◀Ser Pro Ser Ala Gly Thr Cys Lys Lys Phe Val
186 CCG CGA GAA ATA GAG TTG ATC GTC AAA ACC AAC
263◀Arg Ser Phe Tyr Leu Gln Asp Asp Phe Gly Val
219 ATT GCG ACC GAC GGT GGC GAT AGG CAT CCG GGT
252◀Asn Arg Gly Val Thr Ala Ile Pro Met Arg Thr
252 GGT GCT CAA AAG CAG CTT CGC CTG GCT GAT ACG
241◀Thr Ser Leu Leu Leu Lys Ala Gln Ser Ile Arg
285 TTG GTC CTC GCG CCA GCT TAA GAC GCT AAT CCC
230◀Gln Asp Glu Arg Trp Ser Leu Val Ser Ile Gly
318 TAA CTG CTG GCG GAA AAG ATG TGA CAG ACG CGA
219◀Leu Gln Gln Arg Phe Leu His Ser Leu Arg Ser
351 CGG CGA CAA GCA AAC ATG CTG TGC GAC GCT GGC
208◀Pro Ser Leu Cys Val His Gln Ala Val Ser Ala
     EcoRV
384 GAT ATC AAA ATT GCT GTC TGC CAG GTG ATC GCT
197◀Ile Asp Phe Asn Ser Asp Ala Leu His Asp Ser
417 GAT GTA CTG ACA AGC CTC GCG TAC CCG ATT ATC
186◀Ile Tyr Gln Cys Ala Glu Arg Val Arg Asn Asp
```

Figure 7b (cont'd)

```
450 CAT CGG TGG ATG GAG CGA CTC GTT AAT CGC TTC
175 Met Pro Pro His Leu Ser Glu Asn Ile Ala Glu

483 CAT GCG CCG CAG TAA CAA TTG CTC AAG CAG ATT
164 Met Arg Arg Leu Leu Leu Gln Glu Leu Leu Asn

516 TAT CGC CAG CAG CTC CGA ATA GCG CCC TTC CCC
153 Ile Ala Leu Leu Glu Ser Tyr Arg Gly Glu Gly

549 TTG CCC GGC GTT AAT GAT TTG CCC AAA CAG GTC
142 Gln Gly Ala Asn Ile Ile Gln Gly Phe Leu Asp

582 GCT GAA ATG CGG CTG GTG CGC TTC ATC CGG GCG
131 Ser Phe His Pro Gln His Ala Glu Asp Pro Arg

615 AAA GAA CCC CGT ATT GGC AAA TAT TGA CGG CCA
120 Phe Phe Gly Thr Asn Ala Phe Ile Ser Pro Trp

648 GTT AAG CCA TTC ATG CCA GTA GGC GCG CGG ACG
109 Asn Leu Trp Glu His Trp Tyr Ala Arg Pro Arg

681 AAA GTA AAC CCA CTG GTG ATA CCA TTC GCG AGC
 98 Phe Tyr Val Trp Gln His Tyr Trp Glu Arg Ala

714 CTC CGG ATG ACG ACC GTA GTG ATG AAT CTC TCC
 87 Glu Pro His Arg Gly Tyr His His Ile Glu Gly

747 TGG CGG GAA CAG CAA AAT ATC ACC CGG TCG GCA
 76 Pro Pro Phe Leu Leu Ile Asp Gly Pro Arg Cys

780 AAC AAA TTC TCG TCC CTG ATT TTT CAC CAC CCC
 65 Val Phe Glu Arg Gly Gln Asn Lys Val Val Gly

813 CTG ACC GCG AAT GGT GAG ATT GAG AAT ATA ACC
 54 Gln Gly Arg Ile Thr Leu Asn Leu Ile Tyr Gly

846 TTT CAT TCC CAG CGG TCG GTC GAT AAA AAA ATC
 43 Lys Met Gly Leu Pro Arg Asp Ile Phe Phe Asp
```

Figure 7b (cont'd)

879 GAG ATA ACC GTT GGC CTC AAT CGG CGT TAA ACC
32 Leu Tyr Gly Asn Ala Glu Ile Pro Thr Leu Gly

912 CGC CAC CAG ATG GGC ATT AAA CGA GTA TCC CGG
21 Ala Val Leu His Ala Asn Phe Ser Tyr Gly Pro

945 CAG CAG GGG ATC ATT TTG CGC TTC AGC CAT
10 Leu Leu Pro Asp Asn Gln Ala Glu Ala Met

975 ACTTTTCATA CTCCCGCCAT TCAGAGAAGA AACCAATTGT

1015 CCATATTGCA TCAGACATTG CCGTCACTGC GTCTTTTACT

1055 GGCTCTTCTC GCTAACCAAA CCGGTAACCC CGCTTATTAA

1095 AAGCATTCTG TAACAAAGCG GGACCAAAGC CATGACAAAA

1135 ACGCGTAACA AAAGTGTCTA TAATCACGGC AGAAAAGTCC

1175 ACATTGATTA TTTGCACGGC GTCACACTTT GCTATGCCAT

BamHI

1215 AGCATTTTTA TCCATAAGAT TAGCGGATCC TACCTGACGC

1255 TTTTTATCGC AACTCTCTAC TGTTTCTCCA TACCCGTTTT

NheI EcoRI NcoI

1295 TTTGGGCTAG CAGGAGGAAT TCACC ATG GAT CCC GTA

1 Met Asp Pro Val

1332 ATC GTA GAA GAC ATA GAG CCA GGT ATT TAT TAC

5 Ile Val Glu Asp Ile Glu Pro Gly Ile Tyr Tyr

1365 GGA ATT TCG AAT GAG AAT TAC CAC GCG GGT CCC

16 Gly Ile Ser Asn Glu Asn Tyr His Ala Gly Pro

1398 GGT ATC AGT AAG TCT CAG CTC GAT GAC ATT GCT

Figure 7b (cont'd)

```
 27▶Gly Ile Ser Lys Ser Gln Leu Asp Asp Ile Ala
1431 GAT ACT CCG GCA CTA TAT TTG TGG CGT AAA AAT

 38▶Asp Thr Pro Ala Leu Tyr Leu Trp Arg Lys Asn
1464 GCC CCC GTG GAC ACC ACA AAG ACA AAA ACG CTC

 49▶Ala Pro Val Asp Thr Thr Lys Thr Lys Thr Leu
1497 GAT TTA GGA ACT GCT TTC CAC TGC CGG GTA CTT

 60▶Asp Leu Gly Thr Ala Phe His Cys Arg Val Leu
                EcoRI
1530 GAA CCG GAA GAA TTC AGT AAC CGC TTT ATC GTA

 71▶Glu Pro Glu Glu Phe Ser Asn Arg Phe Ile Val
1563 GCA CCT GAA TTT AAC CGC CGT ACA AAC GCC GGA

 82▶Ala Pro Glu Phe Asn Arg Arg Thr Asn Ala Gly
1596 AAA GAA GAA GAG AAA GCG TTT CTG ATG GAA TGC

 93▶Lys Glu Glu Glu Lys Ala Phe Leu Met Glu Cys
1629 GCA AGC ACA GGA AAA ACG GTT ATC ACT GCG GAA

104▶Ala Ser Thr Gly Lys Thr Val Ile Thr Ala Glu
1662 GAA GGC CGG AAA ATT GAA CTC ATG TAT CAA AGC

115▶Glu Gly Arg Lys Ile Glu Leu Met Tyr Gln Ser
```

Figure 7b (cont'd)

```
1695 GTT ATG GCT TTG CCG CTG GGG CAA TGG CTT GTT

 126▶Val Met Ala Leu Pro Leu Gly Gln Trp Leu Val
1728 GAA AGC GCC GGA CAC GCT GAA TCA TCA ATT TAC

 137▶Glu Ser Ala Gly His Ala Glu Ser Ser Ile Tyr
1761 TGG GAA GAT CCT GAA ACA GGA ATT TTG TGT CGG

 148▶Trp Glu Asp Pro Glu Thr Gly Ile Leu Cys Arg
1794 TGC CGT CCG GAC AAA ATT ATC CCT GAA TTT CAC

 159▶Cys Arg Pro Asp Lys Ile Ile Pro Glu Phe His
1827 TGG ATC ATG GAC GTG AAA ACT ACG GCG GAT ATT

 170▶Trp Ile Met Asp Val Lys Thr Thr Ala Asp Ile
1860 CAA CGA TTC AAA ACC GCT TAT TAC GAC TAC CGC

 181▶Gln Arg Phe Lys Thr Ala Tyr Tyr Asp Tyr Arg
1893 TAT CAC GTT CAG GAT GCA TTC TAC AGT GAC GGT

 192▶Tyr His Val Gln Asp Ala Phe Tyr Ser Asp Gly
1926 TAT GAA GCA CAG TTT GGA GTG CAG CCA ACT TTC

 203▶Tyr Glu Ala Gln Phe Gly Val Gln Pro Thr Phe
1959 GTT TTT CTG GTT GCC AGC ACA ACT ATT GAA TGC

 214▶Val Phe Leu Val Ala Ser Thr Thr Ile Glu Cys
1992 GGA CGT TAT CCG GTT GAA ATT TTC ATG ATG GGC
```

Figure 7b (cont'd)

225▶Gly Arg Tyr Pro Val Glu Ile Phe Met Met Gly
2025 GAA GAA GCA AAA CTG GCA GGT CAA CAG GAA TAT

236▶Glu Glu Ala Lys Leu Ala Gly Gln Gln Glu Tyr
2058 CAC CGC AAT CTG CGA ACC CTG TCT GAC TGC CTG

247▶His Arg Asn Leu Arg Thr Leu Ser Asp Cys Leu
BalI
2091 AAT ACC GAT GAA TGG CCA GCT ATT AAG ACA TTA

258▶Asn Thr Asp Glu Trp Pro Ala Ile Lys Thr Leu
2124 TCA CTG CCC CGC TGG GCT AAG GAA TAT GCAA

269▶Ser Leu Pro Arg Trp Ala Lys Glu Tyr AlaA
2155 ATG ACT AAG CAA CCA CCA ATC GCA AAA GCC GAT
1▶Met Thr Lys Gln Pro Pro Ile Ala Lys Ala Asp
279▶s nAs p• • •

2188 CTG CAA AAA ACT CAG GGA AAC CGT GCA CCA GCA
12▶Leu Gln Lys Thr Gln Gly Asn Arg Ala Pro Ala

2221 GCA GTT AAA AAT AGC GAC GTG ATT AGT TTT ATT
23▶Ala Val Lys Asn Ser Asp Val Ile Ser Phe Ile

2254 AAC CAG CCA TCA ATG AAA GAG CAA CTG GCA GCA
34▶Asn Gln Pro Ser Met Lys Glu Gln Leu Ala Ala
NdeI
2287 GCT CTT CCA CGC CAT ATG ACG GCT GAA CGT ATG
45▶Ala Leu Pro Arg His Met Thr Ala Glu Arg Met

Figure 7b (cont'd)

```
2320 ATC CGT ATC GCC ACC ACA GAA ATT CGT AAA GTT
  56▶Ile Arg Ile Ala Thr Thr Glu Ile Arg Lys Val

2353 CCG GCG TTA GGA AAC TGT GAC ACT ATG AGT TTT
  67▶Pro Ala Leu Gly Asn Cys Asp Thr Met Ser Phe

2386 GTC AGT GCG ATC GTA CAG TGT TCA CAG CTC GGA
  78▶Val Ser Ala Ile Val Gln Cys Ser Gln Leu Gly

2419 CTT GAG CCA GGT AGC GCC CTC GGT CAT GCA TAT
  89▶Leu Glu Pro Gly Ser Ala Leu Gly His Ala Tyr

2452 TTA CTG CCT TTT GGT AAT AAA AAC GAA AAG AGC
 100▶Leu Leu Pro Phe Gly Asn Lys Asn Glu Lys Ser

2485 GGT AAA AAG AAC GTT CAG CTA ATC ATT GGC TAT
 111▶Gly Lys Lys Asn Val Gln Leu Ile Ile Gly Tyr

2518 CGC GGC ATG ATT GAT CTG GCT CGC CGT TCT GGT
 122▶Arg Gly Met Ile Asp Leu Ala Arg Arg Ser Gly

2551 CAA ATC GCC AGC CTG TCA GCC CGT GTT GTC CGT
 133▶Gln Ile Ala Ser Leu Ser Ala Arg Val Val Arg

2584 GAA GGT GAC GAG TTT AGC TTC GAA TTT GGC CTT
 144▶Glu Gly Asp Glu Phe Ser Phe Glu Phe Gly Leu

2617 GAT GAA AAG TTA ATA CAC CGC CCG GGA GAA AAC
 155▶Asp Glu Lys Leu Ile His Arg Pro Gly Glu Asn

2650 GAA GAT GCC CCG GTT ACC CAC GTC TAT GCT GTC
 166▶Glu Asp Ala Pro Val Thr His Val Tyr Ala Val

2683 GCA AGA CTG AAA GAC GGA GGT ACT CAG TTT GAA
 177▶Ala Arg Leu Lys Asp Gly Gly Thr Gln Phe Glu

2716 GTT ATG ACG CGC AAA CAG ATT GAG CTG GTG CGC
 188▶Val Met Thr Arg Lys Gln Ile Glu Leu Val Arg
```

Figure 7b (cont'd)

```
2749  AGC CTG AGT AAA GCT GGT AAT AAC GGG CCG TGG
 199▶ Ser Leu Ser Lys Ala Gly Asn Asn Gly Pro Trp

2782  GTA ACT CAC TGG GAA GAA ATG GCA AAG AAA ACG
 210▶ Val Thr His Trp Glu Glu Met Ala Lys Lys Thr

2815  GCT ATT CGT CGC CTG TTC AAA TAT TTG CCC GTA
 221▶ Ala Ile Arg Arg Leu Phe Lys Tyr Leu Pro Val

2848  TCA ATT GAG ATC CAG CGT GCA GTA TCA ATG GAT
 232▶ Ser Ile Glu Ile Gln Arg Ala Val Ser Met Asp
                                          PstI
2881  GAA AAG GAA CCA CTG ACA ATC GAT CCT GCA GAT
 243▶ Glu Lys Glu Pro Leu Thr Ile Asp Pro Ala Asp

2914  TCC TCT GTA TTA ACC GGG GAA TAC AGT GTA ATC
 254▶ Ser Ser Val Leu Thr Gly Glu Tyr Ser Val Ile
                              BglII    HindIII
2947  GAT AAT TCA GAG GAA TAG ATCTAAGCTT
 265▶ Asp Asn Ser Glu Glu •••

2975  GGCTGTTTTG GCGGATGAGA GAAGATTTTC AGCCTGATAC

3015  AGATTAAATC AGAACGCAGA AGCGGTCTGA TAAAACAGAA

3055  TTTGCCTGGC GGCAGTAGCG CGGTGGTCCC ACCTGACCCC

3095  ATGCCGAACT CAGAAGTGAA ACGCCGTAGC GCCGATGGTA

3135  GTGTGGGGTC TCCCCATGCG AGAGTAGGGA ACTGCCAGGC

3175  ATCAAATAAA ACGAAAGGCT CAGTCGAAAG ACTGGGCCTT

3215  TCGTTTTATC TGTTGTTTGT CGGTGAACGC TCTCCTGAGT

3255  AGGACAAATC CGCCGGGAGC GGATTTGAAC GTTGCGAAGC

3295  AACGGCCCGG AGGGTGGCGG GCAGGACGCC CGCCATAAAC

3335  TGCCAGGCAT CAAATTAAGC AGAAGGCCAT CCTGACGGAT
```

Figure 7b (cont'd)

```
3375  GGCCTTTTTG CGTTTCTACA AACTCTTTTG TTTATTTTTC

3415  TAAATACATT CAAATATGTA TCCGCTCATG AGACAATAAC

3455  CCTGATAAAT GCTTCAATAA TATTGAAAAA GGAAGAGT AT
                                              1▶Me

3495  G AGT ATT CAA CAT TTC CGT GTC GCC CTT ATT
    1▶t Ser Ile Gln His Phe Arg Val Ala Leu Ile

3526  CCC TTT TTT GCG GCA TTT TGC CTT CCT GTT TTT
   12▶Pro Phe Phe Ala Ala Phe Cys Leu Pro Val Phe

3559  GCT CAC CCA GAA ACG CTG GTG AAA GTA AAA GAT
   23▶Ala His Pro Glu Thr Leu Val Lys Val Lys Asp

3592  GCT GAA GAT CAG TTG GGT GCA CGA GTG GGT TAC
   34▶Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr

3625  ATC GAA CTG GAT CTC AAC AGC GGT AAG ATC CTT
   45▶Ile Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu

3658  GAG AGT TTT CGC CCC GAA GAA CGT TTT CCA ATG
   56▶Glu Ser Phe Arg Pro Glu Glu Arg Phe Pro Met

3691  ATG AGC ACT TTT AAA GTT CTG CTA TGT GGC GCG
   67▶Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala

3724  GTA TTA TCC CGT GTT GAC GCC GGG CAA GAG CAA
   78▶Val Leu Ser Arg Val Asp Ala Gly Gln Glu Gln

3757  CTC GGT CGC CGC ATA CAC TAT TCT CAG AAT GAC
   89▶Leu Gly Arg Arg Ile His Tyr Ser Gln Asn Asp
                  ScaI
3790  TTG GTT GAG TAC TCA CCA GTC ACA GAA AAG CAT
  100▶Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His

3823  CTT ACG GAT GGC ATG ACA GTA AGA GAA TTA TGC
  111▶Leu Thr Asp Gly Met Thr Val Arg Glu Leu Cys
```

Figure 7b (cont'd)

```
3856 AGT GCT GCC ATA ACC ATG AGT GAT AAC ACT GCG
 122▶Ser Ala Ala Ile Thr Met Ser Asp Asn Thr Ala

3889 GCC AAC TTA CTT CTG ACA ACG ATC GGA GGA CCG
 133▶Ala Asn Leu Leu Leu Thr Thr Ile Gly Gly Pro

3922 AAG GAG CTA ACC GCT TTT TTG CAC AAC ATG GGG
 144▶Lys Glu Leu Thr Ala Phe Leu His Asn Met Gly

3955 GAT CAT GTA ACT CGC CTT GAT CGT TGG GAA CCG
 155▶Asp His Val Thr Arg Leu Asp Arg Trp Glu Pro

3988 GAG CTG AAT GAA GCC ATA CCA AAC GAC GAG CGT
 166▶Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg

4021 GAC ACC ACG ATG CCT GTA GCA ATG GCA ACA ACG
 177▶Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr

4054 TTG CGC AAA CTA TTA ACT GGC GAA CTA CTT ACT
 188▶Leu Arg Lys Leu Leu Thr Gly Glu Leu Leu Thr

4087 CTA GCT TCC CGG CAA CAA TTA ATA GAC TGG ATG
 199▶Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp Met

4120 GAG GCG GAT AAA GTT GCA GGA CCA CTT CTG CGC
 210▶Glu Ala Asp Lys Val Ala Gly Pro Leu Leu Arg

4153 TCG GCC CTT CCG GCT GGC TGG TTT ATT GCT GAT
 221▶Ser Ala Leu Pro Ala Gly Trp Phe Ile Ala Asp

4186 AAA TCT GGA GCC GGT GAG CGT GGG TCT CGC GGT
 232▶Lys Ser Gly Ala Gly Glu Arg Gly Ser Arg Gly

4219 ATC ATT GCA GCA CTG GGG CCA GAT GGT AAG CCC
 243▶Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro

4252 TCC CGT ATC GTA GTT ATC TAC ACG ACG GGG AGT
 254▶Ser Arg Ile Val Val Ile Tyr Thr Thr Gly Ser
```

Figure 7b (cont'd)

```
4285  CAG GCA ACT ATG GAT GAA CGA AAT AGA CAG ATC
 265▶ Gln Ala Thr Met Asp Glu Arg Asn Arg Gln Ile

4318  GCT GAG ATA GGT GCC TCA CTG ATT AAG CAT TGG
 276▶ Ala Glu Ile Gly Ala Ser Leu Ile Lys His Trp

4351  TAA CTGTCAGACC AAGTTTACTC ATATATACTT
 287▶ • • •

4384  TAGATTGATT TACGCGCCCT GTAGCGGCGC ATTAAGCGCG
4424  GCGGGTGTGG TGGTTACGCG CAGCGTGACC GCTACACTTG
4464  CCAGCGCCCT AGCGCCCGCT CCTTTCGCTT TCTTCCCTTC
4504  CTTTCTCGCC ACGTTCGCCG GCTTTCCCCG TCAAGCTCTA
4544  AATCGGGGGC TCCCTTTAGG GTTCCGATTT AGTGCTTTAC
4584  GGCACCTCGA CCCCAAAAAA CTTGATTTGG GTGATGGTTC
4624  ACGTAGTGGG CCATCGCCCT GATAGACGGT TTTTCGCCCT
4664  TTGACGTTGG AGTCCACGTT CTTTAATAGT GGACTCTTGT
4704  TCCAAACTTG AACAACACTC AACCCTATCT CGGGCTATTC
4744  TTTTGATTTA TAAGGGATTT TGCCGATTTC GGCCTATTGG
4784  TTAAAAAATG AGCTGATTTA ACAAAAATTT AACGCGAATT
4824  TTAACAAAAT ATTAACGTTT ACAATTTAAA AGGATCTAGG
4864  TGAAGATCCT TTTTGATAAT CTCATGACCA AAATCCCTTA
4904  ACGTGAGTTT TCGTTCCACT GAGCGTCAGA CCCCGTAGAA
4944  AAGATCAAAG GATCTTCTTG AGATCCTTTT TTTCTGCGCG
4984  TAATCTGCTG CTTGCAAACA AAAAAACCAC CGCTACCAGC
5024  GGTGGTTTGT TTGCCGGATC AAGAGCTACC AACTCTTTTT
```

Figure 7b (cont'd)

5064 CCGAAGGTAA CTGGCTTCAG CAGAGCGCAG ATACCAAATA

5104 CTGTCCTTCT AGTGTAGCCG TAGTTAGGCC ACCACTTCAA

5144 GAACTCTGTA GCACCGCCTA CATACCTCGC TCTGCTAATC

5184 CTGTTACCAG TGGCTGCTGC CAGTGGCGAT AAGTCGTGTC

5224 TTACCGGGTT GGACTCAAGA CGATAGTTAC CGGATAAGGC

5264 GCAGCGGTCG GGCTGAACGG GGGGTTCGTG CACACAGCCC

5304 AGCTTGGAGC GAACGACCTA CACCGAACTG AGATACCTAC

5344 AGCGTGAGCT ATGAGAAAGC GCCACGCTTC CCGAAGGGAG

5384 AAAGGCGGAC AGGTATCCGG TAAGCGGCAG GGTCGGAACA

5424 GGAGAGCGCA CGAGGGAGCT TCCAGGGGGA AACGCCTGGT

5464 ATCTTTATAG TCCTGTCGGG TTTCGCCACC TCTGACTTGA

5504 GCGTCGATTT TTGTGATGCT CGTCAGGGGG GCGGAGCCTA

5544 TGGAAAAACG CCAGCAACGC GGCCTTTTTA CGGTTCCTGG

5584 CCTTTTGCTG GCCTTTTGCT CACATGTTCT TTCCTGCGTT

5624 ATCCCCTGAT TCTGTGGATA ACCGTATTAC CGCCTTTGAG

5664 TGAGCTGATA CCGCTCGCCG CAGCCGAACG ACCGAGCGCA

5704 GCGAGTCAGT GAGCGAGGAA GCGGAAGAGC GCCTGATGCG

5744 GTATTTTCTC CTTACGCATC TGTGCGGTAT TTCACACCGC

5784 ATAGGGTCAT GGCTGCGCCC CGACACCCGC CAACACCCGC

5824 TGACGCGCCC TGACGGGCTT GTCTGCTCCC GGCATCCGCT

5864 TACAGACAAG CTGTGACCGT CTCCGGGAGC TGCATGTGTC

5904 AGAGGTTTTC ACCGTCATCA CCGAAACGCG CGAGGCAGCA

5944 AGGAGATGGC GCCCAACAGT CCCCCGGCCA CGGGGCCTGC

Figure 7b (cont'd)

```
5984  CACCATACCC ACGCCGAAAC AAGCGCTCAT GAGCCCGAAG
6024  TGGCGAGCCC GATCTTCCCC ATCGGTGATG TCGGCGATAT
6064  AGGCGCCAGC AACCGCACCT GTGGCGCCGG TGATGCCGGC
6104  CACGATGCGT CCGGCGTAGA GGATCTGCTC ATGTTTGACA
6144  GCTTATC
```

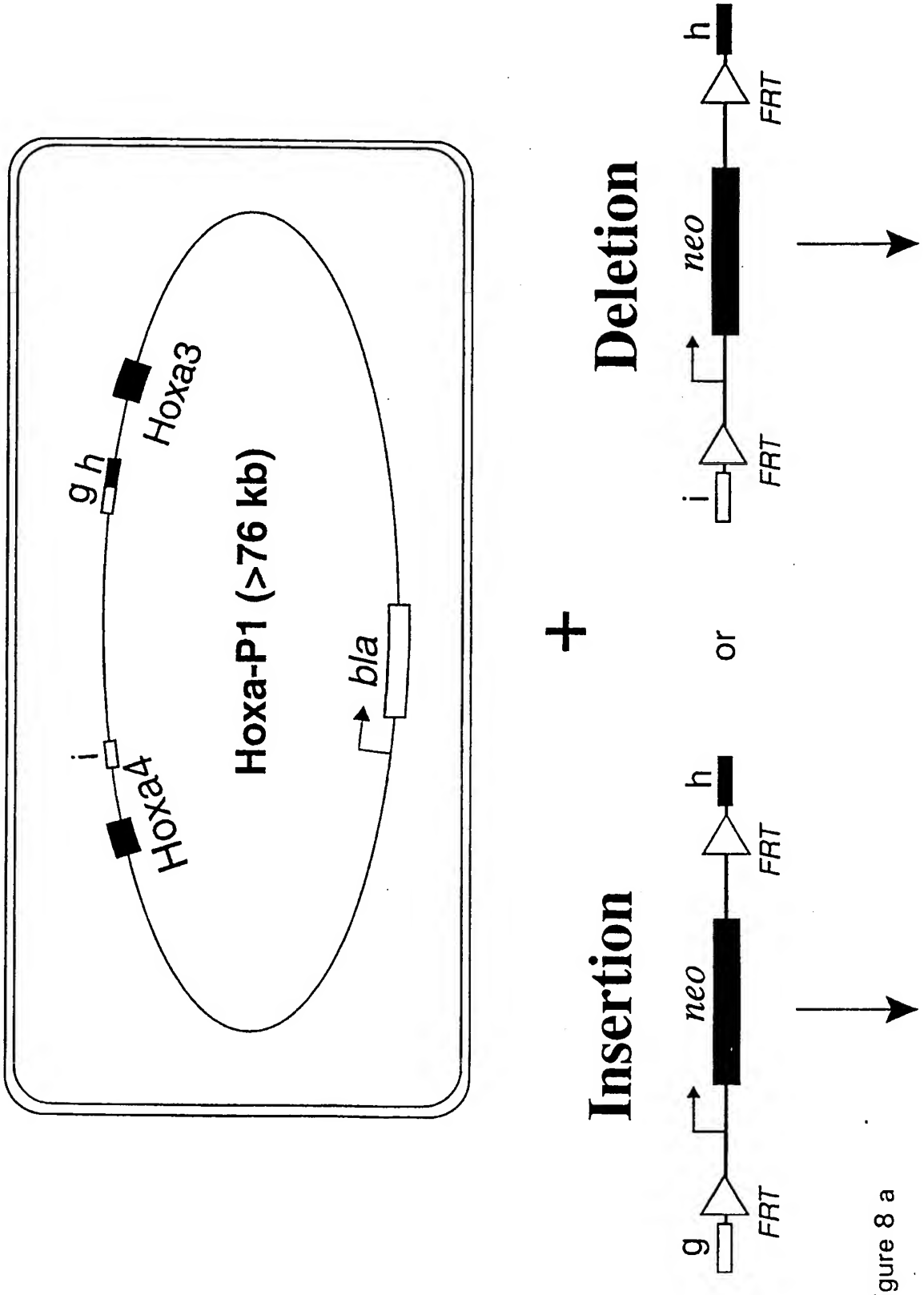

Hoxa-P1 (>76 kb)
Hoxa3
Hoxa4
g
h
i
bla
+
Deletion
or
Insertion
neo
FRT
FRT
h
i
FRT
neo
h
FRT
g
FRT

Figure 8 a

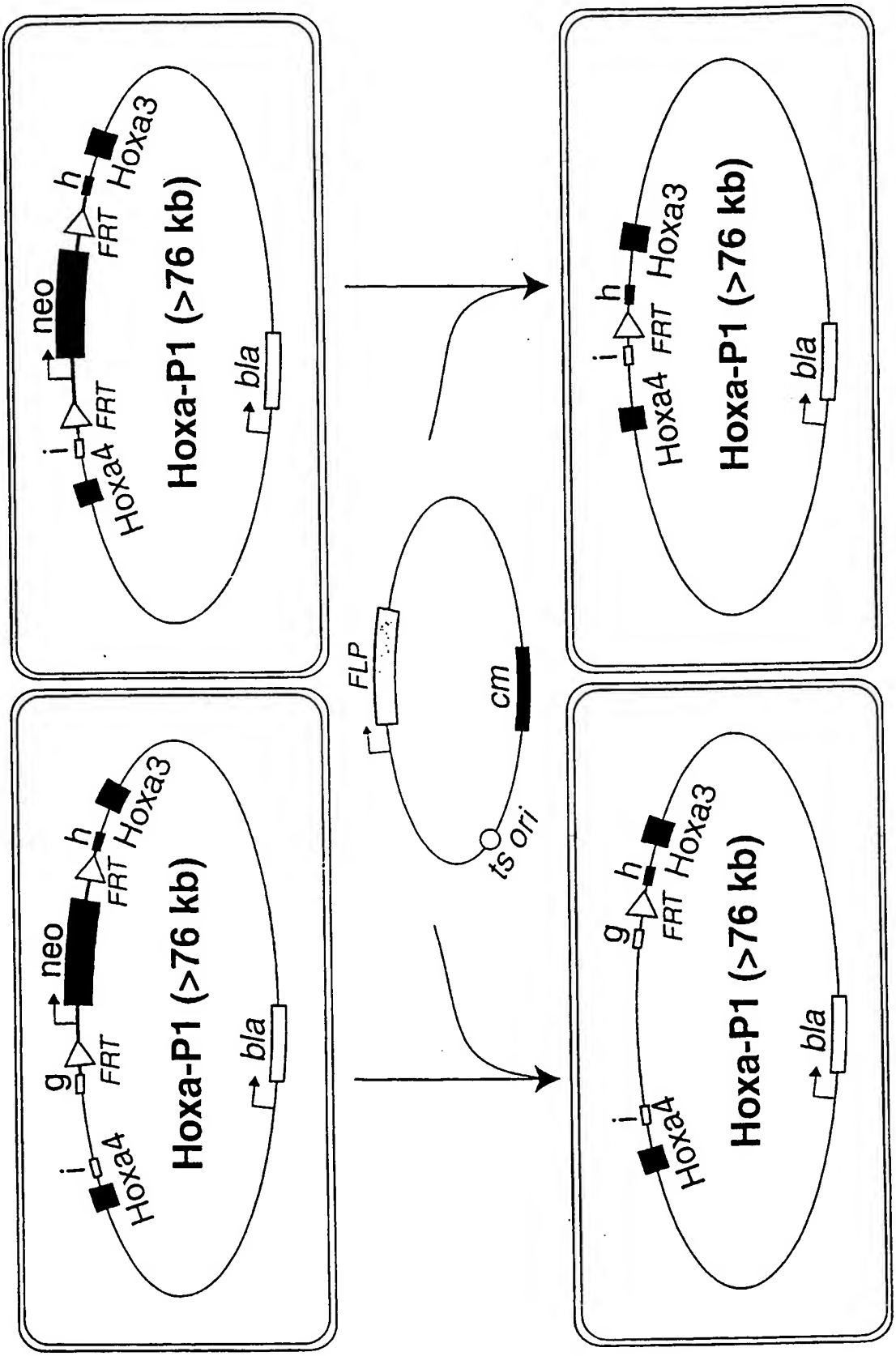


Figure 8 a (continuing)

Figure 8 b

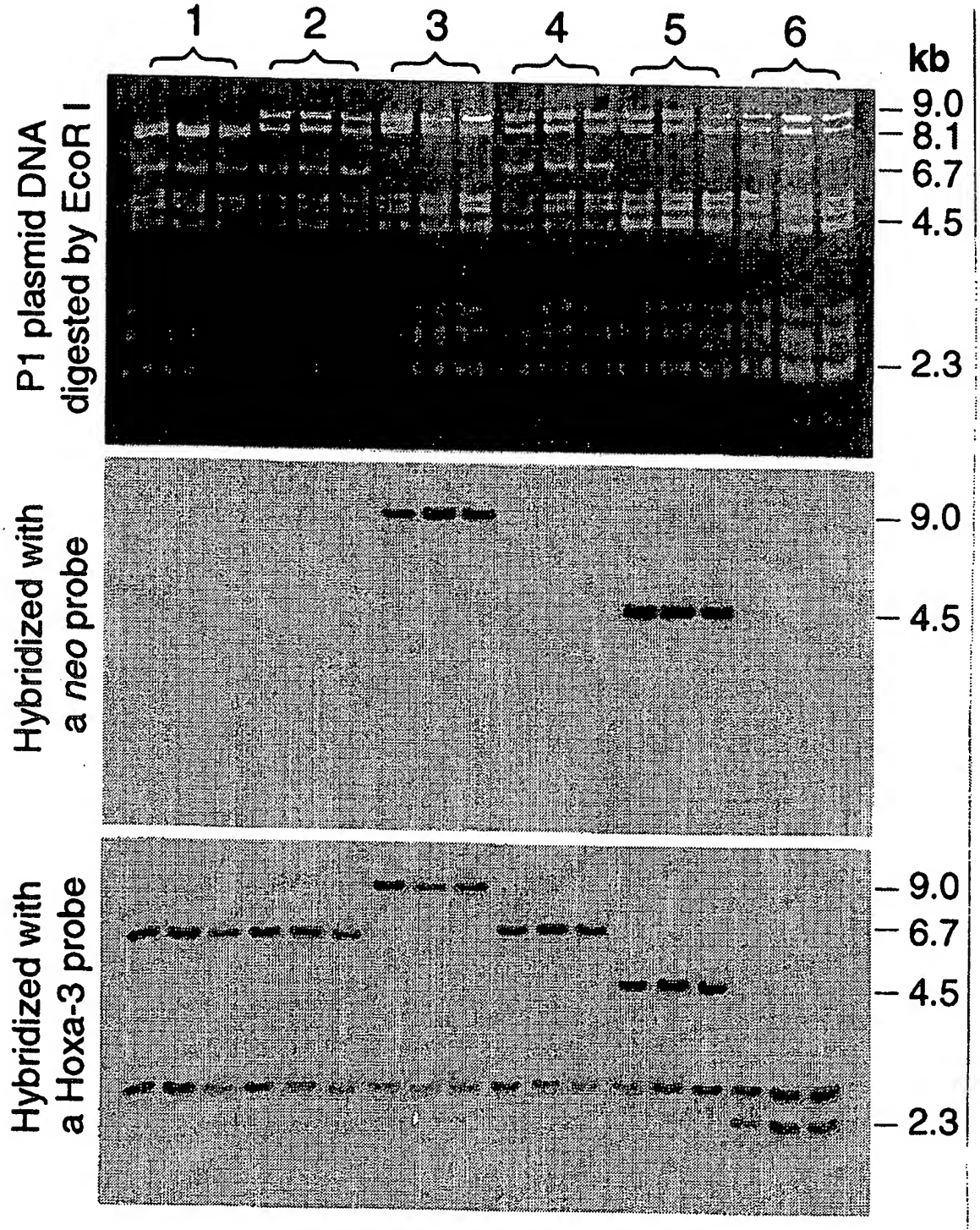

1
2
3
4
5
6
kb
9.0
8.1
6.7
4.5
2.3
P1 plasmid DNA digested by EcoR I
Hybridized with a neo probe
9.0
4.5
Hybridized with a Hoxa-3 probe
9.0
6.7
4.5
2.3

Figure 9a

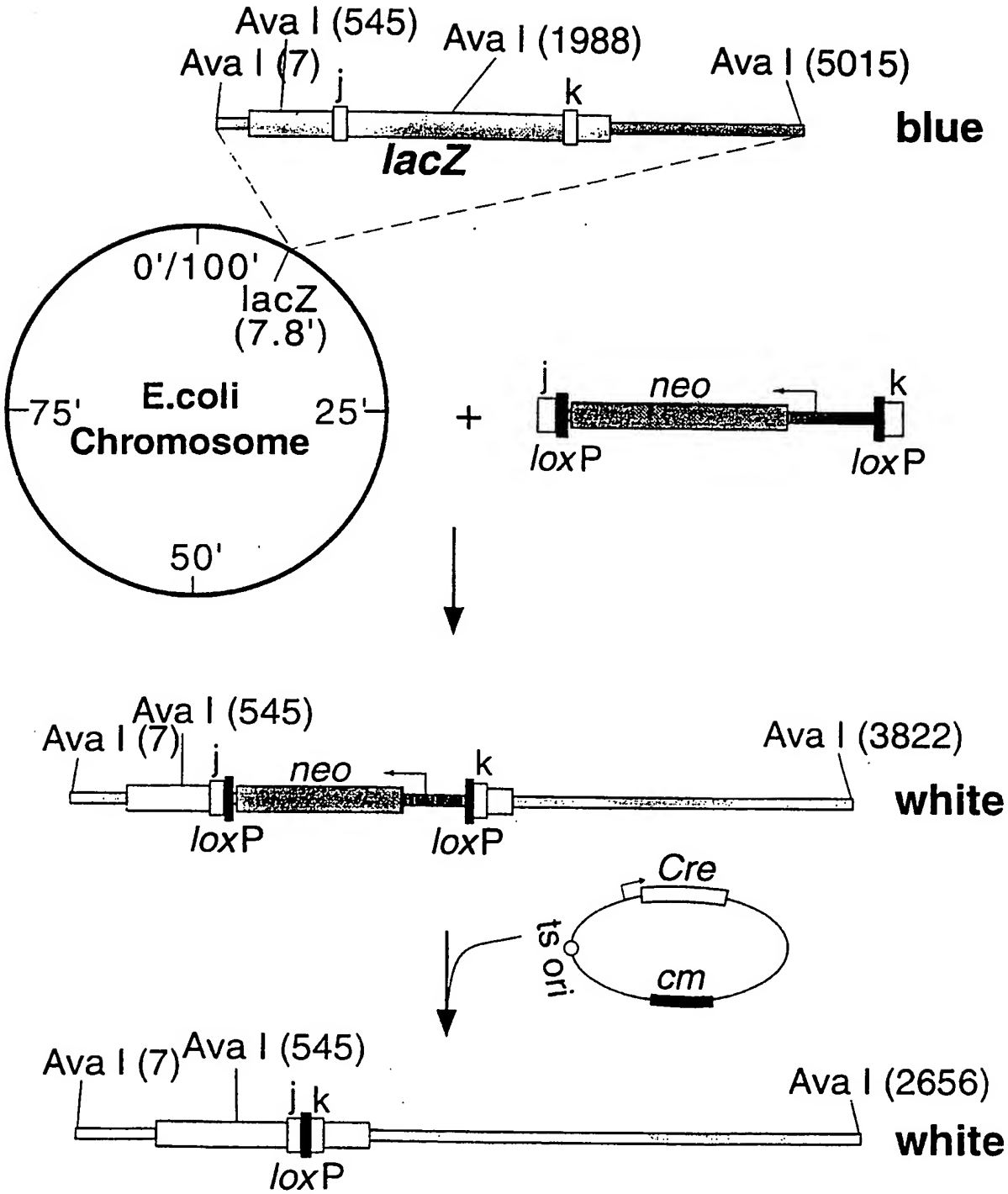

Ava I (545)
Ava I (7)
Ava I (1988)
Ava I (5015)
j
k
lacZ
blue
0'/100'
lacZ
(7.8')
E.coli
Chromosome
75'
25'
50'
+
j
neo
k
loxP
loxP
Ava I (545)
Ava I (7)
Ava I (3822)
j
neo
k
loxP
loxP
white
Cre
ts ori
cm
Ava I (7)
Ava I (545)
Ava I (2656)
j k
loxP
white

Figure 9

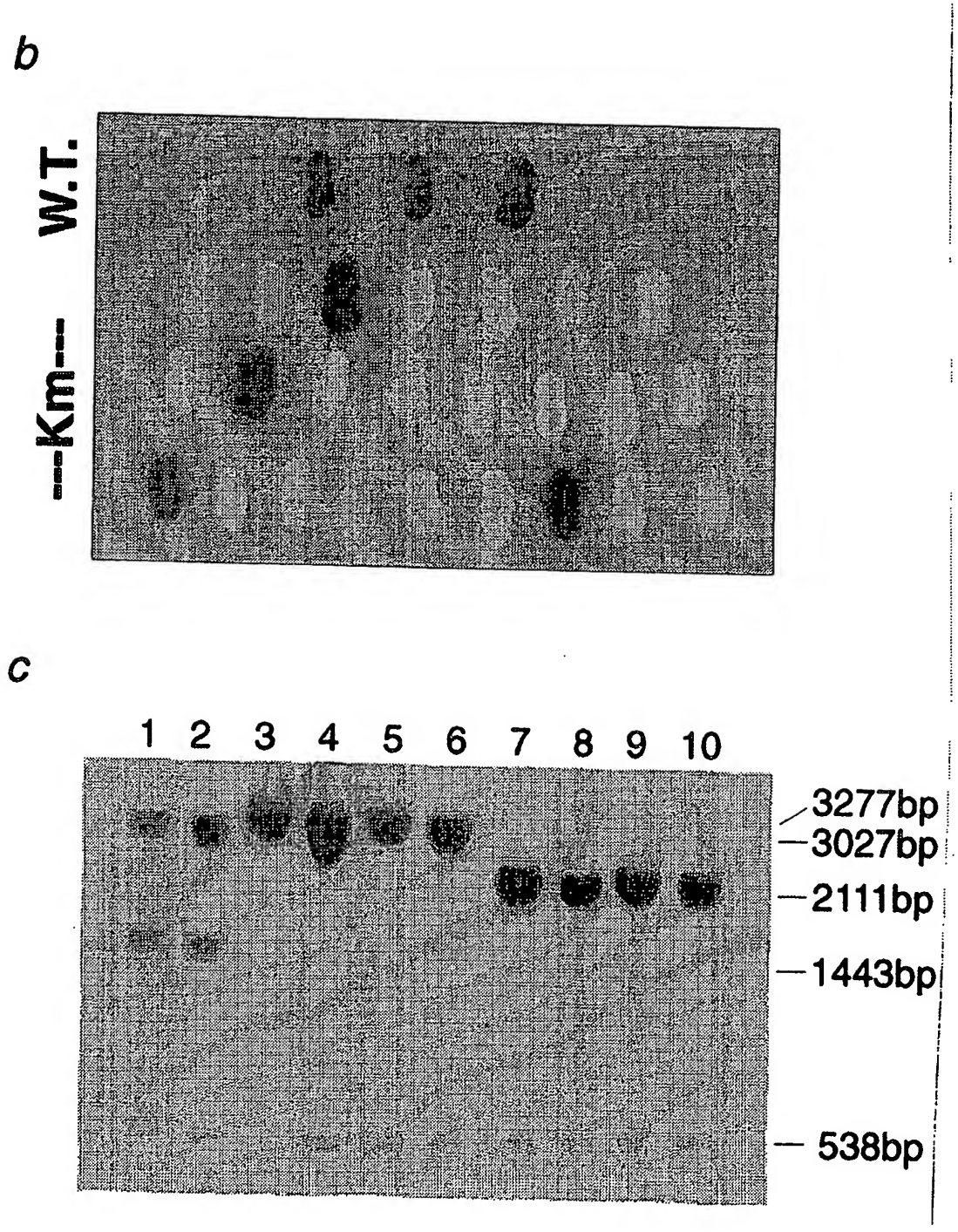


Figure 10a

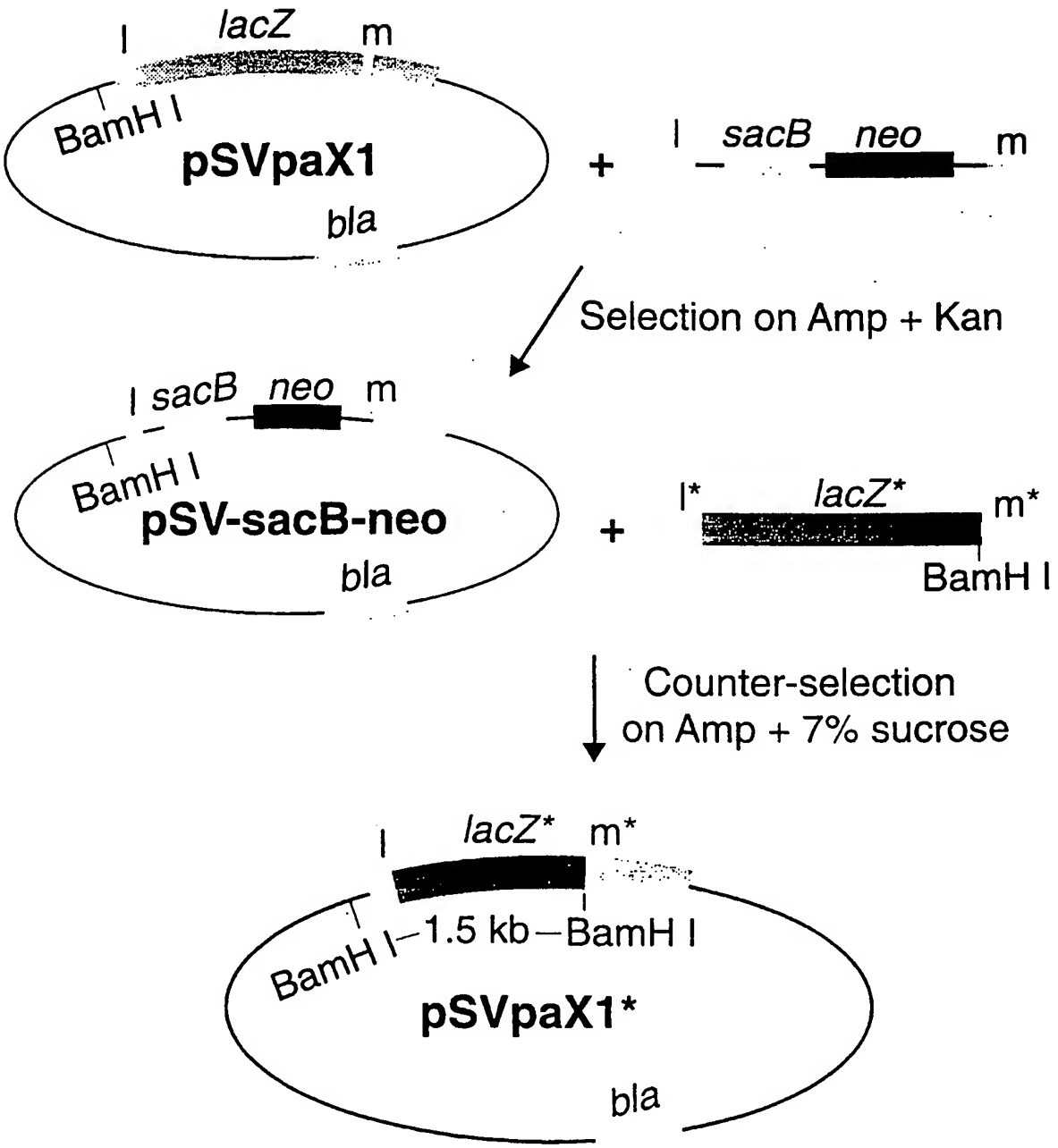

lacZ
m
BamH I
pSVpaX1
bla
+
sacB
neo
m
Selection on Amp + Kan
sacB
neo
m
BamH I
pSV-sacB-neo
bla
+
lacZ*
m*
BamH I
Counter-selection
on Amp + 7% sucrose
lacZ*
m*
BamH I
1.5 kb
BamH I
pSVpaX1*
bla

Figure 10

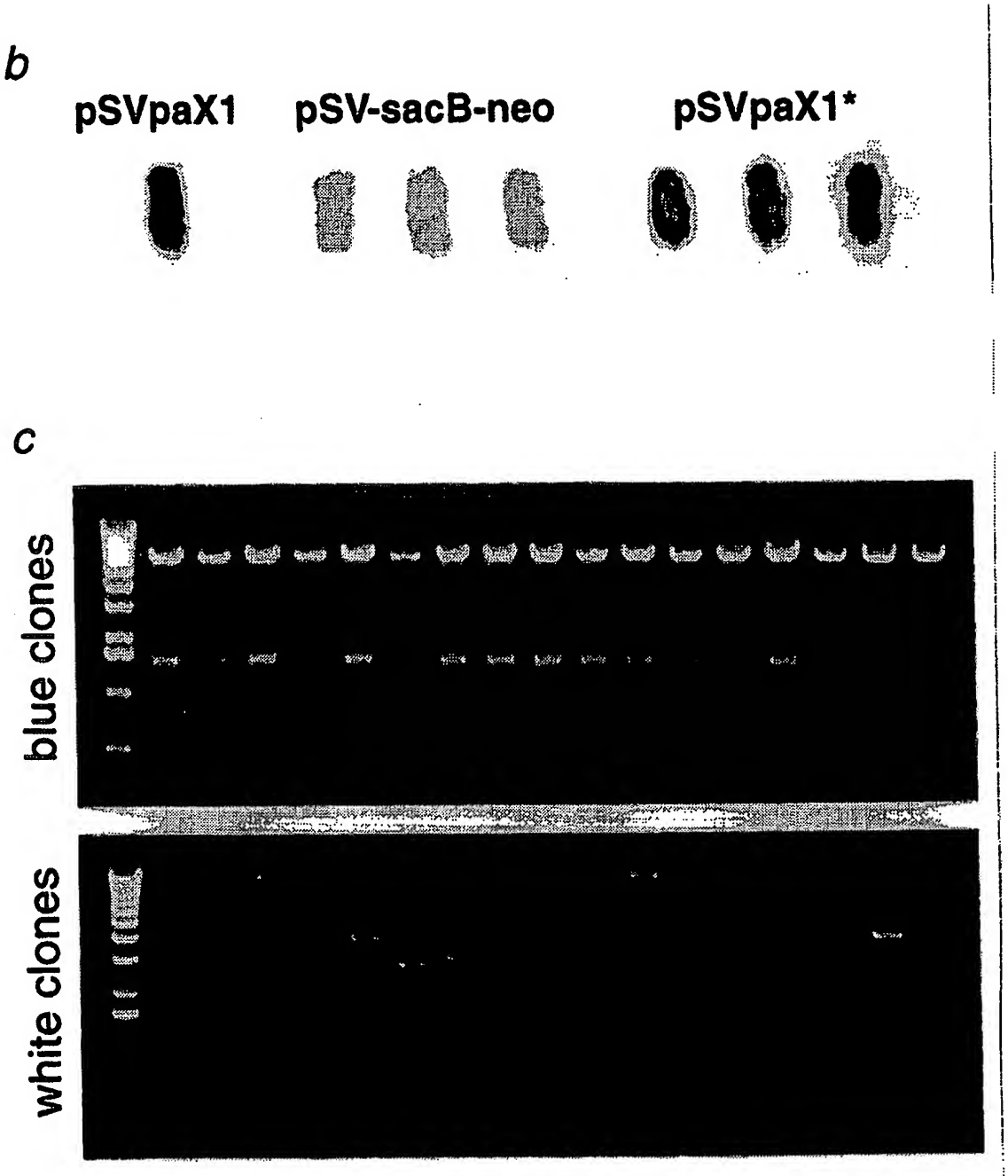


Figure 11a

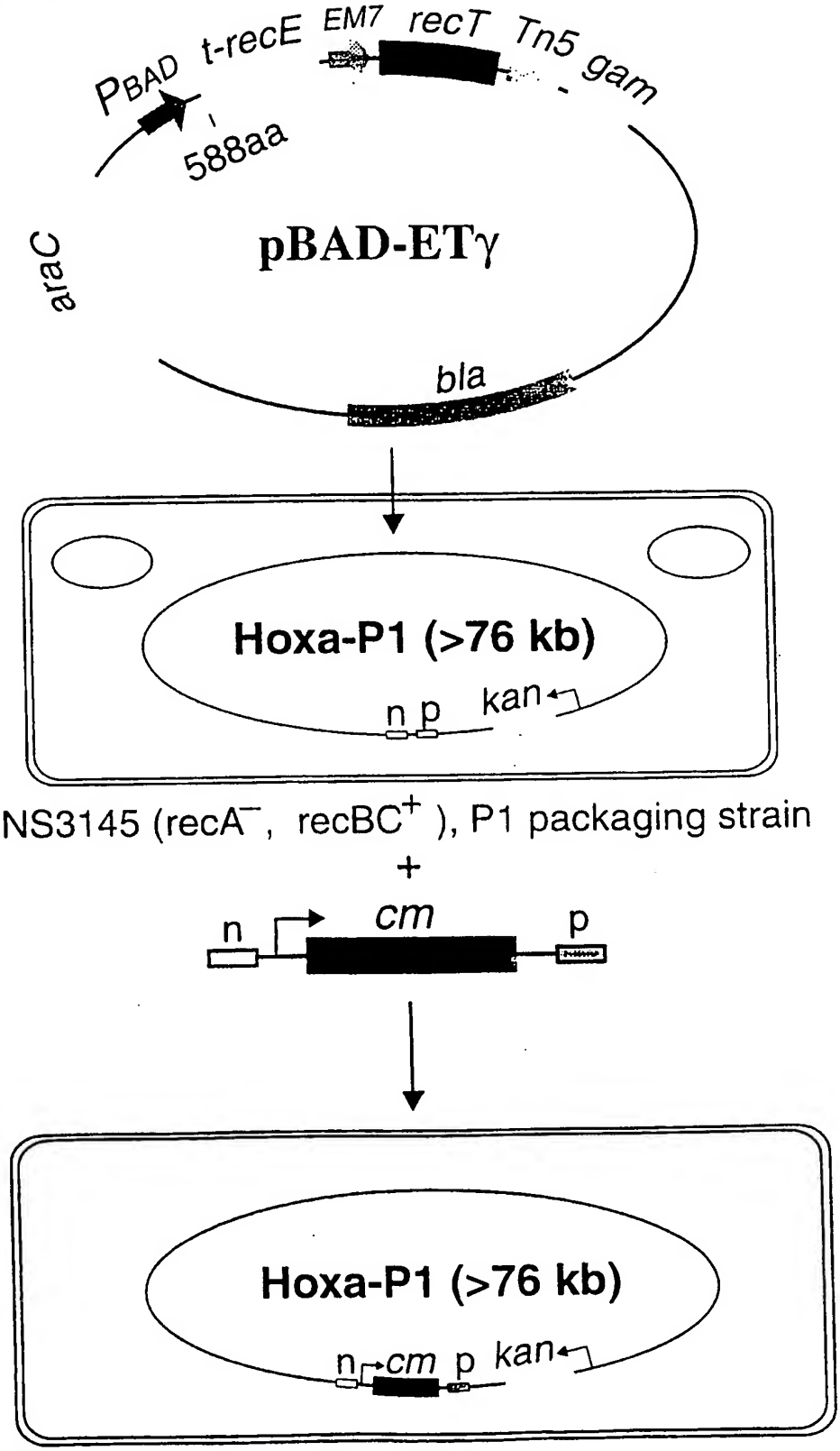

PBAD
t-recE
EM7
recT
Tn5
gam
588aa
araC
pBAD-ETγ
bla
Hoxa-P1 (>76 kb)
n p
kan
NS3145 (recA⁻, recBC⁺), P1 packaging strain
+
n
cm
p
Hoxa-P1 (>76 kb)
n cm p kan

Figure 11 b

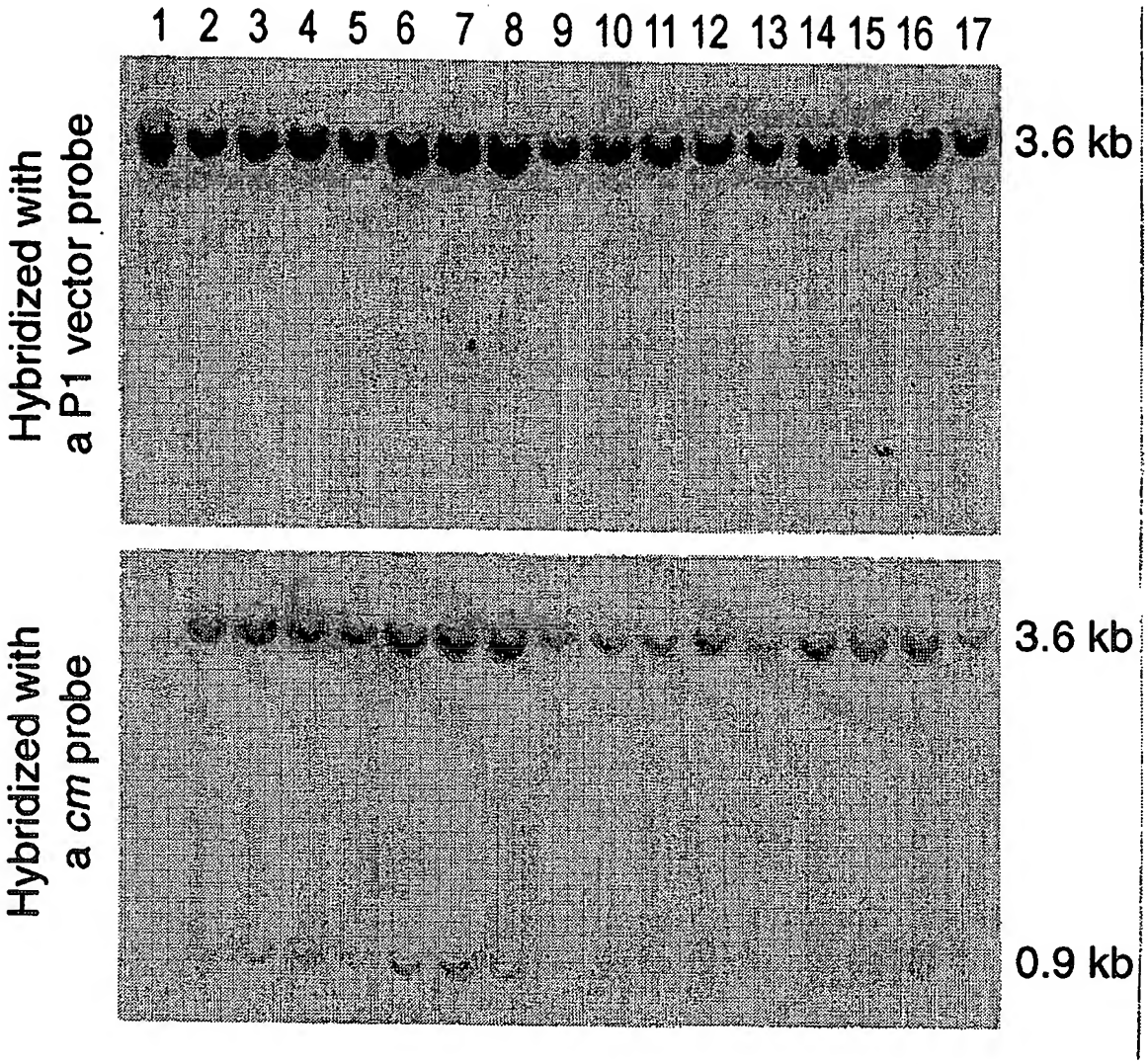
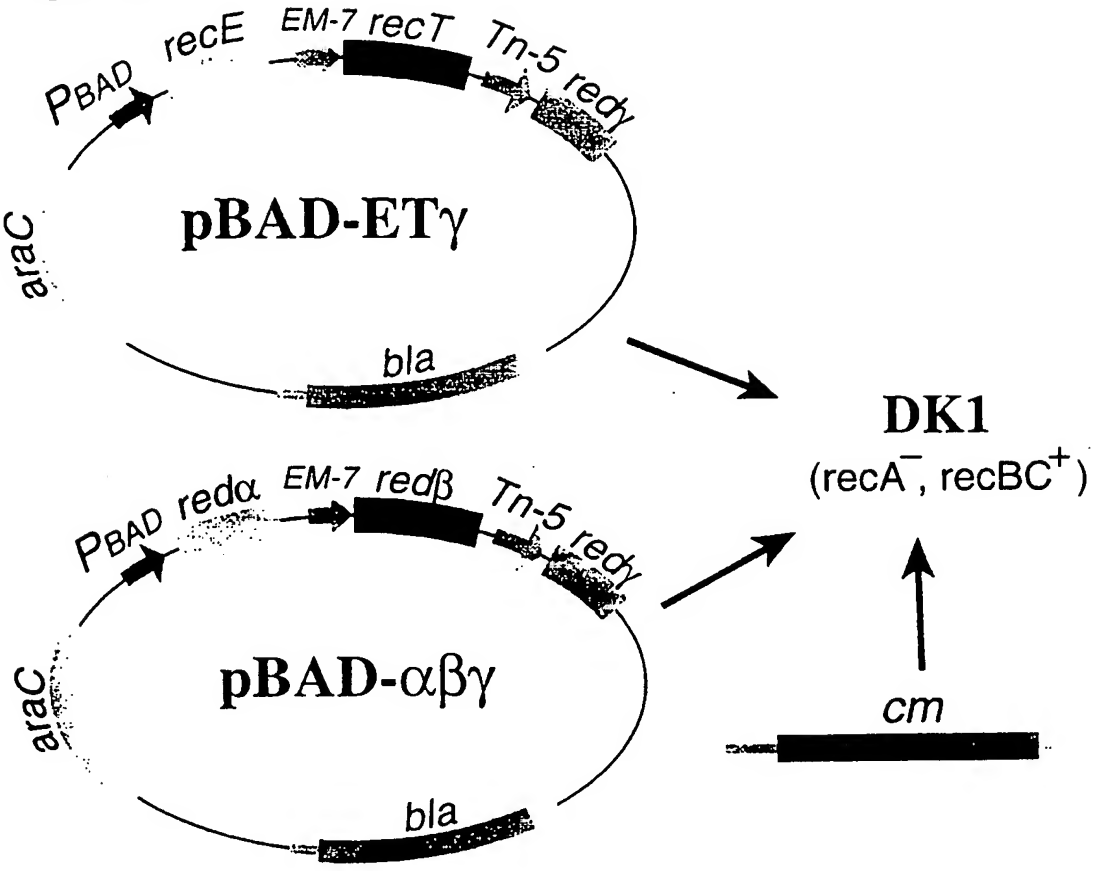

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17
Hybridized with a P1 vector probe
3.6 kb
Hybridized with a cm probe
3.6 kb
0.9 kb

Figure 12


PBAD
recE
EM-7 recT
Tn-5 redγ
pBAD-ETγ
araC
bla
DK1
(recA⁻, recBC⁺)
PBAD redα
EM-7 redβ
Tn-5 redγ
pBAD-αβγ
araC
bla
cm

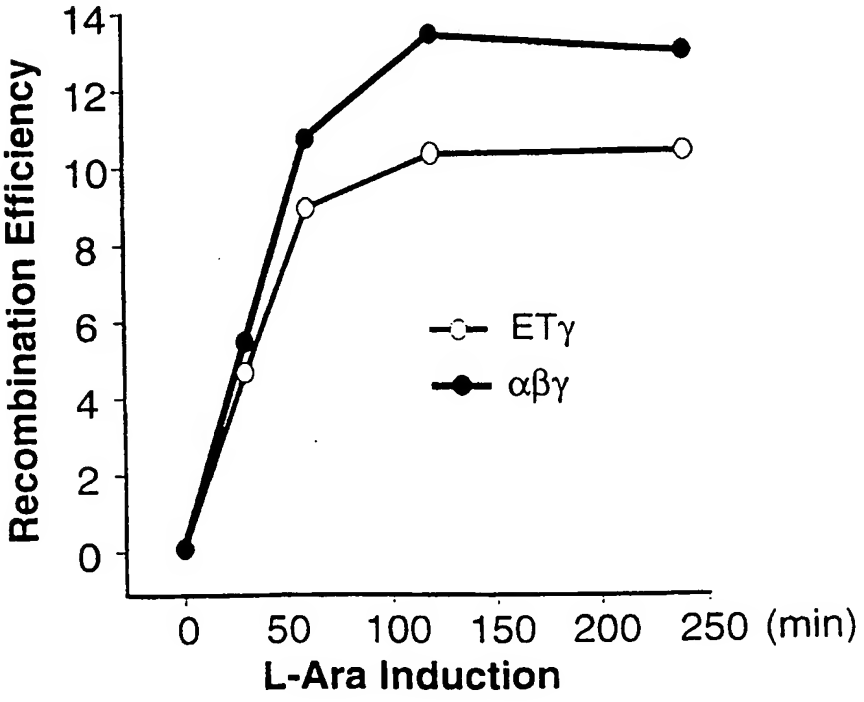

Recombination Efficiency
14
12
10
8
6
4
2
0
ETγ
αβγ
0
50
100
150
200
250 (min)
L-Ara Induction

Figure 13 a

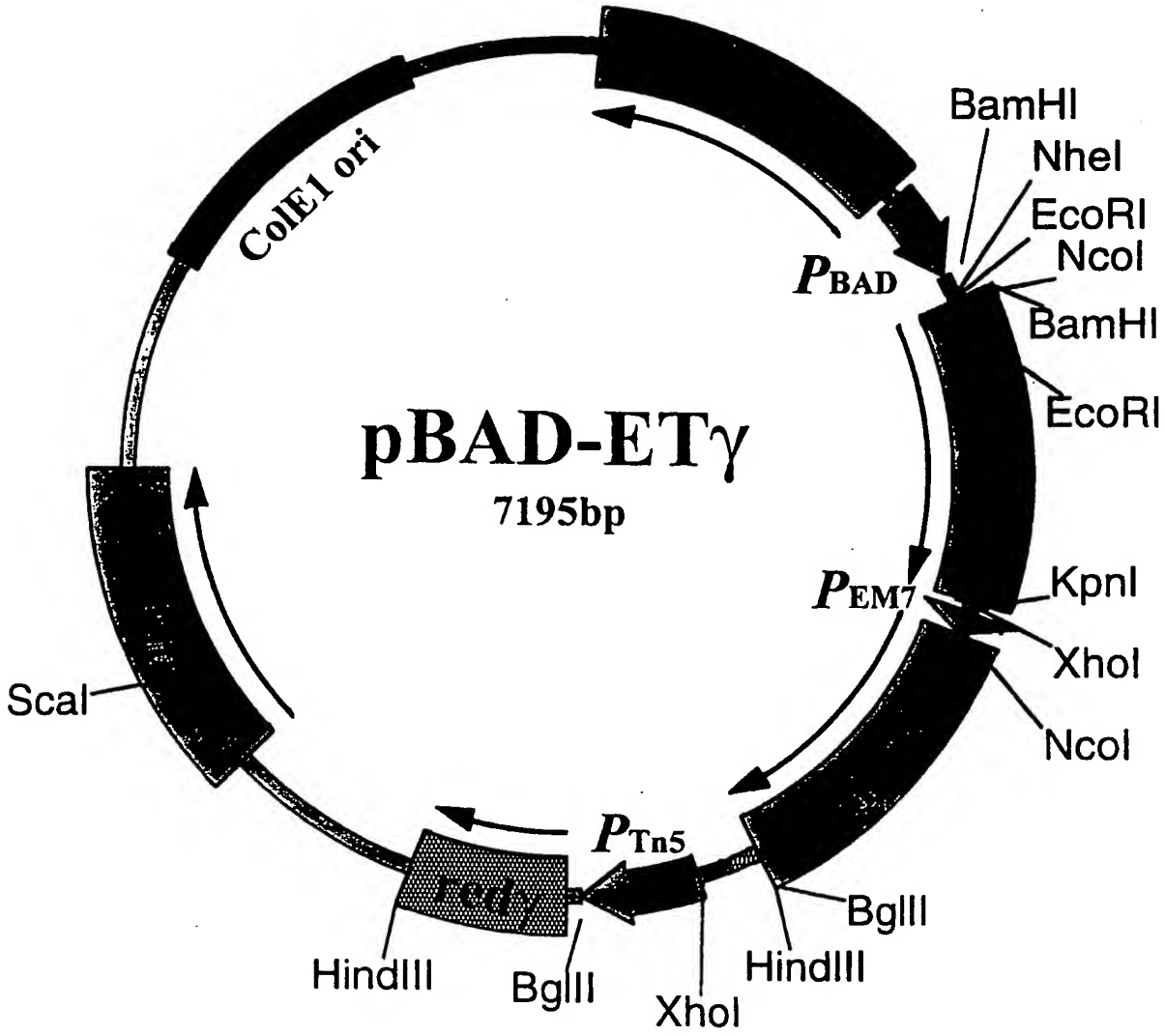

pBAD-ETγ
7195bp
ColE1 ori
BamHI
NheI
EcoRI
NcoI
BamHI
EcoRI
KpnI
XhoI
NcoI
BglII
HindIII
XhoI
BglII
HindIII
ScaI
P_{BAD}
P_{EM7}
P_{Tn5}
tetA

Figure 13b

```
  1 ATCGATGCATAATGTGCCTGTCAAATGGACGAAGCAGGG

 40 ATTCTGCAAACCCTATGCTACTCCGTCAAGCCGTCAATT

 79 GTCTGATTCGTTACCAA TTA TGA CAA CTT GAC
                   293◀••• Ser Leu Lys Val

111 GGC TAC ATC ATT CAC TTT TTC TTC ACA ACC
288◀Ala Val Asp Asn Val Lys Glu Glu Cys Gly

141 GGC ACG GAA CTC GCT CGG GCT GGC CCC GGT
278◀Ala Arg Phe Glu Ser Pro Ser Ala Gly Thr

171 GCA TTT TTT AAA TAC CCG CGA GAA ATA GAG
268◀Cys Lys Lys Phe Val Arg Ser Phe Tyr Leu

201 TTG ATC GTC AAA ACC AAC ATT GCG ACC GAC
258◀Gln Asp Asp Phe Gly Val Asn Arg Gly Val

231 GGT GGC GAT AGG CAT CCG GGT GGT GCT CAA
248◀Thr Ala Ile Pro Met Arg Thr Thr Ser Leu

261 AAG CAG CTT CGC CTG GCT GAT ACG TTG GTC
238◀Leu Leu Lys Ala Gln Ser Ile Arg Gln Asp

291 CTC GCG CCA GCT TAA GAC GCT AAT CCC TAA
228◀Glu Arg Trp Ser Leu Val Ser Ile Gly Leu

321 CTG CTG GCG GAA AAG ATG TGA CAG ACG CGA
218◀Gln Gln Arg Phe Leu His Ser Leu Arg Ser

351 CGG CGA CAA GCA AAC ATG CTG TGC GAC GCT
208◀Pro Ser Leu Cys Val His Gln Ala Val Ser

381 GGC GAT ATC AAA ATT GCT GTC TGC CAG GTG
198◀Ala Ile Asp Phe Asn Ser Asp Ala Leu His

411 ATC GCT GAT GTA CTG ACA AGC CTC GCG TAC
```

Figure 13b (cont'd)

188 Asp Ser Ile Tyr Gln Cys Ala Glu Arg Val

441 CCG ATT ATC CAT CGG TGG ATG GAG CGA CTC
178 Arg Asn Asp Met Pro Pro His Leu Ser Glu

471 GTT AAT CGC TTC CAT GCG CCG CAG TAA CAA
168 Asn Ile Ala Glu Met Arg Arg Leu Leu Leu

501 TTG CTC AAG CAG ATT TAT CGC CAG CAG CTC
158 Gln Glu Leu Leu Asn Ile Ala Leu Leu Glu

531 CGA ATA GCG CCC TTC CCC TTG CCC GGC GTT
148 Ser Tyr Arg Gly Glu Gly Gln Gly Ala Asn

561 AAT GAT TTG CCC AAA CAG GTC GCT GAA ATG
138 Ile Ile Gln Gly Phe Leu Asp Ser Phe His

591 CGG CTG GTG CGC TTC ATC CGG GCG AAA GAA
128 Pro Gln His Ala Glu Asp Pro Arg Phe Phe

621 CCC CGT ATT GGC AAA TAT TGA CGG CCA GTT
118 Gly Thr Asn Ala Phe Ile Ser Pro Trp Asn

651 AAG CCA TTC ATG CCA GTA GGC GCG CGG ACG
108 Leu Trp Glu. His Trp Tyr Ala Arg Pro Arg

681 AAA GTA AAC CCA CTG GTG ATA CCA TTC GCG
98 Phe Tyr Val Trp Gln His Tyr Trp Glu Arg

711 AGC CTC CGG ATG ACG ACC GTA GTG ATG AAT
88 Ala Glu Pro His Arg Gly Tyr His His Ile

741 CTC TCC TGG CGG GAA CAG CAA AAT ATC ACC
78 Glu Gly Pro Pro Phe Leu Leu Ile Asp Gly

771 CGG TCG GCA AAC AAA TTC TCG TCC CTG ATT
68 Pro Arg Cys Val Phe Glu Arg Gly Gln Asn

Figure 13b (cont'd)

```
801  TTT CAC CAC CCC CTG ACC GCG AAT GGT GAG
 58◀Lys Val Val Gly Gln Gly Arg Ile Thr Leu

831  ATT GAG AAT ATA ACC TTT CAT TCC CAG CGG
 48◀Asn Leu Ile Tyr Gly Lys Met Gly Leu Pro

861  TCG GTC GAT AAA AAA ATC GAG ATA ACC GTT
 38◀Arg Asp Ile Phe Phe Asp Leu Tyr Gly Asn

891  GGC CTC AAT CGG CGT TAA ACC CGC CAC CAG
 28◀Ala Glu Ile Pro Thr Leu Gly Ala Val Leu

921  ATG GGC ATT AAA CGA GTA TCC CGG CAG CAG
 18◀His Ala Asn Phe Ser Tyr Gly Pro Leu Leu

951  GGG ATC ATT TTG CGC TTC AGC CAT ACTTTTC
  8◀Pro Asp Asn Gln Ala Glu Ala Met

982  ATACTCCCGCCATTCAGAGAAGAAACCAATTGTCCATAT

1021 TGCATCAGACATTGCCGTCACTGCGTCTTTTACTGGCTC

1060 TTCTCGCTAACCAAACCGGTAACCCCGCTTATTAAAAGC

1099 ATTCTGTAACAAAGCGGGACCAAAGCCATGACAAAAACG

1138 CGTAACAAAAGTGTCTATAATCACGGCAGAAAAGTCCAC

1177 ATTGATTATTTGCACGGCGTCACACTTTGCTATGCCATA

                                    BamHI
1216 GCATTTTTATCCATAAGATTAGCGGATCCTACCTGACGC
```

Figure 13b (cont'd)

```
1255 TTTTTATCGCAACTCTCTACTGTTTCTCCATACCCGTTT
                NheI          EcoRI     NcoI  BamHI
1294 TTTTGGGCTAGCAGGAGGAAT TCACC ATG GAT CCC
                                    1▶Met Asp Pro

1329 GTA ATC GTA GAA GAC ATA GAG CCA GGT ATT
   4▶Val Ile Val Glu Asp Ile Glu Pro Gly Ile

1359 TAT TAC GGA ATT TCG AAT GAG AAT TAC CAC
  14▶Tyr Tyr Gly Ile Ser Asn Glu Asn Tyr His

1389 GCG GGT CCC GGT ATC AGT AAG TCT CAG CTC
  24▶Ala Gly Pro Gly Ile Ser Lys Ser Gln Leu

1419 GAT GAC ATT GCT GAT ACT CCG GCA CTA TAT
  34▶Asp Asp Ile Ala Asp Thr Pro Ala Leu Tyr

1449 TTG TGG CGT AAA AAT GCC CCC GTG GAC ACC
  44▶Leu Trp Arg Lys Asn Ala Pro Val Asp Thr

1479 ACA AAG ACA AAA ACG CTC GAT TTA GGA ACT
  54▶Thr Lys Thr Lys Thr Leu Asp Leu Gly Thr

1509 GCT TTC CAC TGC CGG GTA CTT GAA CCG GAA
  64▶Ala Phe His Cys Arg Val Leu Glu Pro Glu
       EcoRI
1539 GAA TTC AGT AAC CGC TTT ATC GTA GCA CCT
  74▶Glu Phe Ser Asn Arg Phe Ile Val Ala Pro

1569 GAA TTT AAC CGC CGT ACA AAC GCC GGA AAA
  84▶Glu Phe Asn Arg Arg Thr Asn Ala Gly Lys

1599 GAA GAA GAG AAA GCG TTT CTG ATG GAA TGC
  94▶Glu Glu Glu Lys Ala Phe Leu Met Glu Cys

1629 GCA AGC ACA GGA AAA ACG GTT ATC ACT GCG
 104▶Ala Ser Thr Gly Lys Thr Val Ile Thr Ala
```

Figure 13b (cont'd)

```
1659 GAA GAA GGC CGG AAA ATT GAA CTC ATG TAT
 114▶Glu Glu Gly Arg Lys Ile Glu Leu Met Tyr

1689 CAA AGC GTT ATG GCT TTG CCG CTG GGG CAA
 124▶Gln Ser Val Met Ala Leu Pro Leu Gly Gln

1719 TGG CTT GTT GAA AGC GCC GGA CAC GCT GAA
 134▶Trp Leu Val Glu Ser Ala Gly His Ala Glu

1749 TCA TCA ATT TAC TGG GAA GAT CCT GAA ACA
 144▶Ser Ser Ile Tyr Trp Glu Asp Pro Glu Thr

1779 GGA ATT TTG TGT CGG TGC CGT CCG GAC AAA
 154▶Gly Ile Leu Cys Arg Cys Arg Pro Asp Lys

1809 ATT ATC CCT GAA TTT CAC TGG ATC ATG GAC
 164▶Ile Ile Pro Glu Phe His Trp Ile Met Asp

1839 GTG AAA ACT ACG GCG GAT ATT CAA CGA TTC
 174▶Val Lys Thr Thr Ala Asp Ile Gln Arg Phe

1869 AAA ACC GCT TAT TAC GAC TAC CGC TAT CAC
 184▶Lys Thr Ala Tyr Tyr Asp Tyr Arg Tyr His

1899 GTT CAG GAT GCA TTC TAC AGT GAC GGT TAT
 194▶Val Gln Asp Ala Phe Tyr Ser Asp Gly Tyr

1929 GAA GCA CAG TTT GGA GTG CAG CCA ACT TTC
 204▶Glu Ala Gln Phe Gly Val Gln Pro Thr Phe

1959 GTT TTT CTG GTT GCC AGC ACA ACT ATT GAA
 214▶Val Phe Leu Val Ala Ser Thr Thr Ile Glu

1989 TGC GGA CGT TAT CCG GTT GAA ATT TTC ATG
 224▶Cys Gly Arg Tyr Pro Val Glu Ile Phe Met

2019 ATG GGC GAA GAA GCA AAA CTG GCA GGT CAA
 234▶Met Gly Glu Glu Ala Lys Leu Ala Gly Gln
```

Figure 13b (cont'd)

```
2049  CAG GAA TAT CAC CGC AAT CTG CGA ACC CTG
 244▶ Gln Glu Tyr His Arg Asn Leu Arg Thr Leu

2079  TCT GAC TGC CTG AAT ACC GAT GAA TGG CCA
 254▶ Ser Asp Cys Leu Asn Thr Asp Glu Trp Pro

2109  GCT ATT AAG ACA TTA TCA CTG CCC CGC TGG
 264▶ Ala Ile Lys Thr Leu Ser Leu Pro Arg Trp
                                        XhoI  KpnI
2139  GCT AAG GAA TAT GCA AAT GAC TAGATCTCGAG
 274▶ Ala Lys Glu Tyr Ala Asn Asp

2171  GTACCCGAGCACGTGTTGACAATTAATCATCGGCATAGT

2210  ATATCGGCATAGTATAATACGACAAGGTGAGGAACTAAA
      NcoI
2249  CC ATG GCT AAG CAA CCA CCA ATC GCA AAA
    1▶ Met Ala Lys Gln Pro Pro Ile Ala Lys

2278  GCC GAT CTG CAA AAA ACT CAG GGA AAC CGT
   10▶ Ala Asp Leu Gln Lys Thr Gln Gly Asn Arg

2308  GCA CCA GCA GCA GTT AAA AAT AGC GAC GTG
   20▶ Ala Pro Ala Ala Val Lys Asn Ser Asp Val

2338  ATT AGT TTT ATT AAC CAG CCA TCA ATG AAA
   30▶ Ile Ser Phe Ile Asn Gln Pro Ser Met Lys

2368  GAG CAA CTG GCA GCA GCT CTT CCA CGC CAT
   40▶ Glu Gln Leu Ala Ala Ala Leu Pro Arg His

2398  ATG ACG GCT GAA CGT ATG ATC CGT ATC GCC
   50▶ Met Thr Ala Glu Arg Met Ile Arg Ile Ala

2428  ACC ACA GAA ATT CGT AAA GTT CCG GCG TTA
   60▶ Thr Thr Glu Ile Arg Lys Val Pro Ala Leu
```

Figure 13b (cont'd)

```
2458  GGA AAC TGT GAC ACT ATG AGT TTT GTC AGT
  70▶ Gly Asn Cys Asp Thr Met Ser Phe Val Ser

2488  GCG ATC GTA CAG TGT TCA CAG CTC GGA CTT
  80▶ Ala Ile Val Gln Cys Ser Gln Leu Gly Leu

2518  GAG CCA GGT AGC GCC CTC GGT CAT GCA TAT
  90▶ Glu Pro Gly Ser Ala Leu Gly His Ala Tyr

2548  TTA CTG CCT TTT GGT AAT AAA AAC GAA AAG
 100▶ Leu Leu Pro Phe Gly Asn Lys Asn Glu Lys

2578  AGC GGT AAA AAG AAC GTT CAG CTA ATC ATT
 110▶ Ser Gly Lys Lys Asn Val Gln Leu Ile Ile

2608  GGC TAT CGC GGC ATG ATT GAT CTG GCT CGC
 120▶ Gly Tyr Arg Gly Met Ile Asp Leu Ala Arg

2638  CGT TCT GGT CAA ATC GCC AGC CTG TCA GCC
 130▶ Arg Ser Gly Gln Ile Ala Ser Leu Ser Ala

2668  CGT GTT GTC CGT GAA GGT GAC GAG TTT AGC
 140▶ Arg Val Val Arg Glu Gly Asp Glu Phe Ser

2698  TTC GAA TTT GGC CTT GAT GAA AAG TTA ATA
 150▶ Phe Glu Phe Gly Leu Asp Glu Lys Leu Ile

2728  CAC CGC CCG GGA GAA AAC GAA GAT GCC CCG
 160▶ His Arg Pro Gly Glu Asn Glu Asp Ala Pro

2758  GTT ACC CAC GTC TAT GCT GTC GCA AGA CTG
 170▶ Val Thr His Val Tyr Ala Val Ala Arg Leu

2788  AAA GAC GGA GGT ACT CAG TTT GAA GTT ATG
 180▶ Lys Asp Gly Gly Thr Gln Phe Glu Val Met

2818  ACG CGC AAA CAG ATT GAG CTG GTG CGC AGC
 190▶ Thr Arg Lys Gln Ile Glu Leu Val Arg Ser
```

Figure 13b (cont'd)

```
2848  CTG AGT AAA GCT GGT AAT AAC GGG CCG TGG
 200▶ Leu Ser Lys Ala Gly Asn Asn Gly Pro Trp

2878  GTA ACT CAC TGG GAA GAA ATG GCA AAG AAA
 210▶ Val Thr His Trp Glu Glu Met Ala Lys Lys

2908  ACG GCT ATT CGT CGC CTG TTC AAA TAT TTG
 220▶ Thr Ala Ile Arg Arg Leu Phe Lys Tyr Leu

2938  CCC GTA TCA ATT GAG ATC CAG CGT GCA GTA
 230▶ Pro Val Ser Ile Glu Ile Gln Arg Ala Val

2968  TCA ATG GAT GAA AAG GAA CCA CTG ACA ATC
 240▶ Ser Met Asp Glu Lys Glu Pro Leu Thr Ile

2998  GAT CCT GCA GAT TCC TCT GTA TTA ACC GGG
 250▶ Asp Pro Ala Asp Ser Ser Val Leu Thr Gly

3028  GAA TAC AGT GTA ATC GAT AAT TCA GAG GAA
 260▶ Glu Tyr Ser Val Ile Asp Asn Ser Glu Glu
        BglII     HindIII
3058  TAG ATCTAAGCTTCCTGCTGAACATCAAAGGCAAGAAA
 270▶ • • •

3096  ACATCTGTTGTCAAAGACAGCATCCTTGAACAAGGACAA

3135  TTAACAGTTAACAAATAAAAACGCAAAAGAAAATGCCGA

3174  TATCCTATTGGCATTTTCTTTTATTTCTTATCAACATAA
                        XhoI
3213  AGGTGAATCCCATACCTCGAGCTTCACGCTGCCGCAAGC

3252  ACTCAGGGCGCAAGGGCTGCTAAAAGGAAGCGGAACACG

3291  TAGAAAGCCAGTCCGCAGAAACGGTGCTGACCCCGGATG

3330  AATGTCAGCTACTGGGCTATCTGGACAAGGGAAAACGCA

3369  AGCGCAAAGAGAAAGCAGGTAGCTTGCAGTGGGCTTACA
```

Figure 13b (cont'd)

```
3408  TGGCGATAGCTAGACTGGGCGGTTTTATGGACAGCAAGC

3447  GAACCGGAATTGCCAGCTGGGGCGCCCTCTGGTAAGGTT

3486  GGGAAGCCCTGCAAAGTAAACTGGATGGCTTTCTTGCCG
                                        BglII
3525  CCAAGGATCTGATGGCGCAGGGGATCAAGATCTGATCAA

3564  GAGACAGGATGAGGATCGTTTCGC ATG GAT ATT
                                  1▶Met Asp Ile

3597  AAT ACT GAA ACT GAG ATC AAG CAA AAG CAT
   4▶Asn Thr Glu Thr Glu Ile Lys Gln Lys His

3627  TCA CTA ACC CCC TTT CCT GTT TTC CTA ATC
  14▶Ser Leu Thr Pro Phe Pro Val Phe Leu Ile

3657  AGC CCG GCA TTT CGC GGG CGA TAT TTT CAC
  24▶Ser Pro Ala Phe Arg Gly Arg Tyr Phe His

3687  AGC TAT TTC AGG AGT TCA GCC ATG AAC GCT
  34▶Ser Tyr Phe Arg Ser Ser Ala Met Asn Ala

3717  TAT TAC ATT CAG GAT CGT CTT GAG GCT CAG
  44▶Tyr Tyr Ile Gln Asp Arg Leu Glu Ala Gln

3747  AGC TGG GCG CGT CAC TAC CAG CAG CTC GCC
  54▶Ser Trp Ala Arg His Tyr Gln Gln Leu Ala

3777  CGT GAA GAG AAA GAG GCA GAA CTG GCA GAC
  64▶Arg Glu Glu Lys Glu Ala Glu Leu Ala Asp

3807  GAC ATG GAA AAA GGC CTG CCC CAG CAC CTG
  74▶Asp Met Glu Lys Gly Leu Pro Gln His Leu

3837  TTT GAA TCG CTA TGC ATC GAT CAT TTG CAA
  84▶Phe Glu Ser Leu Cys Ile Asp His Leu Gln

3867  CGC CAC GGG GCC AGC AAA AAA TCC ATT ACC
  94▶Arg His Gly Ala Ser Lys Lys Ser Ile Thr
```

Figure 13b (cont'd)

```
3897  CGT GCG TTT GAT GAC GAT GTT GAG TTT CAG
 104▶Arg Ala Phe Asp Asp Asp Val Glu Phe Gln

3927  GAG CGC ATG GCA GAA CAC ATC CGG TAC ATG
 114▶Glu Arg Met Ala Glu His Ile Arg Tyr Met

3957  GTT GAA ACC ATT GCT CAC CAC CAG GTT GAT
 124▶Val Glu Thr Ile Ala His His Gln Val Asp
                                        HindIII
3987  ATT GAT TCA GAG GTA TAA AACGAGTAGA AGCT
 134▶Ile Asp Ser Glu Val •••

4019  TGGCTGTTTTGGCGGATGAGAGAAGATTTTCAGCCTGAT

4058  ACAGATTAAATCAGAACGCAGAAGCGGTCTGATAAAACA

4097  GAATTTGCCTGGCGGCAGTAGCGCGGTGGTCCCACCTGA

4136  CCCCATGCCGAACTCAGAAGTGAAACGCCGTAGCGCCGA

4175  TGGTAGTGTGGGGTCTCCCCATGCGAGAGTAGGGAACTG

4214  CCAGGCATCAAATAAAACGAAAGGCTCAGTCGAAAGACT

4253  GGGCCTTTCGTTTTATCTGTTGTTTGTCGGTGAACGCTC

4292  TCCTGAGTAGGACAAATCCGCCGGGAGCGGATTTGAACG

4331  TTGCGAAGCAACGGCCCGGAGGGTGGCGGGCAGGACGCC

4370  CGCCATAAACTGCCAGGCATCAAATTAAGCAGAAGGCCA

4409  TCCTGACGGATGGCCTTTTTGCGTTTCTACAAACTCTTT

4448  TGTTTATTTTTCTAAATACATTCAAATATGTATCCGCTC

4487  ATGAGACAATAACCCTGATAAATGCTTCAATAATATTGA

4526  AAAAGGAAGAGT ATG AGT ATT CAA CAT TTC
                  1▶Met Ser Ile Gln His Phe
```

Figure 13b (cont'd)

```
4556 CGT GTC GCC CTT ATT CCC TTT TTT GCG GCA
   7▶Arg Val Ala Leu Ile Pro Phe Phe Ala Ala

4586 TTT TGC CTT CCT GTT TTT GCT CAC CCA GAA
  17▶Phe Cys Leu Pro Val Phe Ala His Pro Glu

4616 ACG CTG GTG AAA GTA AAA GAT GCT GAA GAT
  27▶Thr Leu Val Lys Val Lys Asp Ala Glu Asp

4646 CAG TTG GGT GCA CGA GTG GGT TAC ATC GAA
  37▶Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu

4676 CTG GAT CTC AAC AGC GGT AAG ATC CTT GAG
  47▶Leu Asp Leu Asn Ser Gly Lys Ile Leu Glu

4706 AGT TTT CGC CCC GAA GAA CGT TTT CCA ATG
  57▶Ser Phe Arg Pro Glu Glu Arg Phe Pro Met

4736 ATG AGC ACT TTT AAA GTT CTG CTA TGT GGC
  67▶Met Ser Thr Phe Lys Val Leu Leu Cys Gly

4766 GCG GTA TTA TCC CGT GTT GAC GCC GGG CAA
  77▶Ala Val Leu Ser Arg Val Asp Ala Gly Gln

4796 GAG CAA CTC GGT CGC CGC ATA CAC TAT TCT
  87▶Glu Gln Leu Gly Arg Arg Ile His Tyr Ser
                          ScaI
4826 CAG AAT GAC TTG GTT GAG TAC TCA CCA GTC
  97▶Gln Asn Asp Leu Val Glu Tyr Ser Pro Val

4856 ACA GAA AAG CAT CTT ACG GAT GGC ATG ACA
 107▶Thr Glu Lys His Leu Thr Asp Gly Met Thr

4886 GTA AGA GAA TTA TGC AGT GCT GCC ATA ACC
 117▶Val Arg Glu Leu Cys Ser Ala Ala Ile Thr

4916 ATG AGT GAT AAC ACT GCG GCC AAC TTA CTT
 127▶Met Ser Asp Asn Thr Ala Ala Asn Leu Leu
```

Figure 13b (cont'd)

```
4946 CTG ACA ACG ATC GGA GGA CCG AAG GAG CTA
 137▶Leu Thr Thr Ile Gly Gly Pro Lys Glu Leu

4976 ACC GCT TTT TTG CAC AAC ATG GGG GAT CAT
 147▶Thr Ala Phe Leu His Asn Met Gly Asp His

5006 GTA ACT CGC CTT GAT CGT TGG GAA CCG GAG
 157▶Val Thr Arg Leu Asp Arg Trp Glu Pro Glu

5036 CTG AAT GAA GCC ATA CCA AAC GAC GAG CGT
 167▶Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg

5066 GAC ACC ACG ATG CCT GTA GCA ATG GCA ACA
 177▶Asp Thr Thr Met Pro Val Ala Met Ala Thr

5096 ACG TTG CGC AAA CTA TTA ACT GGC GAA CTA
 187▶Thr Leu Arg Lys Leu Leu Thr Gly Glu Leu

5126 CTT ACT CTA GCT TCC CGG CAA CAA TTA ATA
 197▶Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile

5156 GAC TGG ATG GAG GCG GAT AAA GTT GCA GGA
 207▶Asp Trp Met Glu Ala Asp Lys Val Ala Gly

5186 CCA CTT CTG CGC TCG GCC CTT CCG GCT GGC
 217▶Pro Leu Leu Arg Ser Ala Leu Pro Ala Gly

5216 TGG TTT ATT GCT GAT AAA TCT GGA GCC GGT
 227▶Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly

5246 GAG CGT GGG TCT CGC GGT ATC ATT GCA GCA
 237▶Glu Arg Gly Ser Arg Gly Ile Ile Ala Ala

5276 CTG GGG CCA GAT GGT AAG CCC TCC CGT ATC
 247▶Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile

5306 GTA GTT ATC TAC ACG ACG GGG AGT CAG GCA
 257▶Val Val Ile Tyr Thr Thr Gly Ser Gln Ala
```

Figure 13b (cont'd)

```
5336  ACT ATG GAT GAA CGA AAT AGA CAG ATC GCT
 267▶ Thr Met Asp Glu Arg Asn Arg Gln Ile Ala

5366  GAG ATA GGT GCC TCA CTG ATT AAG CAT TGG
 277▶ Glu Ile Gly Ala Ser Leu Ile Lys His Trp

5396  TAA CTGTCAGACCAAGTTTACTCATATATACTTTAGAT
 287▶ • • •

5434  TGATTTACGCGCCCTGTAGCGGCGCATTAAGCGCGGCGG

5473  GTGTGGTGGTTACGCGCAGCGTGACCGCTACACTTGCCA

5512  GCGCCCTAGCGCCCGCTCCTTTCGCTTTCTTCCCTTCCT

5551  TTCTCGCCACGTTCGCCGGCTTTCCCCGTCAAGCTCTAA

5590  ATCGGGGGCTCCCTTTAGGGTTCCGATTTAGTGCTTTAC

5629  GGCACCTCGACCCCAAAAAACTTGATTTGGGTGATGGTT

5668  CACGTAGTGGGCCATCGCCCTGATAGACGGTTTTTCGCC

5707  CTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGACTCT

5746  TGTTCCAAACTTGAACAACACTCAACCCTATCTCGGGCT

5785  ATTCTTTTGATTTATAAGGGATTTTGCCGATTTCGGCCT

5824  ATTGGTTAAAAAATGAGCTGATTTAACAAAAATTTAACG

5863  CGAATTTTAACAAAATATTAACGTTTACAATTTAAAAGG

5902  ATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAA

5941  ATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCAGAC

5980  CCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTT

6019  TTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAAACCA

6058  CCGCTACCAGCGGTGGTTTGTTTGCCGGATCAAGAGCTA

6097  CCAACTCTTTTTCCGAAGGTAACTGGCTTCAGCAGAGCG
```

Figure 13b (cont'd)

6136 CAGATACCAAATACTGTCCTTCTAGTGTAGCCGTAGTTA

6175 GGCCACCACTTCAAGAACTCTGTAGCACCGCCTACATAC

6214 CTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGT

6253 GGCGATAAGTCGTGTCTTACCGGGTTGGACTCAAGACGA

6292 TAGTTACCGGATAAGGCGCAGCGGTCGGGCTGAACGGGG

6331 GGTTCGTGCACACAGCCCAGCTTGGAGCGAACGACCTAC

6370 ACCGAACTGAGATACCTACAGCGTGAGCTATGAGAAAGC

6409 GCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCG

6448 GTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAG

6487 CTTCCAGGGGGAAACGCCTGGTATCTTTATAGTCCTGTC

6526 GGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTTGTGA

6565 TGCTCGTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGC

6604 AACGCGGCCTTTTTACGGTTCCTGGCCTTTTGCTGGCCT

6643 TTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGATTCT

6682 GTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACC

6721 GCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTCAGTG

6760 AGCGAGGAAGCGGAAGAGCGCCTGATGCGGTATTTTCTC

6799 CTTACGCATCTGTGCGGTATTTCACACCGCATAGGGTCA

6838 TGGCTGCGCCCCGACACCCGCCAACACCCGCTGACGCGC

6877 CCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTACAGAC

6916 AAGCTGTGACCGTCTCCGGGAGCTGCATGTGTCAGAGGT

6955 TTTCACCGTCATCACCGAAACGCGCGAGGCAGCAAGGAG

Figure 13b (cont'd)

6994 ATGGCGCCCAACAGTCCCCCGGCCACGGGGCCTGCCACC

7033 ATACCCACGCCGAAACAAGCGCTCATGAGCCCGAAGTGG

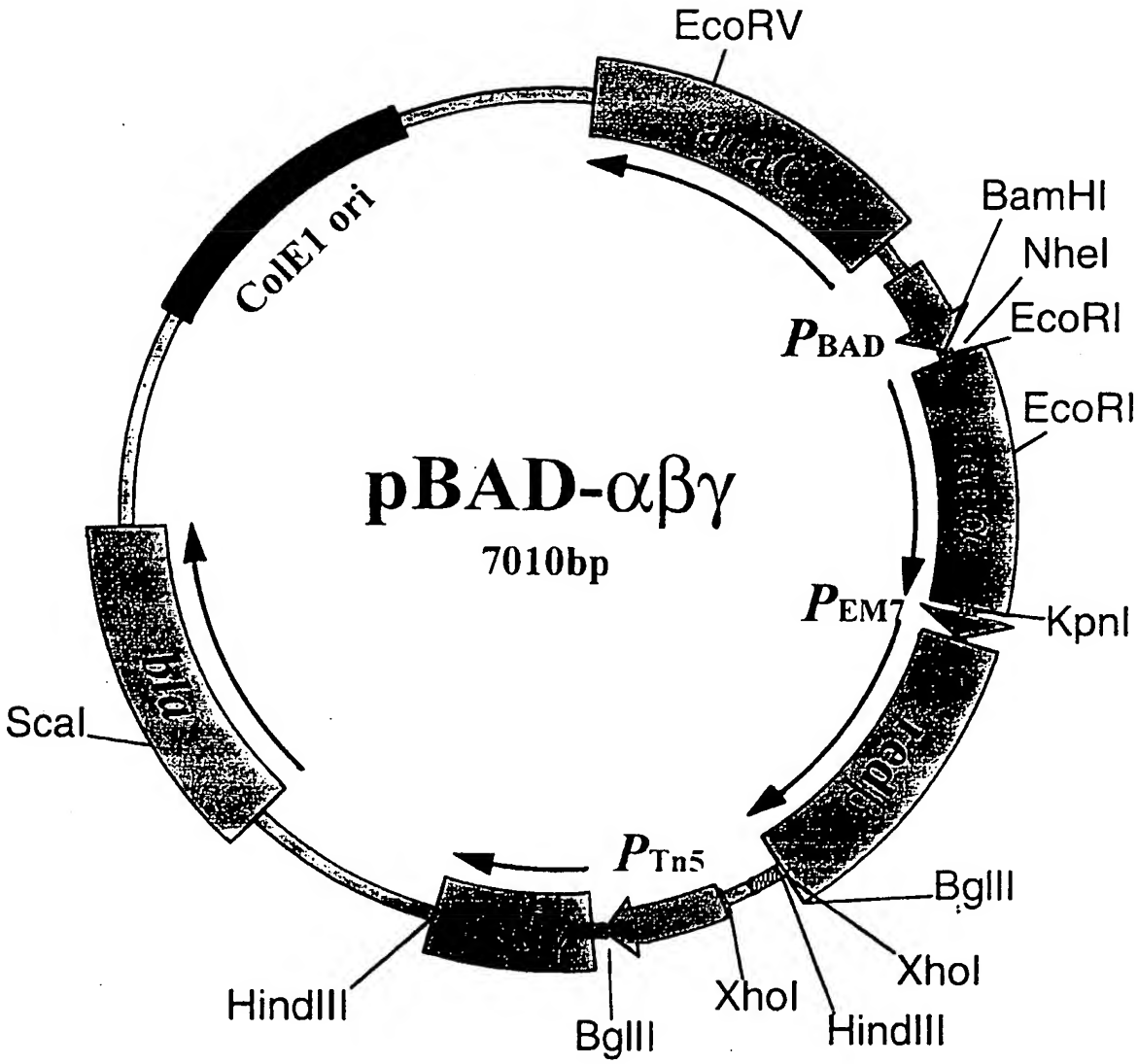
7072 CGAGCCCGATCTTCCCCATCGGTGATGTCGGCGATATAG

7111 GCGCCAGCAACCGCACCTGTGGCGCCGGTGATGCCGGCC

7150 ACGATGCGTCCGGCGTAGAGGATCTGCTCATGTTTGACA

7189 GCTTATC

Figure 14 a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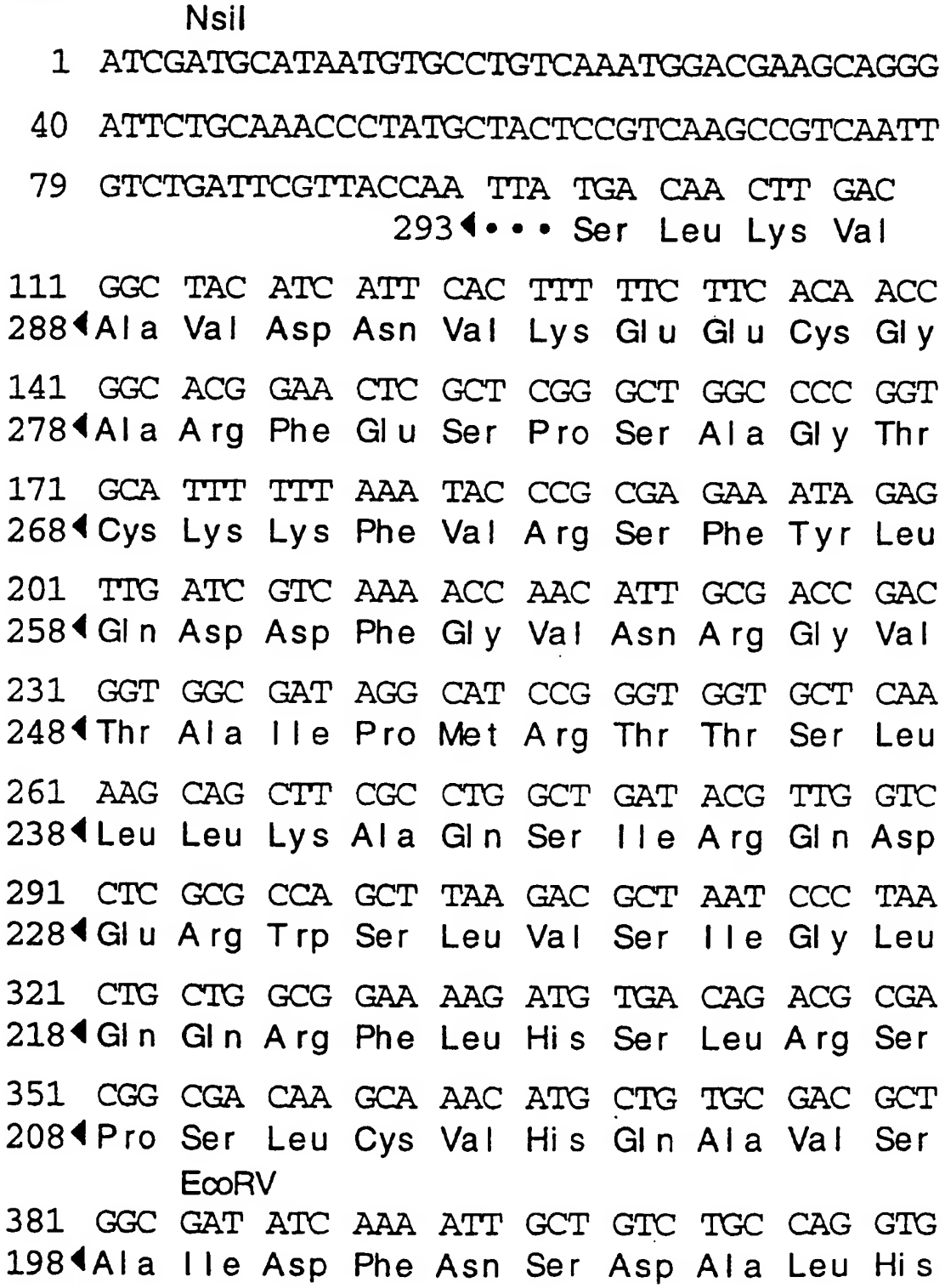
BamHI
NheI
EcoRI
EcoRI
KpnI
BglII
XhoI
HindIII
XhoI
BglII
HindIII
ScaI
ColE1 ori
pBAD-αβγ
7010bp
P_{BAD}
P_{EM7}
P_{Tn5}

Figure 14b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79 GTCTGATTCGTTACCAA TTA TGA CAA CTT GAC
293 • • • Ser Leu Lys Val
111 GGC TAC ATC ATT CAC TTT TTC TTC ACA ACC
288 Ala Val Asp Asn Val Lys Glu Glu Cys Gly
141 GGC ACG GAA CTC GCT CGG GCT GGC CCC GGT
278 Ala Arg Phe Glu Ser Pro Ser Ala Gly Thr
171 GCA TTT TTT AAA TAC CCG CGA GAA ATA GAG
268 Cys Lys Lys Phe Val Arg Ser Phe Tyr Leu
201 TTG ATC GTC AAA ACC AAC ATT GCG ACC GAC
258 Gln Asp Asp Phe Gly Val Asn Arg Gly Val
231 GGT GGC GAT AGG CAT CCG GGT GGT GCT CAA
248 Thr Ala Ile Pro Met Arg Thr Thr Ser Leu
261 AAG CAG CTT CGC CTG GCT GAT ACG TTG GTC
238 Leu Leu Lys Ala Gln Ser Ile Arg Gln Asp
291 CTC GCG CCA GCT TAA GAC GCT AAT CCC TAA
228 Glu Arg Trp Ser Leu Val Ser Ile Gly Leu
321 CTG CTG GCG GAA AAG ATG TGA CAG ACG CGA
218 Gln Gln Arg Phe Leu His Ser Leu Arg Ser
351 CGG CGA CAA GCA AAC ATG CTG TGC GAC GCT
208 Pro Ser Leu Cys Val His Gln Ala Val Ser
EcoRV
381 GGC GAT ATC AAA ATT GCT GTC TGC CAG GTG
198 Ala Ile Asp Phe Asn Ser Asp Ala Leu His

Figure 14b (cont'd)

411 ATC GCT GAT GTA CTG ACA AGC CTC GCG TAC
188◀Asp Ser Ile Tyr Gln Cys Ala Glu Arg Val

441 CCG ATT ATC CAT CGG TGG ATG GAG CGA CTC
178◀Arg Asn Asp Met Pro Pro His Leu Ser Glu

471 GTT AAT CGC TTC CAT GCG CCG CAG TAA CAA
168◀Asn Ile Ala Glu Met Arg Arg Leu Leu Leu

501 TTG CTC AAG CAG ATT TAT CGC CAG CAG CTC
158◀Gln Glu Leu Leu Asn Ile Ala Leu Leu Glu

531 CGA ATA GCG CCC TTC CCC TTG CCC GGC GTT
148◀Ser Tyr Arg Gly Glu Gly Gln Gly Ala Asn

561 AAT GAT TTG CCC AAA CAG GTC GCT GAA ATG
138◀Ile Ile Gln Gly Phe Leu Asp Ser Phe His

591 CGG CTG GTG CGC TTC ATC CGG GCG AAA GAA
128◀Pro Gln His Ala Glu Asp Pro Arg Phe Phe

621 CCC CGT ATT GGC AAA TAT TGA CGG CCA GTT
118◀Gly Thr Asn Ala Phe Ile Ser Pro Trp Asn

651 AAG CCA TTC ATG CCA GTA GGC GCG CGG ACG
108◀Leu Trp Glu His Trp Tyr Ala Arg Pro Arg

681 AAA GTA AAC CCA CTG GTG ATA CCA TTC GCG
98◀Phe Tyr Val Trp Gln His Tyr Trp Glu Arg

711 AGC CTC CGG ATG ACG ACC GTA GTG ATG AAT
88◀Ala Glu Pro His Arg Gly Tyr His His Ile

741 CTC TCC TGG CGG GAA CAG CAA AAT ATC ACC
78◀Glu Gly Pro Pro Phe Leu Leu Ile Asp Gly

771 CGG TCG GCA AAC AAA TTC TCG TCC CTG ATT
68◀Pro Arg Cys Val Phe Glu Arg Gly Gln Asn

Figure 14b (cont'd)

```
801  TTT CAC CAC CCC CTG ACC GCG AAT GGT GAG
 58◀Lys Val Val Gly Gln Gly Arg Ile Thr Leu

831  ATT GAG AAT ATA ACC TTT CAT TCC CAG CGG
 48◀Asn Leu Ile Tyr Gly Lys Met Gly Leu Pro

861  TCG GTC GAT AAA AAA ATC GAG ATA ACC GTT
 38◀Arg Asp Ile Phe Phe Asp Leu Tyr Gly Asn

891  GGC CTC AAT CGG CGT TAA ACC CGC CAC CAG
 28◀Ala Glu Ile Pro Thr Leu Gly Ala Val Leu

921  ATG GGC ATT AAA CGA GTA TCC CGG CAG CAG
 18◀His Ala Asn Phe Ser Tyr Gly Pro Leu Leu

951  GGG ATC ATT TTG CGC TTC AGC CAT ACTTTTC
   8◀Pro Asp Asn Gln Ala Glu Ala Met

 982 ATACTCCCGCCATTCAGAGAAGAAACCAATTGTCCATAT

1021 TGCATCAGACATTGCCGTCACTGCGTCTTTTACTGGCTC

1060 TTCTCGCTAACCAAACCGGTAACCCCGCTTATTAAAAGC

1099 ATTCTGTAACAAAGCGGGACCAAAGCCATGACAAAAACG

1138 CGTAACAAAAGTGTCTATAATCACGGCAGAAAAGTCCAC

1177 ATTGATTATTTGCACGGCGTCACACTTTGCTATGCCATA
                                  BamHI
1216 GCATTTTTATCCATAAGATTAGCGGATCCTACCTGACGC

1255 TTTTTATCGCAACTCTCTACTGTTTCTCCATACCCGTTT
          NheI          EcoRI
1294 TTTTGGGCTAGCAGGAGGAATTCACC ATG ACA CCG
                                  1▶Met Thr Pro
                    PstI
1329 GAC ATT ATC CTG CAG CGT ACC GGG ATC GAT
```

Figure 14b (cont'd)

```
   4▶Asp Ile Ile Leu Gln Arg Thr Gly Ile Asp

1359 GTG AGA GCT GTC GAA CAG GGG GAT GAT GCG
  14▶Val Arg Ala Val Glu Gln Gly Asp Asp Ala

1389 TGG CAC AAA TTA CGG CTC GGC GTC ATC ACC
  24▶Trp His Lys Leu Arg Leu Gly Val Ile Thr

1419 GCT TCA GAA GTT CAC AAC GTG ATA GCA AAA
  34▶Ala Ser Glu Val His Asn Val Ile Ala Lys

1449 CCC CGC TCC GGA AAG AAG TGG CCT GAC ATG
  44▶Pro Arg Ser Gly Lys Lys Trp Pro Asp Met

1479 AAA ATG TCC TAC TTC CAC ACC CTG CTT GCT
  54▶Lys Met Ser Tyr Phe His Thr Leu Leu Ala

1509 GAG GTT TGC ACC GGT GTG GCT CCG GAA GTT
  64▶Glu Val Cys Thr Gly Val Ala Pro Glu Val

1539 AAC GCT AAA GCA CTG GCC TGG GGA AAA CAG
  74▶Asn Ala Lys Ala Leu Ala Trp Gly Lys Gln
                                          EcoRI
1569 TAC GAG AAC GAC GCC AGA ACC CTG TTT GAA
  84▶Tyr Glu Asn Asp Ala Arg Thr Leu Phe Glu

1599 TTC ACT TCC GGC GTG AAT GTT ACT GAA TCC
  94▶Phe Thr Ser Gly Val Asn Val Thr Glu Ser

1629 CCG ATC ATC TAT CGC GAC GAA AGT ATG CGT
 104▶Pro Ile Ile Tyr Arg Asp Glu Ser Met Arg

1659 ACC GCC TGC TCT CCC GAT GGT TTA TGC AGT
 114▶Thr Ala Cys Ser Pro Asp Gly Leu Cys Ser

1689 GAC GGC AAC GGC CTT GAA CTG AAA TGC CCG
 124▶Asp Gly Asn Gly Leu Glu Leu Lys Cys Pro
```

Figure 14b (cont'd)

```
1719  TTT ACC TCC CGG GAT TTC ATG AAG TTC CGG
 134▶ Phe Thr Ser Arg Asp Phe Met Lys Phe Arg

1749  CTC GGT GGT TTC GAG GCC ATA AAG TCA GCT
 144▶ Leu Gly Gly Phe Glu Ala Ile Lys Ser Ala

1779  TAC ATG GCC CAG GTG CAG TAC AGC ATG TGG
 154▶ Tyr Met Ala Gln Val Gln Tyr Ser Met Trp

1809  GTG ACG CGA AAA AAT GCC TGG TAC TTT GCC
 164▶ Val Thr Arg Lys Asn Ala Trp Tyr Phe Ala

1839  AAC TAT GAC CCG CGT ATG AAG CGT GAA GGC
 174▶ Asn Tyr Asp Pro Arg Met Lys Arg Glu Gly

1869  CTG CAT TAT GTC GTG ATT GAG CGG GAT GAA
 184▶ Leu His Tyr Val Val Ile Glu Arg Asp Glu

1899  AAG TAC ATG GCG AGT TTT GAC GAG ATC GTG
 194▶ Lys Tyr Met Ala Ser Phe Asp Glu Ile Val

1929  CCG GAG TTC ATC GAA AAA ATG GAC GAG GCA
 204▶ Pro Glu Phe Ile Glu Lys Met Asp Glu Ala

1959  CTG GCT GAA ATT GGT TTT GTA TTT GGG GAG
 214▶ Leu Ala Glu Ile Gly Phe Val Phe Gly Glu
                          KpnI
1989  CAA TGG CGA TAGATCCGGTACCCGAGCACGTGTTGA
 224▶ Gln Trp Arg •••

2025  CAATTAATCATCGGCATAGTATATCGGCATAGTATAATA

2064  CGACAAGGTGAGGAACTAAACC ATG AGT ACT GCA
                            1▶ Met Ser Thr Ala

2098  CTC GCA ACG CTG GCT GGG AAG CTG GCT GAA
   5▶ Leu Ala Thr Leu Ala Gly Lys Leu Ala Glu
```

Figure 14b (cont'd)

```
                                                SalI
2128  CGT GTC GGC ATG GAT TCT GTC GAC CCA CAG
  15▶Arg Val Gly Met Asp Ser Val Asp Pro Gln

2158  GAA CTG ATC ACC ACT CTT CGC CAG ACG GCA
  25▶Glu Leu Ile Thr Thr Leu Arg Gln Thr Ala

2188  TTT AAA GGT GAT GCC AGC GAT GCG CAG TTC
  35▶Phe Lys Gly Asp Ala Ser Asp Ala Gln Phe

2218  ATC GCA TTA CTG ATC GTT GCC AAC CAG TAC
  45▶Ile Ala Leu Leu Ile Val Ala Asn Gln Tyr

2248  GGC CTT AAT CCG TGG ACG AAA GAA ATT TAC
  55▶Gly Leu Asn Pro Trp Thr Lys Glu Ile Tyr

2278  GCC TTT CCT GAT AAG CAG AAT GGC ATC GTT
  65▶Ala Phe Pro Asp Lys Gln Asn Gly Ile Val

2308  CCG GTG GTG GGC GTT GAT GGC TGG TCC CGC
  75▶Pro Val Val Gly Val Asp Gly Trp Ser Arg

2338  ATC ATC AAT GAA AAC CAG CAG TTT GAT GGC
  85▶Ile Ile Asn Glu Asn Gln Gln Phe Asp Gly

2368  ATG GAC TTT GAG CAG GAC AAT GAA TCC TGT
  95▶Met Asp Phe Glu Gln Asp Asn Glu Ser Cys

2398  ACA TGC CGG ATT TAC CGC AAG GAC CGT AAT
 105▶Thr Cys Arg Ile Tyr Arg Lys Asp Arg Asn

2428  CAT CCG ATC TGC GTT ACC GAA TGG ATG GAT
 115▶His Pro Ile Cys Val Thr Glu Trp Met Asp

2458  GAA TGC CGC CGC GAA CCA TTC AAA ACT CGC
 125▶Glu Cys Arg Arg Glu Pro Phe Lys Thr Arg

2488  GAA GGC AGA GAA ATC ACG GGG CCG TGG CAG
 135▶Glu Gly Arg Glu Ile Thr Gly Pro Trp Gln
```

Figure 14b (cont'd)

```
2518 TCG CAT CCC AAA CGG ATG TTA CGT CAT AAA
 145▶Ser His Pro Lys Arg Met Leu Arg His Lys

2548 GCC ATG ATT CAG TGT GCC CGT CTG GCC TTC
 155▶Ala Met Ile Gln Cys Ala Arg Leu Ala Phe

2578 GGA TTT GCT GGT ATC TAT GAC AAG GAT GAA
 165▶Gly Phe Ala Gly Ile Tyr Asp Lys Asp Glu

2608 GCC GAG CGC ATT GTC GAA AAT ACT GCA TAC
 175▶Ala Glu Arg Ile Val Glu Asn Thr Ala Tyr
       PstI
2638 ACT GCA GAA CGT CAG CCG GAA CGC GAC ATC
 185▶Thr Ala Glu Arg Gln Pro Glu Arg Asp Ile

2668 ACT CCG GTT AAC GAT GAA ACC ATG CAG GAG
 195▶Thr Pro Val Asn Asp Glu Thr Met Gln Glu

2698 ATT AAC ACT CTG CTG ATC GCC CTG GAT AAA
 205▶Ile Asn Thr Leu Leu Ile Ala Leu Asp Lys

2728 ACA TGG GAT GAC GAC TTA TTG CCG CTC TGT
 215▶Thr Trp Asp Asp Asp Leu Leu Pro Leu Cys

2758 TCC CAG ATA TTT CGC CGC GAC ATT CGT GCA
 225▶Ser Gln Ile Phe Arg Arg Asp Ile Arg Ala

2788 TCG TCA GAA CTG ACA CAG GCC GAA GCA GTA
 235▶Ser Ser Glu Leu Thr Gln Ala Glu Ala Val

2818 AAA GCT CTT GGA TTC CTG AAA CAG AAA GCC
 245▶Lys Ala Leu Gly Phe Leu Lys Gln Lys Ala
                                BglII XhoI
2848 GCA GAG CAG AAG GTG GCA GCA TAGATCTCGAG
 255▶Ala Glu Gln Lys Val Ala Ala •••
```

Figure 14b (cont'd)

HindIII

2880 AAGCTTCCTGCTGAACATCAAAGGCAAGAAAACATCTGT

2919 TGTCAAAGACAGCATCCTTGAACAAGGACAATTAACAGT

2958 TAACAAATAAAAACGCAAAAGAAAATGCCGATATCCTAT

2997 TGGCATTTTCTTTTATTTCTTATCAACATAAAGGTGAAT

XhoI

3036 CCCATACCTCGAGCTTCACGCTGCCGCAAGCACTCAGGG

3075 CGCAAGGGCTGCTAAAAGGAAGCGGAACACGTAGAAAGC

3114 CAGTCCGCAGAAACGGTGCTGACCCCGGATGAATGTCAG

3153 CTACTGGGCTATCTGGACAAGGGAAAACGCAAGCGCAAA

3192 GAGAAAGCAGGTAGCTTGCAGTGGGCTTACATGGCGATA

3231 GCTAGACTGGGCGGTTTTATGGACAGCAAGCGAACCGGA

PvuII

3270 ATTGCCAGCTGGGGCGCCCTCTGGTAAGGTTGGGAAGCC

3309 CTGCAAAGTAAACTGGATGGCTTTCTTGCCGCCAAGGAT

BglII

3348 CTGATGGCGCAGGGGATCAAGATCTGATCAAGAGACAGG

3387 ATGAGGATCGTTTCGC ATG GAT ATT AAT ACT
1▶Met Asp Ile Asn Thr

3418 GAA ACT GAG ATC AAG CAA AAG CAT TCA CTA
6▶Glu Thr Glu Ile Lys Gln Lys His Ser Leu

3448 ACC CCC TTT CCT GTT TTC CTA ATC AGC CCG
16▶Thr Pro Phe Pro Val Phe Leu Ile Ser Pro

3478 GCA TTT CGC GGG CGA TAT TTT CAC AGC TAT
26▶Ala Phe Arg Gly Arg Tyr Phe His Ser Tyr

3508 TTC AGG AGT TCA GCC ATG AAC GCT TAT TAC
36▶Phe Arg Ser Ser Ala Met Asn Ala Tyr Tyr

Figure 14b (cont'd)

```
3538  ATT CAG GAT CGT CTT GAG GCT CAG AGC TGG
  46▶ Ile Gln Asp Arg Leu Glu Ala Gln Ser Trp

3568  GCG CGT CAC TAC CAG CAG CTC GCC CGT GAA
  56▶ Ala Arg His Tyr Gln Gln Leu Ala Arg Glu

3598  GAG AAA GAG GCA GAA CTG GCA GAC GAC ATG
  66▶ Glu Lys Glu Ala Glu Leu Ala Asp Asp Met

3628  GAA AAA GGC CTG CCC CAG CAC CTG TTT GAA
  76▶ Glu Lys Gly Leu Pro Gln His Leu Phe Glu

3658  TCG CTA TGC ATC GAT CAT TTG CAA CGC CAC
  86▶ Ser Leu Cys Ile Asp His Leu Gln Arg His

3688  GGG GCC AGC AAA AAA TCC ATT ACC CGT GCG
  96▶ Gly Ala Ser Lys Lys Ser Ile Thr Arg Ala

3718  TTT GAT GAC GAT GTT GAG TTT CAG GAG CGC
 106▶ Phe Asp Asp Asp Val Glu Phe Gln Glu Arg

3748  ATG GCA GAA CAC ATC CGG TAC ATG GTT GAA
 116▶ Met Ala Glu His Ile Arg Tyr Met Val Glu

3778  ACC ATT GCT CAC CAC CAG GTT GAT ATT GAT
 126▶ Thr Ile Ala His His Gln Val Asp Ile Asp
                                  HindIII
3808  TCA GAG GTA TAA AACGAGTAGA AGC TTG GCT
 136▶ Ser Glu Val •••

3839  GTT TTG GCG GAT GAG AGA AGA TTT TCA GCC

3869  TGA TACAGATTAAATCAGAACGCAGAAGCGGTCTGATA

3907  AAACAGAATTTGCCTGGCGGCAGTAGCGCGGTGGTCCCA

3946  CCTGACCCCATGCCGAACTCAGAAGTGAAACGCCGTAGC

3985  GCCGATGGTAGTGTGGGGTCTCCCCATGCGAGAGTAGGG
```

Figure 14b (cont'd)

```
4024  AACTGCCAGGCATCAAATAAAACGAAAGGCTCAGTCGAA

4063  AGACTGGGCCTTTCGTTTTATCTGTTGTTTGTCGGTGAA

4102  CGCTCTCCTGAGTAGGACAAATCCGCCGGGAGCGGATTT

4141  GAACGTTGCGAAGCAACGGCCCGGAGGGTGGCGGGCAGG

4180  ACGCCCGCCATAAACTGCCAGGCATCAAATTAAGCAGAA

4219  GGCCATCCTGACGGATGGCCTTTTTGCGTTTCTACAAAC

4258  TCTTTTGTTTATTTTTCTAAATACATTCAAATATGTATC

4297  CGCTCATGAGACAATAACCCTGATAAATGCTTCAATAAT

4336  ATTGAAAAAGGAAGAGT ATG AGT ATT CAA CAT
                        1▶Met Ser Ile Gln His

4368  TTC CGT GTC GCC CTT ATT CCC TTT TTT GCG
    6▶Phe Arg Val Ala Leu Ile Pro Phe Phe Ala

4398  GCA TTT TGC CTT CCT GTT TTT GCT CAC CCA
   16▶Ala Phe Cys Leu Pro Val Phe Ala His Pro

4428  GAA ACG CTG GTG AAA GTA AAA GAT GCT GAA
   26▶Glu Thr Leu Val Lys Val Lys Asp Ala Glu

4458  GAT CAG TTG GGT GCA CGA GTG GGT TAC ATC
   36▶Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile

4488  GAA CTG GAT CTC AAC AGC GGT AAG ATC CTT
   46▶Glu Leu Asp Leu Asn Ser Gly Lys Ile Leu

4518  GAG AGT TTT CGC CCC GAA GAA CGT TTT CCA
   56▶Glu Ser Phe Arg Pro Glu Glu Arg Phe Pro

4548  ATG ATG AGC ACT TTT AAA GTT CTG CTA TGT
   66▶Met Met Ser Thr Phe Lys Val Leu Leu Cys
```

Figure 14b (cont'd)

```
4578  GGC GCG GTA TTA TCC CGT GTT GAC GCC GGG
  76▶ Gly Ala Val Leu Ser Arg Val Asp Ala Gly

4608  CAA GAG CAA CTC GGT CGC CGC ATA CAC TAT
  86▶ Gln Glu Gln Leu Gly Arg Arg Ile His Tyr
                                  ScaI
4638  TCT CAG AAT GAC TTG GTT GAG TAC TCA CCA
  96▶ Ser Gln Asn Asp Leu Val Glu Tyr Ser Pro

4668  GTC ACA GAA AAG CAT CTT ACG GAT GGC ATG
 106▶ Val Thr Glu Lys His Leu Thr Asp Gly Met

4698  ACA GTA AGA GAA TTA TGC AGT GCT GCC ATA
 116▶ Thr Val Arg Glu Leu Cys Ser Ala Ala Ile

4728  ACC ATG AGT GAT AAC ACT GCG GCC AAC TTA
 126▶ Thr Met Ser Asp Asn Thr Ala Ala Asn Leu

4758  CTT CTG ACA ACG ATC GGA GGA CCG AAG GAG
 136▶ Leu Leu Thr Thr Ile Gly Gly Pro Lys Glu

4788  CTA ACC GCT TTT TTG CAC AAC ATG GGG GAT
 146▶ Leu Thr Ala Phe Leu His Asn Met Gly Asp

4818  CAT GTA ACT CGC CTT GAT CGT TGG GAA CCG
 156▶ His Val Thr Arg Leu Asp Arg Trp Glu Pro

4848  GAG CTG AAT GAA GCC ATA CCA AAC GAC GAG
 166▶ Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu

4878  CGT GAC ACC ACG ATG CCT GTA GCA ATG GCA
 176▶ Arg Asp Thr Thr Met Pro Val Ala Met Ala

4908  ACA ACG TTG CGC AAA CTA TTA ACT GGC GAA
 186▶ Thr Thr Leu Arg Lys Leu Leu Thr Gly Glu

4938  CTA CTT ACT CTA GCT TCC CGG CAA CAA TTA
 196▶ Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu
```

Figure 14b (cont'd)

```
4968  ATA GAC TGG ATG GAG GCG GAT AAA GTT GCA
 206▶ Ile Asp Trp Met Glu Ala Asp Lys Val Ala

4998  GGA CCA CTT CTG CGC TCG GCC CTT CCG GCT
 216▶ Gly Pro Leu Leu Arg Ser Ala Leu Pro Ala

5028  GGC TGG TTT ATT GCT GAT AAA TCT GGA GCC
 226▶ Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala

5058  GGT GAG CGT GGG TCT CGC GGT ATC ATT GCA
 236▶ Gly Glu Arg Gly Ser Arg Gly Ile Ile Ala

5088  GCA CTG GGG CCA GAT GGT AAG CCC TCC CGT
 246▶ Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg

5118  ATC GTA GTT ATC TAC ACG ACG GGG AGT CAG
 256▶ Ile Val Val Ile Tyr Thr Thr Gly Ser Gln

5148  GCA ACT ATG GAT GAA CGA AAT AGA CAG ATC
 266▶ Ala Thr Met Asp Glu Arg Asn Arg Gln Ile

5178  GCT GAG ATA GGT GCC TCA CTG ATT AAG CAT
 276▶ Ala Glu Ile Gly Ala Ser Leu Ile Lys His

5208  TGG TAA CTGTCAGACCAAGTTTACTCATATATACTTT
 286▶ Trp •••

5245  AGATTGATTTACGCGCCCTGTAGCGGCGCATTAAGCGCG

5284  GCGGGTGTGGTGGTTACGCGCAGCGTGACCGCTACACTT

5323  GCCAGCGCCCTAGCGCCCGCTCCTTTCGCTTTCTTCCCT

5362  TCCTTTCTCGCCACGTTCGCCGGCTTTCCCCGTCAAGCT

5401  CTAAATCGGGGGCTCCCTTTAGGGTTCCGATTTAGTGCT

5440  TTACGGCACCTCGACCCCAAAAAACTTGATTTGGGTGAT

5479  GGTTCACGTAGTGGGCCATCGCCCTGATAGACGGTTTTT
```

Figure 14b (cont'd)

5518 CGCCCTTTGACGTTGGAGTCCACGTTCTTTAATAGTGGA

5557 CTCTTGTTCCAAACTTGAACAACACTCAACCCTATCTCG

5596 GGCTATTCTTTTGATTTATAAGGGATTTTGCCGATTTCG

5635 GCCTATTGGTTAAAAAATGAGCTGATTTAACAAAAATTT

5674 AACGCGAATTTTAACAAAATATTAACGTTTACAATTTAA

5713 AAGGATCTAGGTGAAGATCCTTTTTTGATAATCTCATGAC

5752 CAAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTC

5791 AGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCC

5830 TTTTTTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAA

5869 ACCACCGCTACCAGCGGTGGTTTGTTTGCCGGATCAAGA

5908 GCTACCAACTCTTTTTCCGAAGGTAACTGGCTTCAGCAG

5947 AGCGCAGATACCAAATACTGTCCTTCTAGTGTAGCCGTA

5986 GTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTAC

6025 ATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGC

6064 CAGTGGCGATAAGTCGTGTCTTACCGGGTTGGACTCAAG

6103 ACGATAGTTACCGGATAAGGCGCAGCGGTCGGGCTGAAC

6142 GGGGGGTTCGTGCACACAGCCCAGCTTGGAGCGAACGAC

6181 CTACACCGAACTGAGATACCTACAGCGTGAGCTATGAGA

6220 AAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTA

6259 TCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAG

6298 GGAGCTTCCAGGGGGAAACGCCTGGTATCTTTATAGTCC

6337 TGTCGGGTTTCGCCACCTCTGACTTGAGCGTCGATTTTT

Figure 14b (cont'd)

6376 GTGATGCTCGTCAGGGGGGCGGAGCCTATGGAAAAACGC

6415 CAGCAACGCGGCCTTTTTACGGTTCCTGGCCTTTTGCTG

6454 GCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCCTGA

6493 TTCTGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGA

6532 TACCGCTCGCCGCAGCCGAACGACCGAGCGCAGCGAGTC

6571 AGTGAGCGAGGAAGCGGAAGAGCGCCTGATGCGGTATTT

6610 TCTCCTTACGCATCTGTGCGGTATTTCACACCGCATAGG

6649 GTCATGGCTGCGCCCCGACACCCGCCAACACCCGCTGAC

6688 GCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCTTAC

6727 AGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTCAG

6766 AGGTTTTCACCGTCATCACCGAAACGCGCGAGGCAGCAA

6805 GGAGATGGCGCCCAACAGTCCCCCGGCCACGGGGCCTGC

6844 CACCATACCCACGCCGAAACAAGCGCTCATGAGCCCGAA

6883 GTGGCGAGCCCGATCTTCCCCATCGGTGATGTCGGCGAT

6922 ATAGGCGCCAGCAACCGCACCTGTGGCGCCGGTGATGCC

6961 GGCCACGATGCGTCCGGCGTAGAGGATCTGCTCATGTTT

7000 GACAGCTTATC

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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(B) STREET: Meyerhofstrasse 1
(C) CITY: Heidelberg
(E) COUNTRY: DE
(F) POSTAL CODE (ZIP): D-69117

(ii) TITLE OF INVENTION: Novel DNA Cloning Method

(iii) NUMBER OF SEQUENCES: 14

(iv) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: EP 97121562.2
(B) FILING DATE: 05-DEC-1997

(vi) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: EP 98118756.0
(B) FILING DATE: 05-OCT-1998

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6150 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: pBAD24-recET

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION:complement (96..974)
(D) OTHER INFORMATION:/product= "araC"

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION:1320..2162
(D) OTHER INFORMATION:/product= "t-recE"

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION:2155..2972
(D) OTHER INFORMATION:/product= "recT"

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION:3493..4353
(D) OTHER INFORMATION:/product= "bla"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATCGATGCAT AATGTGCCTG TCAAATGGAC GAAGCAGGGA TTCTGCAAAC CCTATGCTAC 60

TCCGTCAAGC CGTCAATTGT CTGATTCGTT ACCAATTATG ACAACTTGAC GGCTACATCA 120

TTCACTTTTT CTTCACAACC GGCACGGAAC TCGCTCGGGC TGGCCCCGGT GCATTTTTTA 180

AATACCCGCG AGAAATAGAG TTGATCGTCA AAACCAACAT TGCGACCGAC GGTGGCGATA 240

GGCATCCGGG TGGTGCTCAA AAGCAGCTTC GCCTGGCTGA TACGTTGGTC CTCGCGCCAG 300

CTTAAGACGC TAATCCCTAA CTGCTGGCGG AAAAGATGTG ACAGACGCGA CGGCGACAAG 360

CAAACATGCT GTGCGACGCT GGCGATATCA AAATTGCTGT CTGCCAGGTG ATCGCTGATG 420

TACTGACAAG CCTCGCGTAC CCGATTATCC ATCGGTGGAT GGAGCGACTC GTTAATCGCT 480

TCCATGCGCC GCAGTAACAA TTGCTCAAGC AGATTTATCG CCAGCAGCTC CGAATAGCGC 540

CCTTCCCCTT GCCCGGCGTT AATGATTTGC CCAAACAGGT CGCTGAAATG CGGCTGGTGC 600

GCTTCATCCG GGCGAAAGAA CCCCGTATTG GCAAATATTG ACGGCCAGTT AAGCCATTCA 660

TGCCAGTAGG CGCGCGGACG AAAGTAAACC CACTGGTGAT ACCATTCGCG AGCCTCCGGA 720

TGACGACCGT AGTGATGAAT CTCTCCTGGC GGGAACAGCA AAATATCACC CGGTCGGCAA 780

ACAAATTCTC GTCCCTGATT TTTCACCACC CCCTGACCGC GAATGGTGAG ATTGAGAATA 840

TAACCTTTCA TTCCCAGCGG TCGGTCGATA AAAAAATCGA GATAACCGTT GGCCTCAATC 900

GGCGTTAAAC CCGCCACCAG ATGGGCATTA AACGAGTATC CCGGCAGCAG GGGATCATTT 960

TGCGCTTCAG CCATACTTTT CATACTCCCG CCATTCAGAG AAGAAACCAA TTGTCCATAT 1020

TGCATCAGAC ATTGCCGTCA CTGCGTCTTT TACTGGCTCT TCTCGCTAAC CAAACCGGTA 1080

ACCCCGCTTA TTAAAAGCAT TCTGTAACAA AGCGGGACCA AAGCCATGAC AAAAACGCGT 1140

AACAAAAGTG TCTATAATCA CGGCAGAAAA GTCCACATTG ATTATTTGCA CGGCGTCACA 1200

CTTTGCTATG CCATAGCATT TTTATCCATA AGATTAGCGG ATCCTACCTG ACGCTTTTTA 1260

TCGCAACTCT CTACTGTTTC TCCATACCCG TTTTTTTGGG CTAGCAGGAG GAATTCACCA 1320

TGGATCCCGT AATCGTAGAA GACATAGAGC CAGGTATTTA TTACGGAATT TCGAATGAGA 1380

ATTACCACGC GGGTCCCGGT ATCAGTAAGT CTCAGCTCGA TGACATTGCT GATACTCCGG 1440

CACTATATTT GTGGCGTAAA AATGCCCCCG TGGACACCAC AAAGACAAAA ACGCTCGATT 1500

TAGGAACTGC TTTCCACTGC CGGGTACTTG AACCGGAAGA ATTCAGTAAC CGCTTTATCG 1560

TAGCACCTGA ATTTAACCGC CGTACAAACG CCGGAAAAGA AGAAGAGAAA GCGTTTCTGA 1620

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CGGACAAAAT TATCCCTGAA TTTCACTGGA TCATGGACGT GAAAACTACG GCGGATATTC 1860

AACGATTCAA AACCGCTTAT TACGACTACC GCTATCACGT TCAGGATGCA TTCTACAGTG 1920

ACGGTTATGA AGCACAGTTT GGAGTGCAGC CAACTTTCGT TTTTCTGGTT GCCAGCACAA 1980

CTATTGAATG CGGACGTTAT CCGGTTGAAA TTTTCATGAT GGGCGAAGAA GCAAAACTGG 2040

CAGGTCAACA GGAATATCAC CGCAATCTGC GAACCCTGTC TGACTGCCTG AATACCGATG 2100

AATGGCCAGC TATTAAGACA TTATCACTGC CCCGCTGGGC TAAGGAATAT GCAAATGACT 2160

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GCAGTTAAAA ATAGCGACGT GATTAGTTTT ATTAACCAGC CATCAATGAA AGAGCAACTG 2280

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CAGTGTTCAC AGCTCGGACT TGAGCCAGGT AGCGCCCTCG GTCATGCATA TTTACTGCCT 2460

TTTGGTAATA AAAACGAAAA GAGCGGTAAA AAGAACGTTC AGCTAATCAT TGGCTATCGC 2520

GGCATGATTG ATCTGGCTCG CCGTTCTGGT CAAATCGCCA GCCTGTCAGC CCGTGTTGTC 2580

CGTGAAGGTG ACGAGTTTAG CTTCGAATTT GGCCTTGATG AAAAGTTAAT ACACCGCCCG 2640

GGAGAAAACG AAGATGCCCC GGTTACCCAC GTCTATGCTG TCGCAAGACT GAAAGACGGA 2700

GGTACTCAGT TTGAAGTTAT GACGCGCAAA CAGATTGAGC TGGTGCGCAG CCTGAGTAAA 2760

GCTGGTAATA ACGGGCCGTG GGTAACTCAC TGGGAAGAAA TGGCAAAGAA AACGGCTATT 2820

CGTCGCCTGT TCAAATATTT GCCCGTATCA ATTGAGATCC AGCGTGCAGT ATCAATGGAT 2880

GAAAAGGAAC CACTGACAAT CGATCCTGCA GATTCCTCTG TATTAACCGG GGAATACAGT 2940

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CCGGAGGGTG GCGGGCAGGA CGCCCGCCAT AAACTGCCAG GCATCAAATT AAGCAGAAGG 3360

CCATCCTGAC GGATGGCCTT TTTGCGTTTC TACAAACTCT TTTGTTTATT TTTCTAAATA 3420

CATTCAAATA TGTATCCGCT CATGAGACAA TAACCCTGAT AAATGCTTCA ATAATATTGA 3480

AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTCGCCC TTATTCCCTT TTTTGCGGCA 3540

TTTTGCCTTC CTGTTTTTGC TCACCCAGAA ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT 3600

CAGTTGGGTG CACGAGTGGG TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG 3660

AGTTTTCGCC CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 3720

GCGGTATTAT CCCGTGTTGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT ACACTATTCT 3780

CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC ATCTTACGGA TGGCATGACA 3840

GTAAGAGAAT TATGCAGTGC TGCCATAACC ATGAGTGATA ACACTGCGGC CAACTTACTT 3900

CTGACAACGA TCGGAGGACC GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT 3960

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CCACTTCTGC GCTCGGCCCT TCCGGCTGGC TGGTTTATTG CTGATAAATC TGGAGCCGGT 4200

GAGCGTGGGT CTCGCGGTAT CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC 4260

GTAGTTATCT ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 4320

GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA CTCATATATA 4380

CTTTAGATTG ATTTACGCGC CCTGTAGCGG CGCATTAAGC GCGGCGGGTG TGGTGGTTAC 4440

GCGCAGCGTG ACCGCTACAC TTGCCAGCGC CCTAGCGCCC GCTCCTTTCG CTTTCTTCCC 4500

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GCGTAATCTG CTGCTTGCAA ACAAAAAAAC CACCGCTACC AGCGGTGGTT TGTTTGCCGG 5040

ATCAAGAGCT ACCAACTCTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA 5100

ATACTGTCCT TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5160

CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC GATAAGTCGT 5220

GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA GGCGCAGCGG TCGGGCTGAA 5280

CGGGGGGTTC GTGCACACAG CCCAGCTTGG AGCGAACGAC CTACACCGAA CTGAGATACC 5340

TACAGCGTGA GCTATGAGAA AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC 5400

CGGTAAGCGG CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 5460

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA TTTTTGTGAT 5520

GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA CGCGGCCTTT TTACGGTTCC 5580

TGGCCTTTTG CTGGCCTTTT GCTCACATGT TCTTTCCTGC GTTATCCCCT GATTCTGTGG 5640

ATAACCGTAT TACCGCCTTT GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC 5700

GCAGCGAGTC AGTGAGCGAG GAAGCGGAAG AGCGCCTGAT GCGGTATTTT CTCCTTACGC 5760

ATCTGTGCGG TATTTCACAC CGCATAGGGT CATGGCTGCG CCCCGACACC CGCCAACACC 5820

CGCTGACGCG CCCTGACGGG CTTGTCTGCT CCCGGCATCC GCTTACAGAC AAGCTGTGAC 5880

CGTCTCCGGG AGCTGCATGT GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA 5940

GCAAGGAGAT GGCGCCCAAC AGTCCCCCGG CCACGGGGCC TGCCACCATA CCCACGCCGA 6000

AACAAGCGCT CATGAGCCCG AAGTGGCGAG CCCGATCTTC CCCATCGGTG ATGTCGGCGA 6060

TATAGGCGCC AGCAACCGCA CCTGTGGCGC CGGTGATGCC GGCCACGATG CGTCCGGCGT 6120

AGAGGATCTG CTCATGTTTG ACAGCTTATC 6150

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 843 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: t-recE

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:1..843
(D) OTHER INFORMATION:/product= "t-recE"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

```
ATG GAT CCC GTA ATC GTA GAA GAC ATA GAG CCA GGT ATT TAT TAC GGA      48
Met Asp Pro Val Ile Val Glu Asp Ile Glu Pro Gly Ile Tyr Tyr Gly
 1               5                  10                  15

ATT TCG AAT GAG AAT TAC CAC GCG GGT CCC GGT ATC AGT AAG TCT CAG      96
Ile Ser Asn Glu Asn Tyr His Ala Gly Pro Gly Ile Ser Lys Ser Gln
            20                  25                  30

CTC GAT GAC ATT GCT GAT ACT CCG GCA CTA TAT TTG TGG CGT AAA AAT     144
Leu Asp Asp Ile Ala Asp Thr Pro Ala Leu Tyr Leu Trp Arg Lys Asn
        35                  40                  45

GCC CCC GTG GAC ACC ACA AAG ACA AAA ACG CTC GAT TTA GGA ACT GCT     192
Ala Pro Val Asp Thr Thr Lys Thr Lys Thr Leu Asp Leu Gly Thr Ala
    50                  55                  60

TTC CAC TGC CGG GTA CTT GAA CCG GAA GAA TTC AGT AAC CGC TTT ATC     240
Phe His Cys Arg Val Leu Glu Pro Glu Glu Phe Ser Asn Arg Phe Ile
65                  70                  75                  80

GTA GCA CCT GAA TTT AAC CGC CGT ACA AAC GCC GGA AAA GAA GAA GAG     288
Val Ala Pro Glu Phe Asn Arg Arg Thr Asn Ala Gly Lys Glu Glu Glu
                85                  90                  95

AAA GCG TTT CTG ATG GAA TGC GCA AGC ACA GGA AAA ACG GTT ATC ACT     336
Lys Ala Phe Leu Met Glu Cys Ala Ser Thr Gly Lys Thr Val Ile Thr
            100                 105                 110
```

```
GCG GAA GAA GGC CGG AAA ATT GAA CTC ATG TAT CAA AGC GTT ATG GCT      384
Ala Glu Glu Gly Arg Lys Ile Glu Leu Met Tyr Gln Ser Val Met Ala
        115                 120                 125

TTG CCG CTG GGG CAA TGG CTT GTT GAA AGC GCC GGA CAC GCT GAA TCA      432
Leu Pro Leu Gly Gln Trp Leu Val Glu Ser Ala Gly His Ala Glu Ser
    130                 135                 140

TCA ATT TAC TGG GAA GAT CCT GAA ACA GGA ATT TTG TGT CGG TGC CGT      480
Ser Ile Tyr Trp Glu Asp Pro Glu Thr Gly Ile Leu Cys Arg Cys Arg
145                 150                 155                 160

CCG GAC AAA ATT ATC CCT GAA TTT CAC TGG ATC ATG GAC GTG AAA ACT      528
Pro Asp Lys Ile Ile Pro Glu Phe His Trp Ile Met Asp Val Lys Thr
                165                 170                 175

ACG GCG GAT ATT CAA CGA TTC AAA ACC GCT TAT TAC GAC TAC CGC TAT      576
Thr Ala Asp Ile Gln Arg Phe Lys Thr Ala Tyr Tyr Asp Tyr Arg Tyr
            180                 185                 190

CAC GTT CAG GAT GCA TTC TAC AGT GAC GGT TAT GAA GCA CAG TTT GGA      624
His Val Gln Asp Ala Phe Tyr Ser Asp Gly Tyr Glu Ala Gln Phe Gly
        195                 200                 205

GTG CAG CCA ACT TTC GTT TTT CTG GTT GCC AGC ACA ACT ATT GAA TGC      672
Val Gln Pro Thr Phe Val Phe Leu Val Ala Ser Thr Thr Ile Glu Cys
    210                 215                 220

GGA CGT TAT CCG GTT GAA ATT TTC ATG ATG GGC GAA GAA GCA AAA CTG      720
Gly Arg Tyr Pro Val Glu Ile Phe Met Met Gly Glu Glu Ala Lys Leu
225                 230                 235                 240

GCA GGT CAA CAG GAA TAT CAC CGC AAT CTG CGA ACC CTG TCT GAC TGC      768
Ala Gly Gln Gln Glu Tyr His Arg Asn Leu Arg Thr Leu Ser Asp Cys
                245                 250                 255

CTG AAT ACC GAT GAA TGG CCA GCT ATT AAG ACA TTA TCA CTG CCC CGC      816
Leu Asn Thr Asp Glu Trp Pro Ala Ile Lys Thr Leu Ser Leu Pro Arg
            260                 265                 270

TGG GCT AAG GAA TAT GCA AAT GAC TAA                                  843
Trp Ala Lys Glu Tyr Ala Asn Asp  *
        275                 280
```

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 281 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

```
Met Asp Pro Val Ile Val Glu Asp Ile Glu Pro Gly Ile Tyr Tyr Gly
 1               5                  10                  15

Ile Ser Asn Glu Asn Tyr His Ala Gly Pro Gly Ile Ser Lys Ser Gln
            20                  25                  30

Leu Asp Asp Ile Ala Asp Thr Pro Ala Leu Tyr Leu Trp Arg Lys Asn
        35                  40                  45

Ala Pro Val Asp Thr Thr Lys Thr Lys Thr Leu Asp Leu Gly Thr Ala
    50                  55                  60
```

```
Phe His Cys Arg Val Leu Glu Pro Glu Glu Phe Ser Asn Arg Phe Ile
 65                  70                  75                  80

Val Ala Pro Glu Phe Asn Arg Arg Thr Asn Ala Gly Lys Glu Glu Glu
                85                  90                  95

Lys Ala Phe Leu Met Glu Cys Ala Ser Thr Gly Lys Thr Val Ile Thr
            100                 105                 110

Ala Glu Glu Gly Arg Lys Ile Glu Leu Met Tyr Gln Ser Val Met Ala
        115                 120                 125

Leu Pro Leu Gly Gln Trp Leu Val Glu Ser Ala Gly His Ala Glu Ser
    130                 135                 140

Ser Ile Tyr Trp Glu Asp Pro Glu Thr Gly Ile Leu Cys Arg Cys Arg
145                 150                 155                 160

Pro Asp Lys Ile Ile Pro Glu Phe His Trp Ile Met Asp Val Lys Thr
                165                 170                 175

Thr Ala Asp Ile Gln Arg Phe Lys Thr Ala Tyr Tyr Asp Tyr Arg Tyr
            180                 185                 190

His Val Gln Asp Ala Phe Tyr Ser Asp Gly Tyr Glu Ala Gln Phe Gly
        195                 200                 205

Val Gln Pro Thr Phe Val Phe Leu Val Ala Ser Thr Thr Ile Glu Cys
    210                 215                 220

Gly Arg Tyr Pro Val Glu Ile Phe Met Met Gly Glu Glu Ala Lys Leu
225                 230                 235                 240

Ala Gly Gln Gln Glu Tyr His Arg Asn Leu Arg Thr Leu Ser Asp Cys
                245                 250                 255

Leu Asn Thr Asp Glu Trp Pro Ala Ile Lys Thr Leu Ser Leu Pro Arg
            260                 265                 270

Trp Ala Lys Glu Tyr Ala Asn Asp  *
        275                 280
```

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 810 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: recT

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:1..810
(D) OTHER INFORMATION:/product= "recT"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

```
ATG ACT AAG CAA CCA CCA ATC GCA AAA GCC GAT CTG CAA AAA ACT CAG      48
Met Thr Lys Gln Pro Pro Ile Ala Lys Ala Asp Leu Gln Lys Thr Gln
            285                 290                 295
```

```
GGA AAC CGT GCA CCA GCA GCA GTT AAA AAT AGC GAC GTG ATT AGT TTT          96
Gly Asn Arg Ala Pro Ala Ala Val Lys Asn Ser Asp Val Ile Ser Phe
        300                 305                 310

ATT AAC CAG CCA TCA ATG AAA GAG CAA CTG GCA GCA GCT CTT CCA CGC         144
Ile Asn Gln Pro Ser Met Lys Glu Gln Leu Ala Ala Ala Leu Pro Arg
    315                 320                 325

CAT ATG ACG GCT GAA CGT ATG ATC CGT ATC GCC ACC ACA GAA ATT CGT         192
His Met Thr Ala Glu Arg Met Ile Arg Ile Ala Thr Thr Glu Ile Arg
330                 335                 340                 345


AAA GTT CCG GCG TTA GGA AAC TGT GAC ACT ATG AGT TTT GTC AGT GCG         240
Lys Val Pro Ala Leu Gly Asn Cys Asp Thr Met Ser Phe Val Ser Ala
                350                 355                 360

ATC GTA CAG TGT TCA CAG CTC GGA CTT GAG CCA GGT AGC GCC CTC GGT         288
Ile Val Gln Cys Ser Gln Leu Gly Leu Glu Pro Gly Ser Ala Leu Gly
            365                 370                 375

CAT GCA TAT TTA CTG CCT TTT GGT AAT AAA AAC GAA AAG AGC GGT AAA         336
His Ala Tyr Leu Leu Pro Phe Gly Asn Lys Asn Glu Lys Ser Gly Lys
        380                 385                 390

AAG AAC GTT CAG CTA ATC ATT GGC TAT CGC GGC ATG ATT GAT CTG GCT         384
Lys Asn Val Gln Leu Ile Ile Gly Tyr Arg Gly Met Ile Asp Leu Ala
    395                 400                 405

CGC CGT TCT GGT CAA ATC GCC AGC CTG TCA GCC CGT GTT GTC CGT GAA         432
Arg Arg Ser Gly Gln Ile Ala Ser Leu Ser Ala Arg Val Val Arg Glu
410                 415                 420                 425

GGT GAC GAG TTT AGC TTC GAA TTT GGC CTT GAT GAA AAG TTA ATA CAC         480
Gly Asp Glu Phe Ser Phe Glu Phe Gly Leu Asp Glu Lys Leu Ile His
                430                 435                 440

CGC CCG GGA GAA AAC GAA GAT GCC CCG GTT ACC CAC GTC TAT GCT GTC         528
Arg Pro Gly Glu Asn Glu Asp Ala Pro Val Thr His Val Tyr Ala Val
            445                 450                 455

GCA AGA CTG AAA GAC GGA GGT ACT CAG TTT GAA GTT ATG ACG CGC AAA         576
Ala Arg Leu Lys Asp Gly Gly Thr Gln Phe Glu Val Met Thr Arg Lys
        460                 465                 470

CAG ATT GAG CTG GTG CGC AGC CTG AGT AAA GCT GGT AAT AAC GGG CCG         624
Gln Ile Glu Leu Val Arg Ser Leu Ser Lys Ala Gly Asn Asn Gly Pro
    475                 480                 485

TGG GTA ACT CAC TGG GAA GAA ATG GCA AAG AAA ACG GCT ATT CGT CGC         672
Trp Val Thr His Trp Glu Glu Met Ala Lys Lys Thr Ala Ile Arg Arg
490                 495                 500                 505

CTG TTC AAA TAT TTG CCC GTA TCA ATT GAG ATC CAG CGT GCA GTA TCA         720
Leu Phe Lys Tyr Leu Pro Val Ser Ile Glu Ile Gln Arg Ala Val Ser
                510                 515                 520

ATG GAT GAA AAG GAA CCA CTG ACA ATC GAT CCT GCA GAT TCC TCT GTA         768
Met Asp Glu Lys Glu Pro Leu Thr Ile Asp Pro Ala Asp Ser Ser Val
            525                 530                 535

TTA ACC GGG GAA TAC AGT GTA ATC GAT AAT TCA GAG GAA TAG                 810
Leu Thr Gly Glu Tyr Ser Val Ile Asp Asn Ser Glu Glu  *
        540                 545                 550
```

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 270 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

```
Met Thr Lys Gln Pro Pro Ile Ala Lys Ala Asp Leu Gln Lys Thr Gln
 1               5                  10                  15

Gly Asn Arg Ala Pro Ala Ala Val Lys Asn Ser Asp Val Ile Ser Phe
            20                  25                  30

Ile Asn Gln Pro Ser Met Lys Glu Gln Leu Ala Ala Ala Leu Pro Arg
        35                  40                  45

His Met Thr Ala Glu Arg Met Ile Arg Ile Ala Thr Thr Glu Ile Arg
    50                  55                  60

Lys Val Pro Ala Leu Gly Asn Cys Asp Thr Met Ser Phe Val Ser Ala
65                  70                  75                  80

Ile Val Gln Cys Ser Gln Leu Gly Leu Glu Pro Gly Ser Ala Leu Gly
                85                  90                  95

His Ala Tyr Leu Leu Pro Phe Gly Asn Lys Asn Glu Lys Ser Gly Lys
            100                 105                 110

Lys Asn Val Gln Leu Ile Ile Gly Tyr Arg Gly Met Ile Asp Leu Ala
        115                 120                 125

Arg Arg Ser Gly Gln Ile Ala Ser Leu Ser Ala Arg Val Val Arg Glu
    130                 135                 140

Gly Asp Glu Phe Ser Phe Glu Phe Gly Leu Asp Glu Lys Leu Ile His
145                 150                 155                 160

Arg Pro Gly Glu Asn Glu Asp Ala Pro Val Thr His Val Tyr Ala Val
                165                 170                 175

Ala Arg Leu Lys Asp Gly Gly Thr Gln Phe Glu Val Met Thr Arg Lys
            180                 185                 190

Gln Ile Glu Leu Val Arg Ser Leu Ser Lys Ala Gly Asn Asn Gly Pro
        195                 200                 205

Trp Val Thr His Trp Glu Glu Met Ala Lys Lys Thr Ala Ile Arg Arg
    210                 215                 220

Leu Phe Lys Tyr Leu Pro Val Ser Ile Glu Ile Gln Arg Ala Val Ser
225                 230                 235                 240

Met Asp Glu Lys Glu Pro Leu Thr Ile Asp Pro Ala Asp Ser Ser Val
                245                 250                 255

Leu Thr Gly Glu Tyr Ser Val Ile Asp Asn Ser Glu Glu  *
            260                 265                 270
```

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 876 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: araC

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:complement (1..876)
(D) OTHER INFORMATION:/product= "araC"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

TGACAACTTG ACGGCTACAT CATTCACTTT TTCTTCACAA CCGGCACGGA ACTCGCTCGG 60

GCTGGCCCCG GTGCATTTTT TAAATACCCG CGAGAAATAG AGTTGATCGT CAAAACCAAC 120

ATTGCGACCG ACGGTGGCGA TAGGCATCCG GGTGGTGCTC AAAAGCAGCT TCGCCTGGCT 180

GATACGTTGG TCCTCGCGCC AGCTTAAGAC GCTAATCCCT AACTGCTGGC GGAAAAGATG 240

TGACAGACGC GACGGCGACA AGCAAACATG CTGTGCGACG CTGGCGATAT CAAAATTGCT 300

GTCTGCCAGG TGATCGCTGA TGTACTGACA AGCCTCGCGT ACCCGATTAT CCATCGGTGG 360

ATGGAGCGAC TCGTTAATCG CTTCCATGCG CCGCAGTAAC AATTGCTCAA GCAGATTTAT 420

CGCCAGCAGC TCCGAATAGC GCCCTTCCCC TTGCCCGGCG TTAATGATTT GCCCAAACAG 480

GTCGCTGAAA TGCGGCTGGT GCGCTTCATC CGGGCGAAAG AACCCCGTAT TGGCAAATAT 540

TGACGGCCAG TTAAGCCATT CATGCCAGTA GGCGCGCGGA CGAAAGTAAA CCCACTGGTG 600

ATACCATTCG CGAGCCTCCG GATGACGACC GTAGTGATGA ATCTCTCCTG GCGGGAACAG 660

CAAAATATCA CCCGGTCGGC AAACAAATTC TCGTCCCTGA TTTTTCACCA CCCCCTGACC 720

GCGAATGGTG AGATTGAGAA TATAACCTTT CATTCCCAGC GGTCGGTCGA TAAAAAAATC 780

GAGATAACCG TTGGCCTCAA TCGGCGTTAA ACCCGCCACC AGATGGGCAT TAAACGAGTA 840

TCCCGGCAGC AGGGGATCAT TTTGCGCTTC AGCCAT 876

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 292 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

Met Ala Glu Ala Gln Asn Asp Pro Leu Leu Pro Gly Tyr Ser Phe Asn
1 5 10 15

Ala His Leu Val Ala Gly Leu Thr Pro Ile Glu Ala Asn Gly Tyr Leu
20 25 30

```
Asp Phe Phe Ile Asp Arg Pro Leu Gly Met Lys Gly Tyr Ile Leu Asn
        35                  40                  45

Leu Thr Ile Arg Gly Gln Gly Val Val Lys Asn Gln Gly Arg Glu Phe
    50                  55                  60

Val Cys Arg Pro Gly Asp Ile Leu Leu Phe Pro Pro Gly Glu Ile His
65                  70                  75                  80

His Tyr Gly Arg His Pro Glu Ala Arg Glu Trp Tyr His Gln Trp Val
                85                  90                  95

Tyr Phe Arg Pro Arg Ala Tyr Trp His Glu Trp Leu Asn Trp Pro Ser
            100                 105                 110

Ile Phe Ala Asn Thr Gly Phe Phe Arg Pro Asp Glu Ala His Gln Pro
        115                 120                 125

His Phe Ser Asp Leu Phe Gly Gln Ile Ile Asn Ala Gly Gln Gly Glu
    130                 135                 140

Gly Arg Tyr Ser Glu Leu Leu Ala Ile Asn Leu Leu Glu Gln Leu Leu
145                 150                 155                 160

Leu Arg Arg Met Glu Ala Ile Asn Glu Ser Leu His Pro Pro Met Asp
                165                 170                 175

Asn Arg Val Arg Glu Ala Cys Gln Tyr Ile Ser Asp His Leu Ala Asp
            180                 185                 190

Ser Asn Phe Asp Ile Ala Ser Val Ala Gln His Val Cys Leu Ser Pro
        195                 200                 205

Ser Arg Leu Ser His Leu Phe Arg Gln Gln Leu Gly Ile Ser Val Leu
    210                 215                 220

Ser Trp Arg Glu Asp Gln Arg Ile Ser Gln Ala Lys Leu Leu Leu Ser
225                 230                 235                 240

Thr Thr Arg Met Pro Ile Ala Thr Val Gly Arg Asn Val Gly Phe Asp
                245                 250                 255

Asp Gln Leu Tyr Phe Ser Arg Val Phe Lys Lys Cys Thr Gly Ala Ser
            260                 265                 270

Pro Ser Glu Phe Arg Ala Gly Cys Glu Glu Lys Val Asn Asp Val Ala
        275                 280                 285

Val Lys Leu Ser
    290
```

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 861 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: bla

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:1..861
(D) OTHER INFORMATION:/product= "bla"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

```
ATG AGT ATT CAA CAT TTC CGT GTC GCC CTT ATT CCC TTT TTT GCG GCA        48
Met Ser Ile Gln His Phe Arg Val Ala Leu Ile Pro Phe Phe Ala Ala
        295                 300                 305

TTT TGC CTT CCT GTT TTT GCT CAC CCA GAA ACG CTG GTG AAA GTA AAA        96
Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr Leu Val Lys Val Lys
    310                 315                 320

GAT GCT GAA GAT CAG TTG GGT GCA CGA GTG GGT TAC ATC GAA CTG GAT       144
Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu Leu Asp
325                 330                 335                 340

CTC AAC AGC GGT AAG ATC CTT GAG AGT TTT CGC CCC GAA GAA CGT TTT       192
Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg Pro Glu Glu Arg Phe
                345                 350                 355

CCA ATG ATG AGC ACT TTT AAA GTT CTG CTA TGT GGC GCG GTA TTA TCC       240
Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala Val Leu Ser
            360                 365                 370

CGT GTT GAC GCC GGG CAA GAG CAA CTC GGT CGC CGC ATA CAC TAT TCT       288
Arg Val Asp Ala Gly Gln Glu Gln Leu Gly Arg Arg Ile His Tyr Ser
        375                 380                 385

CAG AAT GAC TTG GTT GAG TAC TCA CCA GTC ACA GAA AAG CAT CTT ACG       336
Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His Leu Thr
    390                 395                 400

GAT GGC ATG ACA GTA AGA GAA TTA TGC AGT GCT GCC ATA ACC ATG AGT       384
Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala Ala Ile Thr Met Ser
405                 410                 415                 420

GAT AAC ACT GCG GCC AAC TTA CTT CTG ACA ACG ATC GGA GGA CCG AAG       432
Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr Ile Gly Gly Pro Lys
                425                 430                 435

GAG CTA ACC GCT TTT TTG CAC AAC ATG GGG GAT CAT GTA ACT CGC CTT       480
Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp His Val Thr Arg Leu
            440                 445                 450

GAT CGT TGG GAA CCG GAG CTG AAT GAA GCC ATA CCA AAC GAC GAG CGT       528
Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg
        455                 460                 465

GAC ACC ACG ATG CCT GTA GCA ATG GCA ACA ACG TTG CGC AAA CTA TTA       576
Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr Leu Arg Lys Leu Leu
    470                 475                 480

ACT GGC GAA CTA CTT ACT CTA GCT TCC CGG CAA CAA TTA ATA GAC TGG       624
Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp
485                 490                 495                 500

ATG GAG GCG GAT AAA GTT GCA GGA CCA CTT CTG CGC TCG GCC CTT CCG       672
Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu Arg Ser Ala Leu Pro
                505                 510                 515

GCT GGC TGG TTT ATT GCT GAT AAA TCT GGA GCC GGT GAG CGT GGG TCT       720
Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly Glu Arg Gly Ser
            520                 525                 530
```

```
CGC GGT ATC ATT GCA GCA CTG GGG CCA GAT GGT AAG CCC TCC CGT ATC      768
Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile
        535                 540                 545

GTA GTT ATC TAC ACG ACG GGG AGT CAG GCA ACT ATG GAT GAA CGA AAT      816
Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr Met Asp Glu Arg Asn
    550                 555                 560

AGA CAG ATC GCT GAG ATA GGT GCC TCA CTG ATT AAG CAT TGG TAA          861
Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile Lys His Trp  *
565                 570                 575
```

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 287 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

```
Met Ser Ile Gln His Phe Arg Val Ala Leu Ile Pro Phe Phe Ala Ala
 1               5                  10                  15

Phe Cys Leu Pro Val Phe Ala His Pro Glu Thr Leu Val Lys Val Lys
            20                  25                  30

Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu Leu Asp
        35                  40                  45

Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg Pro Glu Glu Arg Phe
    50                  55                  60

Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala Val Leu Ser
65                  70                  75                  80

Arg Val Asp Ala Gly Gln Glu Gln Leu Gly Arg Arg Ile His Tyr Ser
                85                  90                  95

Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His Leu Thr
            100                 105                 110

Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala Ala Ile Thr Met Ser
        115                 120                 125

Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr Ile Gly Gly Pro Lys
    130                 135                 140

Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp His Val Thr Arg Leu
145                 150                 155                 160

Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg
                165                 170                 175

Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr Leu Arg Lys Leu Leu
            180                 185                 190

Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp
        195                 200                 205

Met Glu Ala Asp Lys Val Ala Gly Pro Leu Leu Arg Ser Ala Leu Pro
    210                 215                 220

Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly Glu Arg Gly Ser
```

```
225                 230                 235                 240

Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile
                245                 250                 255

Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr Met Asp Glu Arg Asn
            260                 265                 270

Arg Gln Ile Ala Glu Ile Gly Ala Ser Leu Ile Lys His Trp  *
        275                 280                 285
```

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7195 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: pBAD-ETgamma

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION:3588..4004
(D) OTHER INFORMATION:/product= "red gamma"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

```
ATCGATGCAT AATGTGCCTG TCAAATGGAC GAAGCAGGGA TTCTGCAAAC CCTATGCTAC     60
TCCGTCAAGC CGTCAATTGT CTGATTCGTT ACCAATTATG ACAACTTGAC GGCTACATCA    120
TTCACTTTTT CTTCACAACC GGCACGGAAC TCGCTCGGGC TGGCCCCGGT GCATTTTTTA    180
AATACCCGCG AGAAATAGAG TTGATCGTCA AAACCAACAT TGCGACCGAC GGTGGCGATA    240
GGCATCCGGG TGGTGCTCAA AAGCAGCTTC GCCTGGCTGA TACGTTGGTC CTCGCGCCAG    300
CTTAAGACGC TAATCCCTAA CTGCTGGCGG AAAAGATGTG ACAGACGCGA CGGCGACAAG    360
CAAACATGCT GTGCGACGCT GGCGATATCA AAATTGCTGT CTGCCAGGTG ATCGCTGATG    420
TACTGACAAG CCTCGCGTAC CCGATTATCC ATCGGTGGAT GGAGCGACTC GTTAATCGCT    480
TCCATGCGCC GCAGTAACAA TTGCTCAAGC AGATTTATCG CCAGCAGCTC CGAATAGCGC    540
CCTTCCCCTT GCCCGGCGTT AATGATTTGC CCAAACAGGT CGCTGAAATG CGGCTGGTGC    600
GCTTCATCCG GGCGAAAGAA CCCCGTATTG GCAAATATTG ACGGCCAGTT AAGCCATTCA    660
TGCCAGTAGG CGCGCGGACG AAAGTAAACC CACTGGTGAT ACCATTCGCG AGCCTCCGGA    720
TGACGACCGT AGTGATGAAT CTCTCCTGGC GGGAACAGCA AAATATCACC CGGTCGGCAA    780
ACAAATTCTC GTCCCTGATT TTTCACCACC CCCTGACCGC GAATGGTGAG ATTGAGAATA    840
TAACCTTTCA TTCCCAGCGG TCGGTCGATA AAAAAATCGA GATAACCGTT GGCCTCAATC    900
GGCGTTAAAC CCGCCACCAG ATGGGCATTA AACGAGTATC CCGGCAGCAG GGGATCATTT    960
TGCGCTTCAG CCATACTTTT CATACTCCCG CCATTCAGAG AAGAAACCAA TTGTCCATAT   1020
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TGCATCAGAC ATTGCCGTCA CTGCGTCTTT TACTGGCTCT TCTCGCTAAC CAAACCGGTA 1080

ACCCCGCTTA TTAAAAGCAT TCTGTAACAA AGCGGGACCA AAGCCATGAC AAAAACGCGT 1140

AACAAAAGTG TCTATAATCA CGGCAGAAAA GTCCACATTG ATTATTTGCA CGGCGTCACA 1200

CTTTGCTATG CCATAGCATT TTTATCCATA AGATTAGCGG ATCCTACCTG ACGCTTTTTA 1260

TCGCAACTCT CTACTGTTTC TCCATACCCG TTTTTTTGGG CTAGCAGGAG GAATTCACCA 1320

TGGATCCCGT AATCGTAGAA GACATAGAGC CAGGTATTTA TTACGGAATT TCGAATGAGA 1380

ATTACCACGC GGGTCCCGGT ATCAGTAAGT CTCAGCTCGA TGACATTGCT GATACTCCGG 1440

CACTATATTT GTGGCGTAAA AATGCCCCCG TGGACACCAC AAAGACAAAA ACGCTCGATT 1500

TAGGAACTGC TTTCCACTGC CGGGTACTTG AACCGGAAGA ATTCAGTAAC CGCTTTATCG 1560

TAGCACCTGA ATTTAACCGC CGTACAAACG CCGGAAAAGA AGAAGAGAAA GCGTTTCTGA 1620

TGGAATGCGC AAGCACAGGA AAAACGGTTA TCACTGCGGA AGAAGGCCGG AAAATTGAAC 1680

TCATGTATCA AAGCGTTATG GCTTTGCCGC TGGGGCAATG GCTTGTTGAA AGCGCCGGAC 1740

ACGCTGAATC ATCAATTTAC TGGGAAGATC CTGAAACAGG AATTTTGTGT CGGTGCCGTC 1800

CGGACAAAAT TATCCCTGAA TTTCACTGGA TCATGGACGT GAAAACTACG GCGGATATTC 1860

AACGATTCAA AACCGCTTAT TACGACTACC GCTATCACGT TCAGGATGCA TTCTACAGTG 1920

ACGGTTATGA AGCACAGTTT GGAGTGCAGC CAACTTTCGT TTTTCTGGTT GCCAGCACAA 1980

CTATTGAATG CGGACGTTAT CCGGTTGAAA TTTTCATGAT GGGCGAAGAA GCAAAACTGG 2040

CAGGTCAACA GGAATATCAC CGCAATCTGC GAACCCTGTC TGACTGCCTG AATACCGATG 2100

AATGGCCAGC TATTAAGACA TTATCACTGC CCCGCTGGGC TAAGGAATAT GCAAATGACT 2160

AGATCTCGAG GTACCCGAGC ACGTGTTGAC AATTAATCAT CGGCATAGTA TATCGGCATA 2220

GTATAATACG ACAAGGTGAG GAACTAAACC ATGGCTAAGC AACCACCAAT CGCAAAAGCC 2280

GATCTGCAAA AAACTCAGGG AAACCGTGCA CCAGCAGCAG TTAAAAATAG CGACGTGATT 2340

AGTTTTATTA ACCAGCCATC AATGAAAGAG CAACTGGCAG CAGCTCTTCC ACGCCATATG 2400

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TTGGCATTTT CTTTTATTTC TTATCAACAT AAAGGTGAAT CCCATACCTC GAGCTTCACG 3240

CTGCCGCAAG CACTCAGGGC GCAAGGGCTG CTAAAAGGAA GCGGAACACG TAGAAAGCCA 3300

GTCCGCAGAA ACGGTGCTGA CCCCGGATGA ATGTCAGCTA CTGGGCTATC TGGACAAGGG 3360

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TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA GAACGTTTTC CAATGATGAG 4740

CACTTTTAAA GTTCTGCTAT GTGGCGCGGT ATTATCCCGT GTTGACGCCG GGCAAGAGCA 4800

ACTCGGTCGC CGCATACACT ATTCTCAGAA TGACTTGGTT GAGTACTCAC CAGTCACAGA 4860

AAAGCATCTT ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 4920

TGATAACACT GCGGCCAACT TACTTCTGAC AACGATCGGA GGACCGAAGG AGCTAACCGC 4980

TTTTTTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT CGTTGGGAAC CGGAGCTGAA 5040

TGAAGCCATA CCAAACGACG AGCGTGACAC CACGATGCCT GTAGCAATGG CAACAACGTT 5100

GCGCAAACTA TTAACTGGCG AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG 5160

GATGGAGGCG GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5220

TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG CAGCACTGGG 5280

GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG ACGGGGAGTC AGGCAACTAT 5340

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GTCAGACCAA GTTTACTCAT ATATACTTTA GATTGATTTA CGCGCCCTGT AGCGGCGCAT 5460

TAAGCGCGGC GGGTGTGGTG GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG 5520

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AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTTAG TGCTTTACGG CACCTCGACC 5640

CCAAAAAACT TGATTTGGGT GATGGTTCAC GTAGTGGGCC ATCGCCCTGA TAGACGGTTT 5700

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TAACGTTTAC AATTTAAAAG GATCTAGGTG AAGATCCTTT TTGATAATCT CATGACCAAA 5940

ATCCCTTAAC GTGAGTTTTC GTTCCACTGA GCGTCAGACC CCGTAGAAAA GATCAAAGGA 6000

TCTTCTTGAG ATCCTTTTTT TCTGCGCGTA ATCTGCTGCT TGCAAACAAA AAAACCACCG 6060

CTACCAGCGG TGGTTTGTTT GCCGGATCAA GAGCTACCAA CTCTTTTTCC GAAGGTAACT 6120

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GCTGCTGCCA GTGGCGATAA GTCGTGTCTT ACCGGGTTGG ACTCAAGACG ATAGTTACCG 6300

GATAAGGCGC AGCGGTCGGG CTGAACGGGG GGTTCGTGCA CACAGCCCAG CTTGGAGCGA 6360

ACGACCTACA CCGAACTGAG ATACCTACAG CGTGAGCTAT GAGAAAGCGC CACGCTTCCC 6420

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CCTGCGTTAT CCCCTGATTC TGTGGATAAC CGTATTACCG CCTTTGAGTG AGCTGATACC 6720

GCTCGCCGCA GCCGAACGAC CGAGCGCAGC GAGTCAGTGA GCGAGGAAGC GGAAGAGCGC 6780

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CTGCGCCCCG ACACCCGCCA ACACCCGCTG ACGCGCCCTG ACGGGCTTGT CTGCTCCCGG 6900

CATCCGCTTA CAGACAAGCT GTGACCGTCT CCGGGAGCTG CATGTGTCAG AGGTTTTCAC 6960

CGTCATCACC GAAACGCGCG AGGCAGCAAG GAGATGGCGC CCAACAGTCC CCCGGCCACG 7020

GGGCCTGCCA CCATACCCAC GCCGAAACAA GCGCTCATGA GCCCGAAGTG GCGAGCCCGA 7080

TCTTCCCCAT CGGTGATGTC GGCGATATAG GCGCCAGCAA CCGCACCTGT GGCGCCGGTG 7140

ATGCCGGCCA CGATGCGTCC GGCGTAGAGG ATCTGCTCAT GTTTGACAGC TTATC 7195

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 7010 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: both
(D) TOPOLOGY: both

(vii) IMMEDIATE SOURCE:
(B) CLONE: pBAD-alpha-beta-gamma

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:1320..2000
(D) OTHER INFORMATION:/product= "red alpha"

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:2086..2871
(D) OTHER INFORMATION:/product= "red beta"

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION:3403..3819
(D) OTHER INFORMATION:/product= "red gamma"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

ATCGATGCAT AATGTGCCTG TCAAATGGAC GAAGCAGGGA TTCTGCAAAC CCTATGCTAC 60

TCCGTCAAGC CGTCAATTGT CTGATTCGTT ACCAATTATG ACAACTTGAC GGCTACATCA 120

TTCACTTTTT CTTCACAACC GGCACGGAAC TCGCTCGGGC TGGCCCCGGT GCATTTTTTA 180

AATACCCGCG AGAAATAGAG TTGATCGTCA AAACCAACAT TGCGACCGAC GGTGGCGATA 240

GGCATCCGGG TGGTGCTCAA AAGCAGCTTC GCCTGGCTGA TACGTTGGTC CTCGCGCCAG 300

CTTAAGACGC TAATCCCTAA CTGCTGGCGG AAAAGATGTG ACAGACGCGA CGGCGACAAG 360

CAAACATGCT GTGCGACGCT GGCGATATCA AAATTGCTGT CTGCCAGGTG ATCGCTGATG 420

TACTGACAAG CCTCGCGTAC CCGATTATCC ATCGGTGGAT GGAGCGACTC GTTAATCGCT 480

TCCATGCGCC GCAGTAACAA TTGCTCAAGC AGATTTATCG CCAGCAGCTC CGAATAGCGC 540

CCTTCCCCTT GCCCGGCGTT AATGATTTGC CCAAACAGGT CGCTGAAATG CGGCTGGTGC 600

GCTTCATCCG GGCGAAAGAA CCCCGTATTG GCAAATATTG ACGGCCAGTT AAGCCATTCA 660

TGCCAGTAGG CGCGCGGACG AAAGTAAACC CACTGGTGAT ACCATTCGCG AGCCTCCGGA 720

TGACGACCGT AGTGATGAAT CTCTCCTGGC GGGAACAGCA AAATATCACC CGGTCGGCAA 780

ACAAATTCTC GTCCCTGATT TTTCACCACC CCCTGACCGC GAATGGTGAG ATTGAGAATA 840

TAACCTTTCA TTCCCAGCGG TCGGTCGATA AAAAAATCGA GATAACCGTT GGCCTCAATC 900

GGCGTTAAAC CCGCCACCAG ATGGGCATTA AACGAGTATC CCGGCAGCAG GGGATCATTT 960

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TGCGCTTCAG CCATACTTTT CATACTCCCG CCATTCAGAG AAGAAACCAA TTGTCCATAT    1020

TGCATCAGAC ATTGCCGTCA CTGCGTCTTT TACTGGCTCT TCTCGCTAAC CAAACCGGTA    1080

ACCCCGCTTA TTAAAAGCAT TCTGTAACAA AGCGGGACCA AAGCCATGAC AAAAACGCGT    1140

AACAAAAGTG TCTATAATCA CGGCAGAAAA GTCCACATTG ATTATTTGCA CGGCGTCACA    1200

CTTTGCTATG CCATAGCATT TTTATCCATA AGATTAGCGG ATCCTACCTG ACGCTTTTTA    1260

TCGCAACTCT CTACTGTTTC TCCATACCCG TTTTTTTGGG CTAGCAGGAG GAATTCACC     1319

ATG ACA CCG GAC ATT ATC CTG CAG CGT ACC GGG ATC GAT GTG AGA GCT       1367
Met Thr Pro Asp Ile Ile Leu Gln Arg Thr Gly Ile Asp Val Arg Ala
      290                 295                 300

GTC GAA CAG GGG GAT GAT GCG TGG CAC AAA TTA CGG CTC GGC GTC ATC      1415
Val Glu Gln Gly Asp Asp Ala Trp His Lys Leu Arg Leu Gly Val Ile
  305                 310                 315

ACC GCT TCA GAA GTT CAC AAC GTG ATA GCA AAA CCC CGC TCC GGA AAG      1463
Thr Ala Ser Glu Val His Asn Val Ile Ala Lys Pro Arg Ser Gly Lys
320                 325                 330                 335

AAG TGG CCT GAC ATG AAA ATG TCC TAC TTC CAC ACC CTG CTT GCT GAG      1511
Lys Trp Pro Asp Met Lys Met Ser Tyr Phe His Thr Leu Leu Ala Glu
              340                 345                 350

GTT TGC ACC GGT GTG GCT CCG GAA GTT AAC GCT AAA GCA CTG GCC TGG      1559
Val Cys Thr Gly Val Ala Pro Glu Val Asn Ala Lys Ala Leu Ala Trp
          355                 360                 365

GGA AAA CAG TAC GAG AAC GAC GCC AGA ACC CTG TTT GAA TTC ACT TCC      1607
Gly Lys Gln Tyr Glu Asn Asp Ala Arg Thr Leu Phe Glu Phe Thr Ser
      370                 375                 380

GGC GTG AAT GTT ACT GAA TCC CCG ATC ATC TAT CGC GAC GAA AGT ATG      1655
Gly Val Asn Val Thr Glu Ser Pro Ile Ile Tyr Arg Asp Glu Ser Met
  385                 390                 395

CGT ACC GCC TGC TCT CCC GAT GGT TTA TGC AGT GAC GGC AAC GGC CTT      1703
Arg Thr Ala Cys Ser Pro Asp Gly Leu Cys Ser Asp Gly Asn Gly Leu
400                 405                 410                 415

GAA CTG AAA TGC CCG TTT ACC TCC CGG GAT TTC ATG AAG TTC CGG CTC      1751
Glu Leu Lys Cys Pro Phe Thr Ser Arg Asp Phe Met Lys Phe Arg Leu
              420                 425                 430

GGT GGT TTC GAG GCC ATA AAG TCA GCT TAC ATG GCC CAG GTG CAG TAC      1799
Gly Gly Phe Glu Ala Ile Lys Ser Ala Tyr Met Ala Gln Val Gln Tyr
          435                 440                 445

AGC ATG TGG GTG ACG CGA AAA AAT GCC TGG TAC TTT GCC AAC TAT GAC      1847
Ser Met Trp Val Thr Arg Lys Asn Ala Trp Tyr Phe Ala Asn Tyr Asp
      450                 455                 460

CCG CGT ATG AAG CGT GAA GGC CTG CAT TAT GTC GTG ATT GAG CGG GAT      1895
Pro Arg Met Lys Arg Glu Gly Leu His Tyr Val Val Ile Glu Arg Asp
  465                 470                 475

GAA AAG TAC ATG GCG AGT TTT GAC GAG ATC GTG CCG GAG TTC ATC GAA      1943
Glu Lys Tyr Met Ala Ser Phe Asp Glu Ile Val Pro Glu Phe Ile Glu
480                 485                 490                 495

AAA ATG GAC GAG GCA CTG GCT GAA ATT GGT TTT GTA TTT GGG GAG CAA      1991
Lys Met Asp Glu Ala Leu Ala Glu Ile Gly Phe Val Phe Gly Glu Gln
              500                 505                 510
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TGG CGA TAG ATCCGGTACC CGAGCACGTG TTGACAATTA ATCATCGGCA                2040
Trp Arg  *

TAGTATATCG GCATAGTATA ATACGACAAG GTGAGGAACT AAACC ATG AGT ACT          2094
                                                  Met Ser Thr
                                                    1

GCA CTC GCA ACG CTG GCT GGG AAG CTG GCT GAA CGT GTC GGC ATG GAT        2142
Ala Leu Ala Thr Leu Ala Gly Lys Leu Ala Glu Arg Val Gly Met Asp
  5                  10                  15

TCT GTC GAC CCA CAG GAA CTG ATC ACC ACT CTT CGC CAG ACG GCA TTT        2190
Ser Val Asp Pro Gln Glu Leu Ile Thr Thr Leu Arg Gln Thr Ala Phe
 20                  25                  30                  35

AAA GGT GAT GCC AGC GAT GCG CAG TTC ATC GCA TTA CTG ATC GTT GCC        2238
Lys Gly Asp Ala Ser Asp Ala Gln Phe Ile Ala Leu Leu Ile Val Ala
                 40                  45                  50

AAC CAG TAC GGC CTT AAT CCG TGG ACG AAA GAA ATT TAC GCC TTT CCT        2286
Asn Gln Tyr Gly Leu Asn Pro Trp Thr Lys Glu Ile Tyr Ala Phe Pro
           55                  60                  65

GAT AAG CAG AAT GGC ATC GTT CCG GTG GTG GGC GTT GAT GGC TGG TCC        2334
Asp Lys Gln Asn Gly Ile Val Pro Val Val Gly Val Asp Gly Trp Ser
         70                  75                  80

CGC ATC ATC AAT GAA AAC CAG CAG TTT GAT GGC ATG GAC TTT GAG CAG        2382
Arg Ile Ile Asn Glu Asn Gln Gln Phe Asp Gly Met Asp Phe Glu Gln
     85                  90                  95

GAC AAT GAA TCC TGT ACA TGC CGG ATT TAC CGC AAG GAC CGT AAT CAT        2430
Asp Asn Glu Ser Cys Thr Cys Arg Ile Tyr Arg Lys Asp Arg Asn His
100                 105                 110                 115

CCG ATC TGC GTT ACC GAA TGG ATG GAT GAA TGC CGC CGC GAA CCA TTC        2478
Pro Ile Cys Val Thr Glu Trp Met Asp Glu Cys Arg Arg Glu Pro Phe
                120                 125                 130

AAA ACT CGC GAA GGC AGA GAA ATC ACG GGG CCG TGG CAG TCG CAT CCC        2526
Lys Thr Arg Glu Gly Arg Glu Ile Thr Gly Pro Trp Gln Ser His Pro
           135                 140                 145

AAA CGG ATG TTA CGT CAT AAA GCC ATG ATT CAG TGT GCC CGT CTG GCC        2574
Lys Arg Met Leu Arg His Lys Ala Met Ile Gln Cys Ala Arg Leu Ala
         150                 155                 160

TTC GGA TTT GCT GGT ATC TAT GAC AAG GAT GAA GCC GAG CGC ATT GTC        2622
Phe Gly Phe Ala Gly Ile Tyr Asp Lys Asp Glu Ala Glu Arg Ile Val
     165                 170                 175

GAA AAT ACT GCA TAC ACT GCA GAA CGT CAG CCG GAA CGC GAC ATC ACT        2670
Glu Asn Thr Ala Tyr Thr Ala Glu Arg Gln Pro Glu Arg Asp Ile Thr
180                 185                 190                 195

CCG GTT AAC GAT GAA ACC ATG CAG GAG ATT AAC ACT CTG CTG ATC GCC        2718
Pro Val Asn Asp Glu Thr Met Gln Glu Ile Asn Thr Leu Leu Ile Ala
                200                 205                 210

CTG GAT AAA ACA TGG GAT GAC GAC TTA TTG CCG CTC TGT TCC CAG ATA        2766
Leu Asp Lys Thr Trp Asp Asp Asp Leu Leu Pro Leu Cys Ser Gln Ile
           215                 220                 225

TTT CGC CGC GAC ATT CGT GCA TCG TCA GAA CTG ACA CAG GCC GAA GCA        2814
Phe Arg Arg Asp Ile Arg Ala Ser Ser Glu Leu Thr Gln Ala Glu Ala
         230                 235                 240
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GTA AAA GCT CTT GGA TTC CTG AAA CAG AAA GCC GCA GAG CAG AAG GTG      2862
Val Lys Ala Leu Gly Phe Leu Lys Gln Lys Ala Ala Glu Gln Lys Val
    245                 250                 255

GCA GCA TAG ATCTCGAGAA GCTTCCTGCT GAACATCAAA GGCAAGAAAA             2911
Ala Ala  *
260

CATCTGTTGT CAAAGACAGC ATCCTTGAAC AAGGACAATT AACAGTTAAC AAATAAAAAC   2971

GCAAAAGAAA ATGCCGATAT CCTATTGGCA TTTTCTTTTA TTTCTTATCA ACATAAAGGT   3031

GAATCCCATA CCTCGAGCTT CACGCTGCCG CAAGCACTCA GGGCGCAAGG GCTGCTAAAA   3091

GGAAGCGGAA CACGTAGAAA GCCAGTCCGC AGAAACGGTG CTGACCCCGG ATGAATGTCA   3151

GCTACTGGGC TATCTGGACA AGGGAAAACG CAAGCGCAAA GAGAAAGCAG GTAGCTTGCA   3211

GTGGGCTTAC ATGGCGATAG CTAGACTGGG CGGTTTTATG GACAGCAAGC GAACCGGAAT   3271

TGCCAGCTGG GGCGCCCTCT GGTAAGGTTG GGAAGCCCTG CAAAGTAAAC TGGATGGCTT   3331

TCTTGCCGCC AAGGATCTGA TGGCGCAGGG GATCAAGATC TGATCAAGAG ACAGGATGAG   3391

GATCGTTTCG C ATG GAT ATT AAT ACT GAA ACT GAG ATC AAG CAA AAG CAT     3441
             Met Asp Ile Asn Thr Glu Thr Glu Ile Lys Gln Lys His
               1               5                  10

TCA CTA ACC CCC TTT CCT GTT TTC CTA ATC AGC CCG GCA TTT CGC GGG      3489
Ser Leu Thr Pro Phe Pro Val Phe Leu Ile Ser Pro Ala Phe Arg Gly
     15                  20                  25

CGA TAT TTT CAC AGC TAT TTC AGG AGT TCA GCC ATG AAC GCT TAT TAC      3537
Arg Tyr Phe His Ser Tyr Phe Arg Ser Ser Ala Met Asn Ala Tyr Tyr
 30                  35                  40                  45

ATT CAG GAT CGT CTT GAG GCT CAG AGC TGG GCG CGT CAC TAC CAG CAG      3585
Ile Gln Asp Arg Leu Glu Ala Gln Ser Trp Ala Arg His Tyr Gln Gln
                 50                  55                  60

CTC GCC CGT GAA GAG AAA GAG GCA GAA CTG GCA GAC GAC ATG GAA AAA      3633
Leu Ala Arg Glu Glu Lys Glu Ala Glu Leu Ala Asp Asp Met Glu Lys
             65                  70                  75

GGC CTG CCC CAG CAC CTG TTT GAA TCG CTA TGC ATC GAT CAT TTG CAA      3681
Gly Leu Pro Gln His Leu Phe Glu Ser Leu Cys Ile Asp His Leu Gln
         80                  85                  90

CGC CAC GGG GCC AGC AAA AAA TCC ATT ACC CGT GCG TTT GAT GAC GAT      3729
Arg His Gly Ala Ser Lys Lys Ser Ile Thr Arg Ala Phe Asp Asp Asp
     95                  100                 105

GTT GAG TTT CAG GAG CGC ATG GCA GAA CAC ATC CGG TAC ATG GTT GAA      3777
Val Glu Phe Gln Glu Arg Met Ala Glu His Ile Arg Tyr Met Val Glu
110                 115                 120                 125

ACC ATT GCT CAC CAC CAG GTT GAT ATT GAT TCA GAG GTA TAA              3819
Thr Ile Ala His His Gln Val Asp Ile Asp Ser Glu Val  *
                130                 135

AACGAGTAGA AGCTTGGCTG TTTTGGCGGA TGAGAGAAGA TTTTCAGCCT GATACAGATT   3879

AAATCAGAAC GCAGAAGCGG TCTGATAAAA CAGAATTTGC CTGGCGGCAG TAGCGCGGTG   3939

GTCCCACCTG ACCCCATGCC GAACTCAGAA GTGAAACGCC GTAGCGCCGA TGGTAGTGTG   3999

GGGTCTCCCC ATGCGAGAGT AGGGAACTGC CAGGCATCAA ATAAAACGAA AGGCTCAGTC   4059
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GAAAGACTGG GCCTTTCGTT TTATCTGTTG TTTGTCGGTG AACGCTCTCC TGAGTAGGAC 4119

AAATCCGCCG GGAGCGGATT TGAACGTTGC GAAGCAACGG CCCGGAGGGT GGCGGGCAGG 4179

ACGCCCGCCA TAAACTGCCA GGCATCAAAT TAAGCAGAAG GCCATCCTGA CGGATGGCCT 4239

TTTTGCGTTT CTACAAACTC TTTTGTTTAT TTTTCTAAAT ACATTCAAAT ATGTATCCGC 4299

TCATGAGACA ATAACCCTGA TAAATGCTTC AATAATATTG AAAAAGGAAG AGTATGAGTA 4359

TTCAACATTT CCGTGTCGCC CTTATTCCCT TTTTTGCGGC ATTTTGCCTT CCTGTTTTTG 4419

CTCACCCAGA AACGCTGGTG AAAGTAAAAG ATGCTGAAGA TCAGTTGGGT GCACGAGTGG 4479

GTTACATCGA ACTGGATCTC AACAGCGGTA AGATCCTTGA GAGTTTTCGC CCCGAAGAAC 4539

GTTTTCCAAT GATGAGCACT TTTAAAGTTC TGCTATGTGG CGCGGTATTA TCCCGTGTTG 4599

ACGCCGGGCA AGAGCAACTC GGTCGCCGCA TACACTATTC TCAGAATGAC TTGGTTGAGT 4659

ACTCACCAGT CACAGAAAAG CATCTTACGG ATGGCATGAC AGTAAGAGAA TTATGCAGTG 4719

CTGCCATAAC CATGAGTGAT AACACTGCGG CCAACTTACT TCTGACAACG ATCGGAGGAC 4779

CGAAGGAGCT AACCGCTTTT TTGCACAACA TGGGGGATCA TGTAACTCGC CTTGATCGTT 4839

GGGAACCGGA GCTGAATGAA GCCATACCAA ACGACGAGCG TGACACCACG ATGCCTGTAG 4899

CAATGGCAAC AACGTTGCGC AAACTATTAA CTGGCGAACT ACTTACTCTA GCTTCCCGGC 4959

AACAATTAAT AGACTGGATG GAGGCGGATA AAGTTGCAGG ACCACTTCTG CGCTCGGCCC 5019

TTCCGGCTGG CTGGTTTATT GCTGATAAAT CTGGAGCCGG TGAGCGTGGG TCTCGCGGTA 5079

TCATTGCAGC ACTGGGGCCA GATGGTAAGC CCTCCCGTAT CGTAGTTATC TACACGACGG 5139

GGAGTCAGGC AACTATGGAT GAACGAAATA GACAGATCGC TGAGATAGGT GCCTCACTGA 5199

TTAAGCATTG GTAACTGTCA GACCAAGTTT ACTCATATAT ACTTTAGATT GATTTACGCG 5259

CCCTGTAGCG GCGCATTAAG CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA 5319

CTTGCCAGCG CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC CTTCCTTTCT CGCCACGTTC 5379

GCCGGCTTTC CCCGTCAAGC TCTAAATCGG GGGCTCCCTT TAGGGTTCCG ATTTAGTGCT 5439

TTACGGCACC TCGACCCCAA AAAACTTGAT TTGGGTGATG GTTCACGTAG TGGGCCATCG 5499

CCCTGATAGA CGGTTTTTCG CCCTTTGACG TTGGAGTCCA CGTTCTTTAA TAGTGGACTC 5559

TTGTTCCAAA CTTGAACAAC ACTCAACCCT ATCTCGGGCT ATTCTTTTGA TTTATAAGGG 5619

ATTTTGCCGA TTTCGGCCTA TTGGTTAAAA AATGAGCTGA TTTAACAAAA ATTTAACGCG 5679

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TAATCTCATG ACCAAAATCC CTTAACGTGA GTTTTCGTTC CACTGAGCGT CAGACCCCGT 5799

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AATCCTGTTA CCAGTGGCTG CTGCCAGTGG CGATAAGTCG TGTCTTACCG GGTTGGACTC 6099

AAGACGATAG TTACCGGATA AGGCGCAGCG GTCGGGCTGA ACGGGGGGTT CGTGCACACA 6159

GCCCAGCTTG GAGCGAACGA CCTACACCGA ACTGAGATAC CTACAGCGTG AGCTATGAGA 6219

AAGCGCCACG CTTCCCGAAG GGAGAAAGGC GGACAGGTAT CCGGTAAGCG GCAGGGTCGG 6279

AACAGGAGAG CGCACGAGGG AGCTTCCAGG GGGAAACGCC TGGTATCTTT ATAGTCCTGT 6339

CGGGTTTCGC CACCTCTGAC TTGAGCGTCG ATTTTTGTGA TGCTCGTCAG GGGGGCGGAG 6399

CCTATGGAAA AACGCCAGCA ACGCGGCCTT TTTACGGTTC CTGGCCTTTT GCTGGCCTTT 6459

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GGAAGCGGAA GAGCGCCTGA TGCGGTATTT TCTCCTTACG CATCTGTGCG GTATTTCACA 6639

CCGCATAGGG TCATGGCTGC GCCCCGACAC CCGCCAACAC CCGCTGACGC GCCCTGACGG 6699

GCTTGTCTGC TCCCGGCATC CGCTTACAGA CAAGCTGTGA CCGTCTCCGG GAGCTGCATG 6759

TGTCAGAGGT TTTCACCGTC ATCACCGAAA CGCGCGAGGC AGCAAGGAGA TGGCGCCCAA 6819

CAGTCCCCCG GCCACGGGGC CTGCCACCAT ACCCACGCCG AAACAAGCGC TCATGAGCCC 6879

GAAGTGGCGA GCCCGATCTT CCCCATCGGT GATGTCGGCG ATATAGGCGC CAGCAACCGC 6939

ACCTGTGGCG CCGGTGATGC CGGCCACGAT GCGTCCGGCG TAGAGGATCT GCTCATGTTT 6999

GACAGCTTAT C 7010

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 227 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

```
Met Thr Pro Asp Ile Ile Leu Gln Arg Thr Gly Ile Asp Val Arg Ala
 1               5                  10                  15

Val Glu Gln Gly Asp Asp Ala Trp His Lys Leu Arg Leu Gly Val Ile
            20                  25                  30

Thr Ala Ser Glu Val His Asn Val Ile Ala Lys Pro Arg Ser Gly Lys
        35                  40                  45

Lys Trp Pro Asp Met Lys Met Ser Tyr Phe His Thr Leu Leu Ala Glu
    50                  55                  60

Val Cys Thr Gly Val Ala Pro Glu Val Asn Ala Lys Ala Leu Ala Trp
65                  70                  75                  80

Gly Lys Gln Tyr Glu Asn Asp Ala Arg Thr Leu Phe Glu Phe Thr Ser
                85                  90                  95

Gly Val Asn Val Thr Glu Ser Pro Ile Ile Tyr Arg Asp Glu Ser Met
            100                 105                 110

Arg Thr Ala Cys Ser Pro Asp Gly Leu Cys Ser Asp Gly Asn Gly Leu
        115                 120                 125
```

```
Glu Leu Lys Cys Pro Phe Thr Ser Arg Asp Phe Met Lys Phe Arg Leu
    130                 135                 140

Gly Gly Phe Glu Ala Ile Lys Ser Ala Tyr Met Ala Gln Val Gln Tyr
145                 150                 155                 160

Ser Met Trp Val Thr Arg Lys Asn Ala Trp Tyr Phe Ala Asn Tyr Asp
                165                 170                 175

Pro Arg Met Lys Arg Glu Gly Leu His Tyr Val Val Ile Glu Arg Asp
            180                 185                 190

Glu Lys Tyr Met Ala Ser Phe Asp Glu Ile Val Pro Glu Phe Ile Glu
        195                 200                 205

Lys Met Asp Glu Ala Leu Ala Glu Ile Gly Phe Val Phe Gly Glu Gln
    210                 215                 220

Trp Arg  *
225
```

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 262 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

```
Met Ser Thr Ala Leu Ala Thr Leu Ala Gly Lys Leu Ala Glu Arg Val
 1               5                  10                  15

Gly Met Asp Ser Val Asp Pro Gln Glu Leu Ile Thr Thr Leu Arg Gln
            20                  25                  30

Thr Ala Phe Lys Gly Asp Ala Ser Asp Ala Gln Phe Ile Ala Leu Leu
        35                  40                  45

Ile Val Ala Asn Gln Tyr Gly Leu Asn Pro Trp Thr Lys Glu Ile Tyr
    50                  55                  60

Ala Phe Pro Asp Lys Gln Asn Gly Ile Val Pro Val Val Gly Val Asp
65                  70                  75                  80

Gly Trp Ser Arg Ile Ile Asn Glu Asn Gln Gln Phe Asp Gly Met Asp
                85                  90                  95

Phe Glu Gln Asp Asn Glu Ser Cys Thr Cys Arg Ile Tyr Arg Lys Asp
            100                 105                 110

Arg Asn His Pro Ile Cys Val Thr Glu Trp Met Asp Glu Cys Arg Arg
        115                 120                 125

Glu Pro Phe Lys Thr Arg Glu Gly Arg Glu Ile Thr Gly Pro Trp Gln
    130                 135                 140

Ser His Pro Lys Arg Met Leu Arg His Lys Ala Met Ile Gln Cys Ala
145                 150                 155                 160

Arg Leu Ala Phe Gly Phe Ala Gly Ile Tyr Asp Lys Asp Glu Ala Glu
                165                 170                 175
```

```
Arg Ile Val Glu Asn Thr Ala Tyr Thr Ala Glu Arg Gln Pro Glu Arg
            180                 185                 190

Asp Ile Thr Pro Val Asn Asp Glu Thr Met Gln Glu Ile Asn Thr Leu
        195                 200                 205

Leu Ile Ala Leu Asp Lys Thr Trp Asp Asp Asp Leu Leu Pro Leu Cys
    210                 215                 220

Ser Gln Ile Phe Arg Arg Asp Ile Arg Ala Ser Ser Glu Leu Thr Gln
225                 230                 235                 240

Ala Glu Ala Val Lys Ala Leu Gly Phe Leu Lys Gln Lys Ala Ala Glu
                245                 250                 255

Gln Lys Val Ala Ala  *
            260
```

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 139 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

```
Met Asp Ile Asn Thr Glu Thr Glu Ile Lys Gln Lys His Ser Leu Thr
 1               5                  10                  15

Pro Phe Pro Val Phe Leu Ile Ser Pro Ala Phe Arg Gly Arg Tyr Phe
            20                  25                  30

His Ser Tyr Phe Arg Ser Ser Ala Met Asn Ala Tyr Tyr Ile Gln Asp
        35                  40                  45

Arg Leu Glu Ala Gln Ser Trp Ala Arg His Tyr Gln Gln Leu Ala Arg
    50                  55                  60

Glu Glu Lys Glu Ala Glu Leu Ala Asp Asp Met Glu Lys Gly Leu Pro
65                  70                  75                  80

Gln His Leu Phe Glu Ser Leu Cys Ile Asp His Leu Gln Arg His Gly
                85                  90                  95

Ala Ser Lys Lys Ser Ile Thr Arg Ala Phe Asp Asp Asp Val Glu Phe
            100                 105                 110

Gln Glu Arg Met Ala Glu His Ile Arg Tyr Met Val Glu Thr Ile Ala
        115                 120                 125

His His Gln Val Asp Ile Asp Ser Glu Val  *
    130                 135
```

Table 1: Sequences of Oligos for PCR

Figure 3ab
left: TGACCCCTCACAAGGAGACGACCTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
right: TACAAATGTGGTATGGCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
template: pJP5603
targeting vector: pSV-paz1 1

Figure 3c
a-left: CTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
a-right: ATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
b-left: AGACGACCTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
b-right: GCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
c-left: CACAAGGAGACGACCTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
c-right: TGGTATGGCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
d-left: TGACCCCTCACAAGGAGACGACCTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
d-right: TACAAATGTGGTATGGCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
e-left:
CACGCCCCTGACCCCTCACAAGGAGACGACCTTCCATGACCGAGTACAAGAGGGATGTAACGCACTGA
e-right:
TAAAACCTCTACAAATGTGGTATGGCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCCCGCTTTCCA
f-left:
TCCCCTGACCCACGCCCCTGACCCCTCACAAGGAGACGACCTTCCATGACCGAGTACAAGAGGGATGT
AACGCACTGA
f-right:
TAAAGCAAGTAAAACCTCTACAAATGTGGTATGGCTGATTATGATCCTCTAGAGTCGGTGCTCACTGCC
CGCTTTCCA
template: pJP5603
targeting vector: pSV-paz1 1

Figure 3d
a-left:
TCATCCTCTGCATGGTCAGGTCATGGATGAGCAGACGATGGTGCAGGATCAAGGGCTGCTAAAGGAA
a-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
b-left:
CACGAGCATCATCCTCTGCATGGTCAGGTCATGGATGAGCAGACGATGGCAAGGGCTGCTAAAGGAA
b-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
c-left:
TTAACCGTCACGAGCATCATCCTCTGCATGGTCAGGTCATGGATGAGCACAAGGGCTGCTAAAGGAA
c-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
d-left:
TGCTGCTGAACGGCAAGCCGTTGCTGATTCGAGGCGTTAACCGTCACGACAAGGGCTGCTAAAGGAA
d-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
e-left:
TCTCTATCGTGCGGTGGTTGAACTGCACACCGCCGACGGCACGCTGATTCAAGGGCTGCTAAAGGAA
e-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
f-left:
TGGAGTGACGGCAGTTATCTGGAAGATCAGGATATGTGGCGGATGAGCGCAAGGGCTGCTAAAGGAA

f-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
g-left:
TGCATTCTAGTTGTGGTTTGTCCAAACTCATCAATGTATCTTATCATGTCAAGGGCTGCTAAAGGAA
g-right:
TAATGCGAACAGCGCACGGCGTTAAAGTTGTTCTGCTTCATCAGCAGGATGGCGAAGAACTCCAGCAT
h-left:
TGCATTCTAGTTGTGGTTTGTCCAAACTCATCAATGTATCTTATCATGTCAAGGGCTGCTAAAGGAA
h-right:
TATTTTTGACACCAGACCAACTGGTAATGGTAGCGACCGGCGCTCAGCTGGCGAAGAACTCCAGCAT
template: pJP5603
targeting vector: pSV-paz11

Figure 4
left:
TCAAGTGAGAAATCACCATGAGTGACGACTGAATCCGGTGAGAATGGCAAAAGCTTATGCCCACCAGC
TGGTATGGCTGATTATGATC
right:
TCCAACATGGATGCTGATTTATATGGGTATAAATGGGCTCGCGATAATGTCGGGCAATCAGGTGCGACA
ATCTACCACCAGCTCTTTTCTACGGGGTCTGACGC
template: pBR322
targeting vector: Hoxa-P1

Figure 5
left:
TGCAGCTGGCACGACAGGTTTCCCGACTGGAAAGCGGGCAGTTAATACGACTCACTATAGGGAGAACA
GGAAACAGCTATGCCCATAACACCCAGAGTA
right:
TGCGCCGCTACAGGGCGCGTCCATTCGCCATTCAGGCCTGACTCACTAGTGATGGTGATGGTGATGTGG
GGGGTGCCGCTCAGT
template: pmtrx (a pBluescipt vector carrying mouse trithorax cDNA)
targeting vector: pZero2.1

Figure 6
left:
TGAGACAATAACCCTGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGGAGAAAAAAATCACT
GGATATACCACCG
right:
TACAGGGCGCGTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACGCCCCGCCCTGC
CACTCATCGCA
template: pMAK705
targeting vector: pBAD-24 backbone Amp resistant gene

Figure 8
i:
TGCCAAGCTTGACCCACTGTGGAAGTGTTCCAAAAAGCGGGAAGGCTCTTGAGCTACTTCACTAACAAC
CGG
g:
TCACCATCTTCGGGCCATTTGTAGACTGGAATATTTCGAGCTATGAGTGTGCTACTTCACTAACAACCG
G
h:
TGGCCCCAGGGTGACGCGGACATGGAGTTGTCGCCAGGGCACTGGTCCATGAGAGTGCCAAGCTACTC
GCGAC
template: pKaZ
targeting vector: Hoxa-P1

Figure 9
j:
TAATAGCGAAGAGGCCCGCACCGATCGCCCTTCCCAACAGTTGCGCAGCCTGAATGGCGAATGGCGCT
TTGCCTGGTTTATAACTTCGTATAGCATACATTATACGAAGTTATGGGCTGCTAAAGGAAGCGGAACAC
G
k:
TGGCAGTTCAGGCCAATCCGCGCCGGATGCGGTGTATCGCTCGCCACTTCAACATCAACGGTAATCGCC
ATTTGACCATATAACTTCGTATAATGTATGCTATACGAAGTTATCCCCAGAGTCCCGCTCAGAAGAACT
template: pJP5603
targeting vector: JC9604 chromosome

Figure 10
l:
TAGCTTGGCACTGGCCGTCGTTTTACAACGTCGTGACTGGGAAAACCCTGGCGTTACCCAACCCATCAC
ATATACCTGCCGTTCACTAT
m:
TATCGGTGGCCGTGGTGTCGGCTCCGCCGCCTTCATACTGCACCGGGCGGGAAGGCGATTCCGAAGCCC
AACCTTTCATAGAAGCC
template: pIB279
targeting vector: pSV-paX1

l*: GCTTGGCACTGGCCGTCGTTTTACAACGTCGTGACTGGGAA
m*:
TCGGTGGCCGTGGTGTCGGCTCCGCCGCCTTCATACTGCACCGGGCGGGAAGGATCCACAGATTTGATC
CAGCGATACAGC
template: pSV-paz11
targeting vector: pSV-sacB-neo

Figure11
n:
TACCGCATTAAAGCTTATCGATGATAAGCTGTCAAACATGAGAATTGACCCGGAACCCTTCTCGAGGAA
GTTCCTATTCTCTAGAAAGTATAGGAACTTCCGAATAAATACCTGTGACGGAAGATCACTT
p:
TTCCCTCAAGAATTTTACTCTGTCAGAAACGGCCTTAACGACGTAGTCGAGGGACCTAGAAGTTCCTAT
ACTTTCTAGAGAATAGGAACTTCATTATCACTTATTCAGGCGTAGCACCAGGCG
template: pMAK705
targeting vector: Hoxa-P1

Figure 12
left:
TGAGACAATAACCCTGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGGAGAAAAAAATCACT
GGATATACCACCG
right:
TACAGGGCGCGTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACGCCCCGCCCTGC
CACTCATCGCA
template: pMAK705
targeting vector: pBAD-24 backbone Amp resistant gene

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(74) Agents: WEICKMANN, H. et al.; Kopernikusstrasse 9, D–81679 München (DE).

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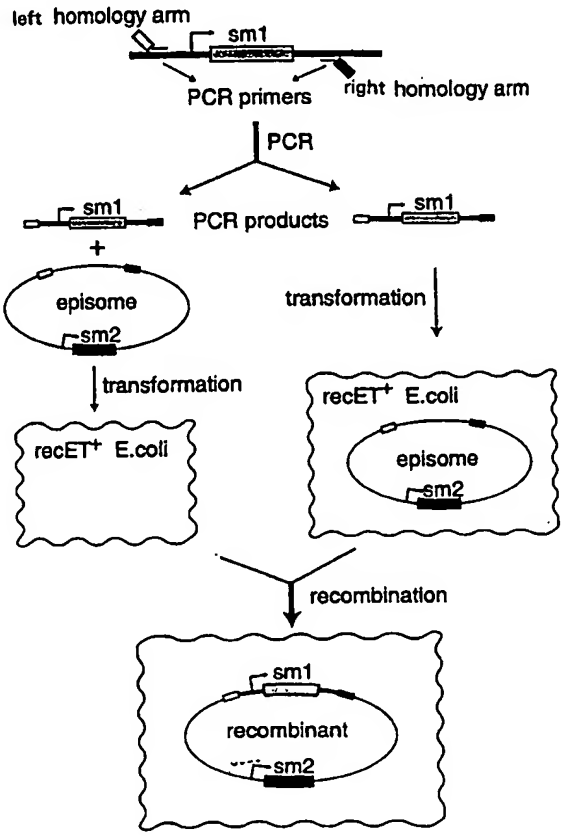
(57) Abstract

The invention refers to a novel method for cloning DNA molecules using a homologous recombination mechanism between at least two DNA molecules comprising: a) providing a host cell capable of performing homologous recombination, b) contacting in said host cell a first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred. In particular, it relies on the use of the E. coli RecE and RecT proteins, the bacteriophage Red-alpha and Red-beta proteins, or the phage P22 recombination system. The beneficial effects of concomitant expression of the RecBC inhibitor genes (e.g. Red-Gamma) is also examplified.



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Minimum documentation searched (classification system followed by classification symbols)
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C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	OLINER J.D. ET AL.: "In vivo Cloning of PCR Products in E. coli" NUCLEIC ACIDS RESEARCH, vol. 21, no. 22, 1993, pages 5192-5197, XP002064297 cited in the application	1-6, 8-13,19, 20, 22-27, 34,35, 37-42, 46-48,50
Y	See page 5192, column 2, line 8 - line 31, and discussion section	18,28

Y	NUSSBAUM A ET AL: "Restriction-stimulated homologous recombination of plasmids by the RecE pathway of Escherichia coli" GENETICS, vol. 130, no. 1, January 1992, pages 37-49, XP002103370 see figures 1,7	18,28

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[X] Further documents are listed in the continuation of box C. [ ] Patent family members are listed in annex.

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21 May 1999	04/06/1999
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Lonnoy, O

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	ZHANG Y ET AL: "A new logic for DNA engineering using recombination in Escherichia coli" NATURE GENETICS, vol. 20, no. 2, October 1998, pages 123-128, XP002103371 see the whole document	1-50
A	KOLODNER R ET AL: "Homologous pairing proteins encoded by the Escherichia coli recE and recT genes" MOLECULAR MICROBIOLOGY, vol. 11, no. 1, 1994, pages 23-30, XP002064301 cited in the application	
A	LUISI-DELUCA C ET AL: "Genetic and physical analysis of plasmid recombination in recB recC sbcB and recB recC sbcA Escherichia coli K-12 mutants" GENETICS, vol. 122, 1989, pages 269-278, XP002064305	
A	DEGRYSE E: "Evaluation of Escherichia coli recBC sbcBC mutants for cloning by recombination in vivo" JOURNAL OF BIOTECHNOLOGY, vol. 2, no. 39, 15 April 1995, page 181-187 XP004036984	
A	YANG X ET AL: "Homologous recombination based modification in Escherichia coli and germ line transmission in transgenic mice of a bacterial artificial chromosome" NATURE BIOTECHNOLOGY, vol. 15, September 1997, pages 859-865, XP002103372	
A	MURPHY K: "Lambda Gam protein inhibits the helicase and Chi-stimulated recombination activities of Escherichia coli RecBCD enzyme" JOURNAL OF BACTERIOLOGY, vol. 173, no. 18, September 1991, pages 5808-5821, XP002103373	

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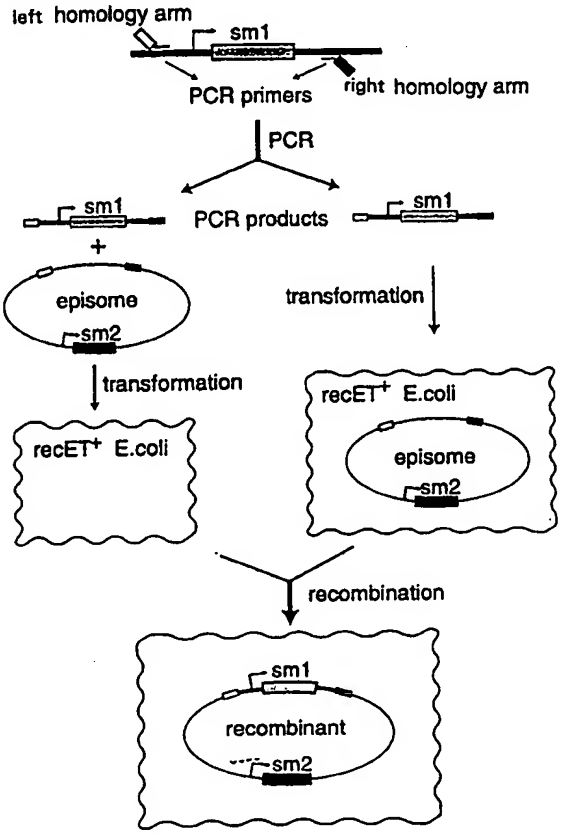
(54) Title: NOVEL DNA CLONING METHOD RELYING ON THE E. COLI RECE/RECT RECOMBINATION SYSTEM

(57) Abstract

The invention refers to a novel method for cloning DNA molecules using a homologous recombination mechanism between at least two DNA molecules comprising: a) providing a host cell capable of performing homologous recombination, b) contacting in said host cell a first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred. In particular, it relies on the use of the E. coli RecE and RecT proteins, the bacteriophage Red-alpha and Red-beta proteins, or the phage P22 recombination system. The beneficial effects of concomitant expression of the RecBC inhibitor genes (e.g. Red-Gamma) is also examplified.



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AMENDED CLAIMS

[received by the International Bureau on 04 August 1999 (04.08.99);
original claims 1-50 replaced by new claims 1-64 (10 pages)]

1. A method for cloning DNA molecules in procaryotic cells comprising the steps of:

   a) providing a procaryotic host cell capable of performing homologous recombination,

   b) contacting in said host cell a circular first DNA molecule which is capable of being replicated in said host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DNA molecules and

   c) selecting a host cell in which homologous recombination between said first and second DNA molecules has occurred.

2. The method according to claim 1 wherein the homologous recombination occurs via the recET cloning mechanism.

3. The method according to claim 2 wherein the host cell is capable of expressing recE and recT genes.

4. The method according to claim 3 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

5. The method according to claim 3 or 4 wherein the host cell is transformed with at least one vector capable of expressing recE and/or recT genes.

6. The method of claim 3, 4 or 5 wherein the expression of the recE and/or recT genes is under control of a regulatable promoter.

7. The method of claim 5 or 6 wherein the recT gene is overexpressed versus the recE gene.

8. The method according to any one of claims 3 to 7 wherein the recE gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 1320 (ATG) to 2159 (GAC) as depicted in Fig.7B,
(b) the nucleic acid sequence from position 1320 (ATG) to 1998 (CGA) as depicted in Fig.13B,
(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequence from (a), (b) and/or (c).

9. The method according to any one of claims 3 to 8 wherein the recT gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 2155 (ATG) to 2961 (GAA) as depicted in Fig.7B,
(b) the nucleic acid sequence from position 2086 (ATG) to 2868 (GCA) as depicted in Fig.13B,
(c) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(d) a nucleic acid sequence which hybridizes under stringent conditions with the nucleic acid sequences from (a), (b) and/or (c).

10. The method according to any one of the previous claims wherein the host cell is a gram-negative bacterial cell.

11. The method according to claim 10 wherein the host cell is an Escherichia coli cell.

12. The method according to claim 11 wherein the host cell is an Escherichia coli K12 strain.

13. The method according to claim 12 wherein the E.coli strain is selected from JC 8679 and JC 9604.

14. The method according to any one of the previous claims wherein the host cell further is capable of expressing a recBC inhibitor gene.

15. The method according to claim 14 wherein the host cell is transformed with a vector expressing the recBC inhibitor gene.

16. The method according to claim 14 or 15 wherein the recBC inhibitor gene is selected from a nucleic acid molecule comprising
(a) the nucleic acid sequence from position 3588 (ATG) to 4002 (GTA) as depicted in Fig.13B,
(b) a nucleic acid encoding the same polypeptide within the degeneracy of the genetic code and/or
(c) a nucleic acid sequence which hybridizes under stringent conditions (as defined above) with the nucleic acid sequence from (a) and/ or (b).

17. The method according to any one of claims 13 to 16 wherein the host cell is a prokaryotic recBC+ cell.

18. The method according to any one of the previous claims wherein the first DNA molecule is an extrachromosomal DNA molecule containing an origin of replication which is operative in the host cell.

19. The method according to claim 18 wherein the first DNA molecule is selected from plasmids, cosmids, P1 vectors, BAC vectors and PAC vectors.

20. The method according to any one of claims 1-18 wherein the first DNA molecule is a host cell chromosome.

21. The method according to any one of the previous claims wherein the second DNA molecule is linear.

22. The method according to any one of the previous claims wherein the regions of sequence homology are at least 15 nucleotides each.

23. The method according to one of claims 1 to 16 wherein the second DNA molecule is obtained by an amplification reaction.

24. The method according to one of the previous claims wherein the first and/or second DNA molecules are introduced into the host cells by transformation.

25. The method according to claim 24 wherein the transformation method is electroporation.

26. The method according to one of claims 1 to 25 wherein the first and second DNA molecules are introduced into the host cell simultaneously by co-transformation.

27. The method according to one of claims 1 to 25 wherein the second DNA molecule is introduced into a host cell in which the first DNA molecule is already present.

28. The method according to one of the previous claims wherein the second DNA molecule contains at least one marker gene placed between the two regions of sequence homology and wherein homologous recombination is detected by expression of said marker gene.

29. The method according to claim 28 wherein gene presence is selected from antibiotic resistance genes, deficiency complementation genes and reporter genes.

30. The method of any one of claims 1 to 29 wherein the first DNA molecule contains at least one marker gene between the two regions of sequence homology and wherein homologous recombination is detected by lack of expression of said marker gene.

31. The method of any one of claims 1 to 30 wherein said marker gene is selected from genes which, under selected conditions, convey a toxic or bacteriostatic effect on the cell, and reporter genes.

32. A method according to any one of the previous claims wherein the first DNA molecule contains at least one target site for a site specific recombinase between the two regions of sequence homology and wherein homologous recombination is detected by removal of said target site.

33. A method for cloning DNA molecules comprising the steps of:
(a) providing a source of RecE and RecT proteins,
(b) contacting a first DNA molecule which is capable of being replicated in a suitable host cell with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, under conditions which favour homologous recombination between said first and second DMA molecules and
(c) selecting DNA molecules in which homologous recombination between said first and second DNA molecules has occurred.

34. The method of claim 33 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

35. The method of claim 33 or 34 wherein the recombination occurs in vitro.

36. The method of claim 33 or 34 wherein the recombination occurs in vivo.

37. A method for making a recombinant DNA molecule comprising introducing into a prokaryotic host cell a circular first DNA molecule which is capable of being replicated in said host cell, and introducing a second DNA molecule comprising a first and a second region of sequence homology to a third and fourth region, respectively, on the first DNA molecule, said host cell being capable of performing homologous recombination, such that a recombinant DNA molecule is made, said recombinant DNA molecule comprising the first DNA molecule wherein the sequences between said third and fourth regions have been replaced by sequences between the first and second regions of the second DNA molecule.

38. The method according to claim 37 which further comprises detecting the recombinant DNA molecule.

39. A method for making a recombinant DNA molecule comprising introducing into a prokaryotic host cell, containing a chromosomal first DNA molecule, a second DNA molecule comprising a first and a second region of sequence homology to a third and a fourth region, respectively, on the host chromosomal first DNA molecule, said host cell being capable of performing homologous recombination, such that a recombinant DNA molecule is made, said recombinant DNA molecule comprising the chromosomal first DNA molecule wherein the sequences between said third and fourth regions have been replaced by sequences between the first and second regions of the second DNA molecule.

40. The method according to claim 39 which further comprises detecting the recombinant DNA molecule.

41. The method according to any one of claims 37 to 40, wherein the host cell is capable of expressing RecE and RecT proteins or λexo and λß proteins.

42. A method for cloning DNA molecules comprising the steps of:
(a) contacting in vitro a first DNA molecule with a second DNA molecule comprising at least two regions of sequence homology to regions on the first DNA molecule, in the presence of RecE and RecT proteins and under conditions which favour homologous recombination between said first and second DNA molecules; and
(b) selecting a DNA molecule in which homologous recombination between said first and second DNA molecules has occurred.

43. A method for making a recombinant DNA molecule comprising contacting in vitro a first DNA molecule with a second DNA molecule comprising a first and a second region of sequence homology to a third and a fourth region on the first DNA molecule, in the presence of RecE and RecT proteins and under conditions in which homologous recombination can occur, such that a recombinant DNA molecule is made, said recombinant DNA molecule comprising the first DNA molecule wherein the sequences between said third and fourth regions have been replaced by sequences between the first and second regions of the second DNA molecule.

44. The method of claim 42, which further comprises between steps (a) and (b) the step of introducing the product step (a) into a cell, wherein recombination occurs in the cell.

45. Use of cells capable of expressing the recE and recT genes as a host cell for a cloning method involving homologous recombination.

46. Use of a vector system capable of expressing recE and recT genes in a host cell for a cloning method involving homologous recombination.

47. Use of claims 45 or 46 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

48. Use of a source of RecE and RecT proteins for a cloning method involving homologous recombination.

49. Use of claim 48 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

50. A reagent kit for cloning comprising
(a) a host cell
(b) means of expressing recE and recT genes in said host cell and
(c) a recipient cloning vehicle capable of being replicated in said cell.

51. The reagent kit according to claim 50 wherein the means (b) comprise a vector system capable of expressing the recE and recT genes in the host cell.

52. The reagent kit according to claim 50 or 51 wherein the recE and recT genes are selected from E.coli recE and recT genes or from λ redα and redß genes.

53. A reagent kit for cloning comprising
(a) a source for RecE and RecT proteins and

(b) a recipient cloning vehicle capable of being propagated in a host cell.

54. The reagent kit according to claim 53 further comprising a host cell suitable for propagating said recipient cloning vehicle.

55. The reagent kit according to claim 53 or 54 wherein said RecE and RecT or proteins are selected from E.coli RecE and RecT proteins or from phage λ Redα and Redß proteins.

56. The reagent kit according to any one of claims 50-55 further comprising means for expressing a site specific recombinase in said host cell.

57. The reagent kit according to any one of claims 50-56 further comprising nucleic acid amplification primers comprising a region of homology to said recipient cloning vehicle.

58. A reagent kit for cloning comprising first and second DNA amplification primers and a recipient cloning vehicle that is a circular DNA molecule, said first DNA amplification primer having a first region of sequence homology to a third region on the circular recipient cloning vehicle, and said second DNA amplification primer having a second region of sequence homology to a fourth region on the circular recipient cloning vehicle.

59. The reagent kit of claim 58, further comprising a prokaryotic host cell that is capable of performing homologous recombination.

60. The reagent kit of claim 58 or 59, further comprising a means of expressing RecE and RecT proteins or Redα and Redß proteins.

61. The reagent kit according to any one of claims 58-60, wherein the means comprises a vector system capable of expressing the recE and recT genes in the host cell.

62. The reagent kit according to any one of claims 58-61, further comprising a phenotypic marker located in the recipient cloning vehicle between the third and fourth regions of sequence homology.

63. The reagent kit according to any one of claims 58-62, wherein the recipient cloning vehicle further comprises a recognition site for a site-specific recombinase on the recipient cloning vehicle between the third and fourth regions of sequence homology.

64. The reagent kit of claim 63, further comprising means for expressing a site-specific recombinase in said host cell.